

SUPPLEMENTARY INFORMATION

Manuscript title: Lupane triterpenes from the leaves of the tropical rain forest tree *Hopea odorata* Roxb. and their cytotoxic activities.

Authors: Malai Satiraphan^{a,b}, Perayot Pamornsilpadaharm^b, Chavalit Sittisombut^b, Uthai Sotanaphun^b, Françoise Raynaud^c, Christiane Garbay^c, Sylvie Michel^a, Xavier Cachet^{a,*}

Contents (S1 to S are the page numbers)

S1 – Title, authors and description of Supplementary information content.

S2-S3 – Evaluation of the cytotoxic activities of isolated lupanes: Experimental details and summary of results.

S4-S8 – Purification and identification of isolated compounds: Experimental with full physical and spectroscopic data for compounds **1-8**, including a complete assignment of ¹H and ¹³C NMR data for lupanes **6-8**.

S9-S35 – Original spectroscopic data:

S9 – MS (ESI-), ¹H, ¹³C and HSQC edit spectra of compound **1**.

S12 – MS (ESI-), ¹H and ¹³C spectra of compound **2**.

S14 – MS (ESI-), ¹H and ¹³C spectra of compound **3**.

S16 – MS (ESI+), ¹H and ¹³C spectra of compound **4**.

S18 – MS (ESI+), ¹H and ¹³C spectra of compound **5**.

S21 – MS (ESI-), ¹H, ¹³C, HSQC edit, HMBC and NOESY spectra of compound **6**.

S25 – MS (ESI+), ¹H, ¹³C, HSQC edit and NOESY spectra of compound **7**.

S28 – MS (ESI-), ¹H, ¹³C and HSQC edit spectra of compound **8**.

S30 – MS (EI), ¹H, ¹³C and HSQC edit spectra of compound **9**.

S34 – MS (EI), ¹H and ¹³C spectra of compound **10**.

Evaluation of the cytotoxic activities of isolated lupanes: Experimental details and summary of results.

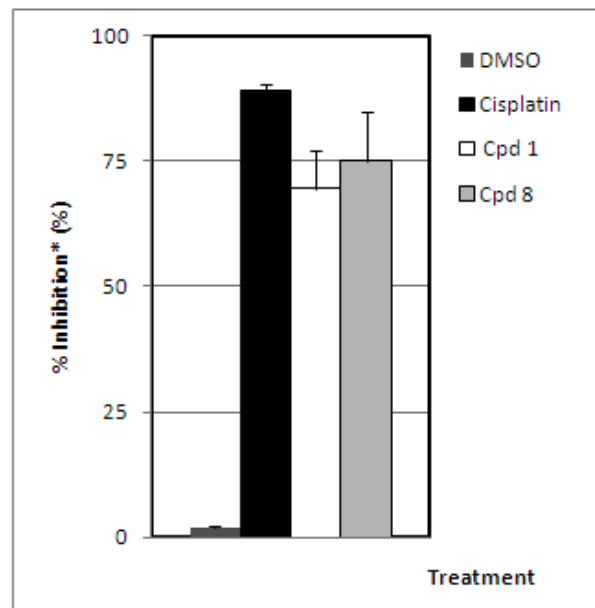
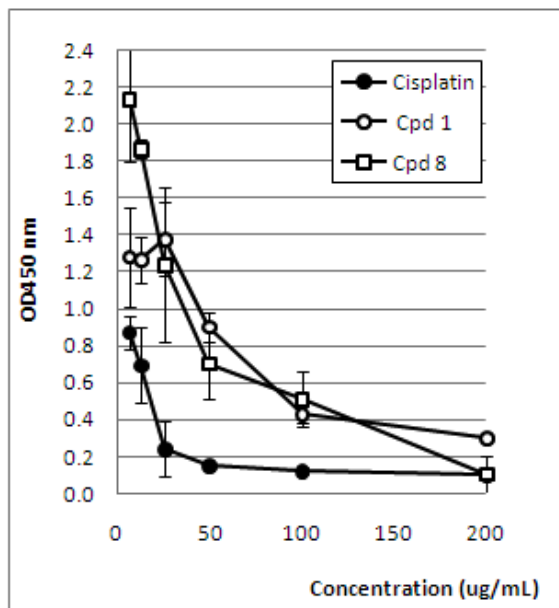
Materials. Dulbecco's Modified Eagle's Medium (DMEM), fetal bovine serum (FBS), penicillin, streptomycin, trypsin/EDTA were purchased from Invitrogen (Carlsbad, CA, USA), WST-1 solution were purchased from Roche Applied Science (Meylan Cedex, France).

In vitro cytotoxic assay. The cytotoxic effects of isolated lupanes on various human cancer cell lines were evaluated using WST-1 method, as previously described (Ishiyama et al., 1996; Tsai et al., 2011). Different human cancer cell lines (*i.e.* prostate cancer cell line (PC3), human breast adenocarcinoma cell line (MDA-MB-231), colorectal adenocarcinoma cell line (HT-29) and colorectal carcinoma cell line (HCT 116)), were selected and seeded onto 96-well plates at a cell density of 5,000 cells/well in 5% CO₂ incubator at 37°C. After 24 h incubation, cells were exposed to various concentrations of each compound **1** to **8** (3 wells for each dilution) for 24 hr. Then, the tetrazolium salt WST-1 solution was added and cultured for further 4 h. To determine the cell survival, Optical Density (OD) was measured with a Bio-Rad Coda microplate analyzer at a wavelength of 450 nm (reference wavelength: 600 nm). Cisplatin was used as a positive control. Results presented in Table 2 were expressed as IC₅₀, the concentration required for 50% inhibition cell growth of treated cells compared to untreated controls. Dose-response curves of the active lupane triterpenes against human cancer cell line compared to cisplatin were shown in Figure 4.

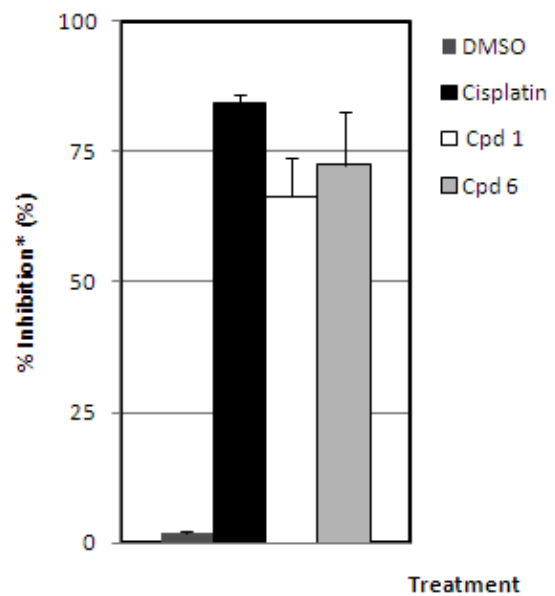
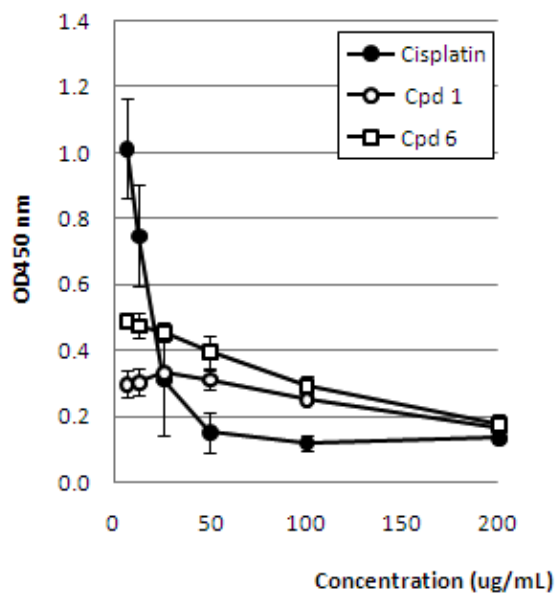
Statistical analysis: IC₅₀ are shown as mean ± standard deviation (SD) of triplicates from each independent experiment. Cell proliferation from WST-1 activities were analyzed using the student's t-test. P-values less than 0.05 were considered statistically significant.

Fig. 4 Dose-response curve and % growth inhibition at 200 µg/mL of HCT116 cells to a 24-hour treatment with compounds **1** and **8** or cisplatin (positive control). (A); Dose-response curve and % growth inhibition at 200 µg/mL of PC3 cells to a 24-hour treatment with compounds **1** and **6** or cisplatin (positive control). (B)

A) HCT116 cell line



B) PC3 cell line



Purification and identification of isolated compounds: Experimental with full physical and spectroscopic data for compounds **1-8**, including a complete assignment of ^1H and ^{13}C NMR data for lupanes **6-8**.

General methods

1D and 2D NMR spectra were recorded on Bruker AC300 (300 MHz) and Avance 400 (400 MHz). Chemical shifts are given in parts per million (ppm, δ) relative to solvent peaks as internal standards (δ : CDCl_3 : 7.27 ppm (^1H), 77.0 ppm (^{13}C)); coupling constants are given in hertz (Hz, J). IR spectra were recorded using Nicolet 510 FT-IR spectrophotometer as film on NaCl pellets. Optical rotations were measured on a Perkin-Elmer Model 341 polarimeter at 20°C. Mass spectra were determined on Thermo Finnigan LCQ Advantage (ESI-ion trap) for low resolution MS and LCT Premier Waters® (ESI-TOF) for high resolution measurement. GC/MS analyses for sesquiterpenes and fatty acid were carried out with a GC chromatograph Hewlett Pack 6890 GC coupled to a 5975 quadrupole MS. Rough fractionation was performed on MPLC (medium pressure liquid chromatography) BUCHI684 Fraction collector. Further separation was carried out on silica gel column chromatography (Silica gel 60A C.C. 20-45 μm chromagel SDS-CARLOERBA).

Betulonic acid (1) $\text{C}_{30}\text{H}_{46}\text{O}_3$; MW 454; white solid; $[\alpha]_{\text{D}}^{20} +12.2$ (CHCl_3 , $c=0.09$); IR (film) ν_{max} (cm^{-1}) 3500-2500 (br), 3066, 2917, 2849, 2863, 1703, 1694, 1462, 1377 and 757; ^1H and ^{13}C NMR (CDCl_3) data see Tables 1 and 2; MS (ESI-) m/z 453 $[\text{M}-\text{H}]^-$.

Betulonic acid (2) $\text{C}_{30}\text{H}_{48}\text{O}_3$; MW 456; colorless crystals; $[\alpha]_{\text{D}}^{20} +2.7^\circ$ (CHCl_3 , $c=0.075$); IR (film) ν_{max} (cm^{-1}) 3466-2500 (br), 3069, 2926, 2851 and 1687; ^1H NMR (δ_{H} , ppm, in CDCl_3) 0.70 (H5), 0.78 (H24), 0.82 (H25), 0.92 (H26), 0.96 (H23), 0.98 (H27), 1.71 (H30), 3.04 (H19), 3.21 (H3), 4.62, 4.75 (H29); ^{13}C NMR (δ_{C} , ppm, in CDCl_3) 38.7 (C1), 27.4 (C2), 79.0 (C-3), 38.8 (C4), 55.3 (C5), 18.3 (C6), 34.3 (C7), 40.7 (C8), 50.5 (C9), 37.2 (C10), 20.8 (C11), 25.5 (C12), 38.4 (C13), 42.4 (C14), 30.5 (C15), 32.1 (C16), 56.3 (C17), 49.2 (C18), 46.9 (C19), 150.4 (C20), 29.7 (C21), 37.1 (C22), 28.0

(C23), 15.4 (C24), 16.1 (C25), 16.1 (C26), 14.7 (C27), 180.5 (C28), 109.7 (C29), 19.4 (C30); MS (ES-) m/z 455 [M-H]⁻.

Epibetulinic acid (3) C₃₀H₄₈O₃; MW 456; white solid; $[\alpha]_D^{20}$ -3.3 (CHCl₃, c=0.09); IR (film) ν_{\max} (cm⁻¹) 3700-2500 (br), 3428, 3069, 2914, 2844, 1700, 1459, 1295 and 762; ¹H NMR (δ_H , ppm, in CDCl₃) 0.82 (H24), 0.84 (H25), 0.94 (H23+H26), 1.00 (H27), 1.70 (H30), 3.02 (H19), 3.40 (H3), 4.62, 4.75 (H29); ¹³C NMR (δ_C , ppm, in CDCl₃) 33.2 (C1), 25.4 (C2), 76.3 (C-3), 37.5 (C4), 49.0 (C5), 18.2 (C6), 34.2 (C7), 40.9 (C8), 50.3 (C9), 37.3 (C10), 20.7 (C11), 25.5 (C12), 38.4 (C13), 42.5 (C14), 30.6 (C15), 32.2 (C16), 56.4 (C17), 49.2 (C18), 46.9 (C19), 150.5 (C20), 29.6 (C21), 37.1 (C22), 28.3 (C23), 22.1 (C24), 15.9 (C25), 16.0 (C26), 14.8 (C27), 181.2 (C28), 109.7 (C29), 19.4 (C30); MS (ES-) m/z 455 [M-H]⁻.

28-hydroxylup-20(29)-en-3-one (Betulone) (4) C₃₀H₄₈O₂; MW 440; white solid; $[\alpha]_D^{20}$ +21.7 (CHCl₃, c=0.12); IR (film) ν_{\max} (cm⁻¹) 3435, 3065, 2917, 2847, 1705, 1463, 1376, 1025 and 757; ¹H NMR (δ_H , ppm, in CDCl₃) 0.93 (H25), 1.00 (H27), 1.03 (H24), 1.07 (H26), 1.08 (H23), 1.70 (H30), 3.36, 3.81 (H28), 4.59, 4.69 (H29); ¹³C NMR (δ_C , ppm, in CDCl₃) 39.6 (C1), 34.2 (C2), 218.3 (C3), 47.4 (C4), 54.9 (C5), 19.7 (C6), 33.5 (C7), 40.9 (C8), 49.7 (C9), 36.9 (C10), 21.4 (C11), 25.2 (C12), 37.4 (C13), 42.8 (C14), 27.0 (C15), 29.3 (C16), 47.8 (C17), 48.7 (C18), 47.8 (C19), 150.4 (C20), 29.6 (C21), 34.0 (C22), 26.6 (C23), 21.1 (C24), 15.8 (C25), 16.0 (C26), 14.7 (C27), 60.5 (C28), 109.8 (C29), 19.1 (C30); MS (ES+) m/z 463 [M+Na]⁺.

30-hydroxylup-20(29)-en-3-one (5) C₃₀H₄₈O₂; MW 440; white powder; $[\alpha]_D^{20}$ +14.4 (CHCl₃, c=0.18); IR (film) ν_{\max} (cm⁻¹) 3448, 3085, 2937, 2851 and 1705; ¹H and ¹³C NMR (CDCl₃) data see Tables 1 and 2; MS (ES+) m/z 463 [M+Na]⁺.

3,30-dioxo-lup-20,29-en-28-oic acid, (6) C₃₀H₄₄O₄; MW 468.3240; colorless crystals; $[\alpha]_D^{20}$ +30.6 (CHCl₃, c=0.11); IR (film) ν_{\max} (cm⁻¹) 3700-2500 (br), 3069, 2924, 2845, 1731, 1685, 1463 and 1276; ¹H and ¹³C NMR (CDCl₃) data see Tables 1 and 2; MS (ESI-) m/z 467 [M-H]⁻. HRMS (ESI-TOF) m/z (ESI-) : 467.3146 [M-H]⁻ (calcd for C₃₀H₄₃O₄ 467.3161)

28,30-dihydroxy-3-oxolup-20(29)-ene, (7) C₃₀H₄₈O₃; MW 456; green oil; $[\alpha]_D^{20} +3.0$ (CHCl₃, c=0.33); IR (film) ν_{\max} (cm⁻¹) 3434, 1699 and 1456; ¹H and ¹³C NMR (CDCl₃) data see Tables 1 and 2; MS (ESI+) m/z 479 [M+Na]⁺.

Messagenic acid G, (8) C₃₀H₄₆O₄; MW 470; amorphous solid; $[\alpha]_D^{20} +14.3$ (CHCl₃, c=0.14); IR (film) ν_{\max} (cm⁻¹) 3700-2500 (br), 3074, 2938, 2864 and 1695; ¹H and ¹³C NMR (CDCl₃) data see Tables 1 and 2; MS (ESI-) m/z 469 [M-H]⁺.

Table 1. ^1H NMR data (400 MHz) of **1**, **5**, **6**, **7**, **8** (in CDCl_3 , J in Hz, δ in ppm)

Position	1	5	6	7	8
	δ_{H}	δ_{H}	δ_{H}	δ_{H}	δ_{H}
1	1.38 (m), 1H 1.91 (m), 1H	1.38 (m), 1H 1.90 (m), 1H	1.37 (m), 1H 1.89 (m), 1H	1.38 (m), 1H 1.88 (m), 1H	1.39 (m), 1H 1.90 (m), 1H
2	2.42 (m), 1H 2.50 (m), 1H	2.43 (m), 1H 2.50 (m), 1H	2.41 (m), 1H 2.47 (m), 1H	2.42 (m), 1H 2.49 (m), 1H	2.43 (m), 1H 2.50 (m), 1H
3					
4					
5	1.35 (m), 1H	1.32 (m), 1H	1.31 (m), 1H	1.33 (m), 1H	1.33 (m), 1H
6	1.52 (m), 2H	1.47 (m), 2H	1.46 (m), 2H	1.47 (m), 2H	1.47 (m), 2H
7	1.44 (m), 2H	1.45 (m), 2H	1.43 (m), 2H	1.45 (m), 1H	1.43 (m), 2H
8					
9	1.38 (s), 1H	1.38 (m), 1H	1.35 (m), 1H	1.38 (m), 1H	1.38 (m), 1H
10					
11	1.34 (m), 1H 1.44 (m), 1H	1.28 (m), 1H 1.42 (m), 1H	1.31 (m), 1H 1.38 (m), 1H	1.27 (m), 1H 1.43 (m), 1H	1.32 (m), 1H 1.45 (m), 1H
12	1.06 (m), 1H 1.73 (m), 1H	1.13 (m), 1H 1.36 (m), 1H	0.91 (m), 1H 1.34 (m), 1H	1.02 (m), 1H 1.41 (m), 1H	1.48 (m), 1H
13	2.23 (m), 1H	1.68 (m), 1H	2.22 (m), 1H	1.67 (m), 1H	2.21 (m), 1H
14					
15	1.42 (m), 1H 1.99 (m), 1H	1.06 (m), 1H 1.71 (m), 1H	1.23 (m), 1H 1.55 (m), 1H	1.12 (m), 1H 1.73 (m), 1H	1.23 (m), 1H 1.54 (m), 1H
16	1.44 (m), 1H 2.28 (m), 1H	1.40 (m), 1H 1.52 (m), 1H	1.51 (m), 1H 2.32 (m), 1H	1.13 (m), 1H 1.90 (m), 1H	1.46 (m), 1H 2.31 (m), 1H
17					
18	1.64 (m), 1H	1.47 (m), 1H	2.02 (m), 1H	1.72 (m), 1H	1.77 (t, $J=11.5$), 1H
19	3.02 (td, $J=10.7$; 4.8), 1H	2.34 (td, $J=5.3$, 10.8), 1H	3.35 (td, $J=11.2$; 4.6), 1H	2.31 (m), 1H	2.90 (td, $J=11.0$, 4.5), 1H
20					
21	1.22 (m), 1H 1.54 (m), 1H	1.33 (m), 1H 2.08 (m), 1H	1.42 (m), 1H 2.16 (m), 1H	1.31 (m), 1H 1.96 (m), 1H	1.42 (m), 1H 2.11 (m), 1H
22	1.47 (m), 1H 1.99 (m), 1H	1.28 (m), 1H 1.40 (m), 1H	1.75 (m), 1H 2.00 (m), 1H	1.43 (m), 1H 2.12 (m), 1H	1.57 (m), 1H 1.98 (m), 1H
23	1.08 (s), 3H	1.08 (s), 3H	1.07 (s), 3H	1.07 (s), 3H	1.08 (s), 3H
24	1.03 (s), 3H	1.03 (s), 3H	1.01 (s), 3H	1.02 (s), 3H	1.02 (s), 3H
25	0.94 (s), 3H	0.93 (s), 3H	0.91 (s), 3H	0.92 (s), 3H	0.92 (s), 3H
26	0.99 (s), 3H	1.07 (s), 3H	0.96 (s), 3H	1.06 (s), 3H	0.97 (s), 3H
27	1.00 (s), 3H	0.96 (s), 3H	0.96 (s), 3H	0.99 (s), 3H	1.02 (s), 3H
28		0.80 (s), 3H		3.33 (d, $J=10.8$), 1H 3.80 (d, $J=10.8$), 1H	
29	4.62 (s), 1H 4.75 (s), 1H	4.91 (s), 1H 4.95 (s), 1H	5.93 (s), 1H 6.31 (s), 1H	4.91 (s), 1H 4.96 (s), 1H	4.94 (s), 1H 4.99 (s), 1H
30	1.70 (s), 3H	4.15 (br d, $J=14.3$), 1H 4.11 (br d, $J=14.3$), 1H	9.53 (s), 1H	4.12 (m), 2H	4.14 (m), 2H

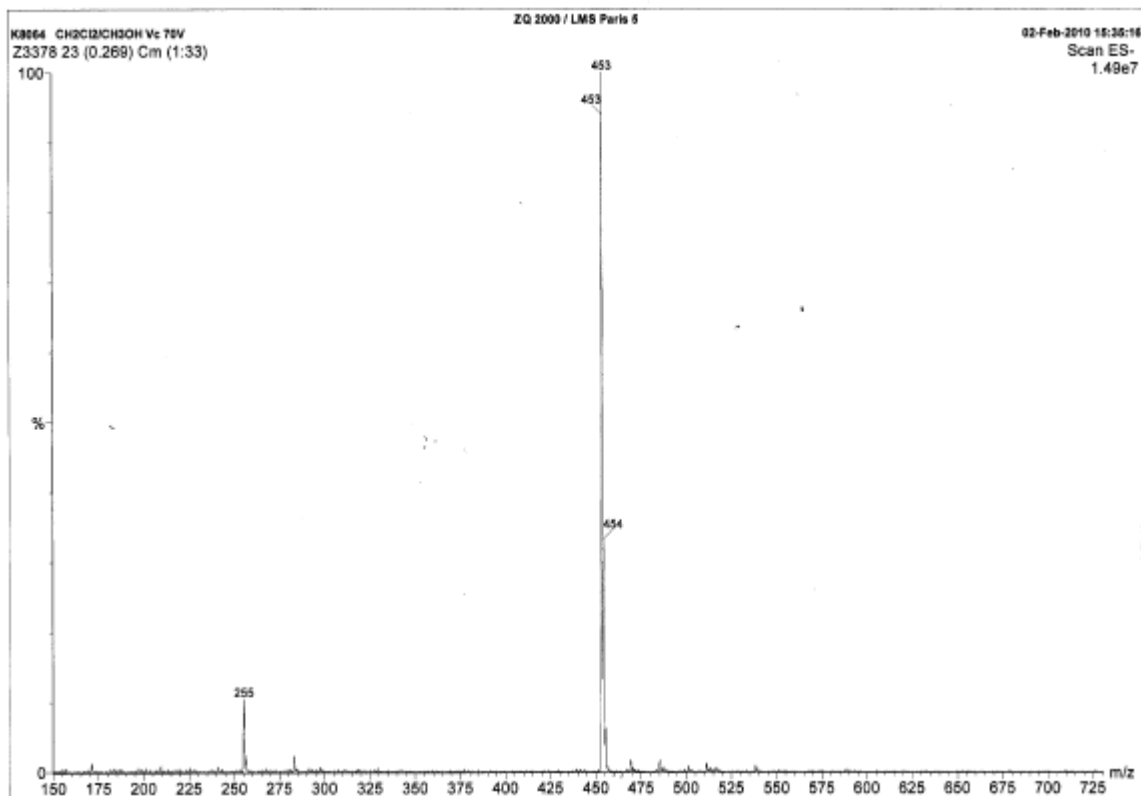
Table 2. ^{13}C NMR data (75 MHz) of **1**, **5**, **6**, **7**, **8** (in CDCl_3 , δ in ppm)

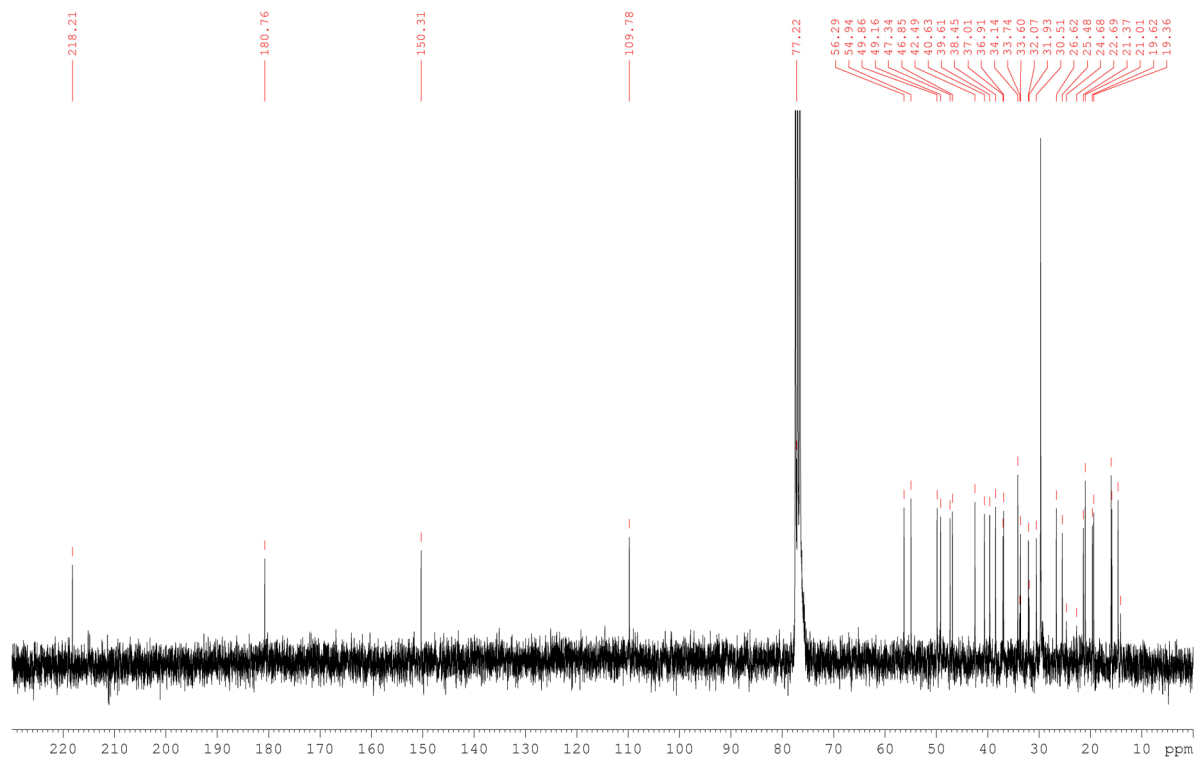
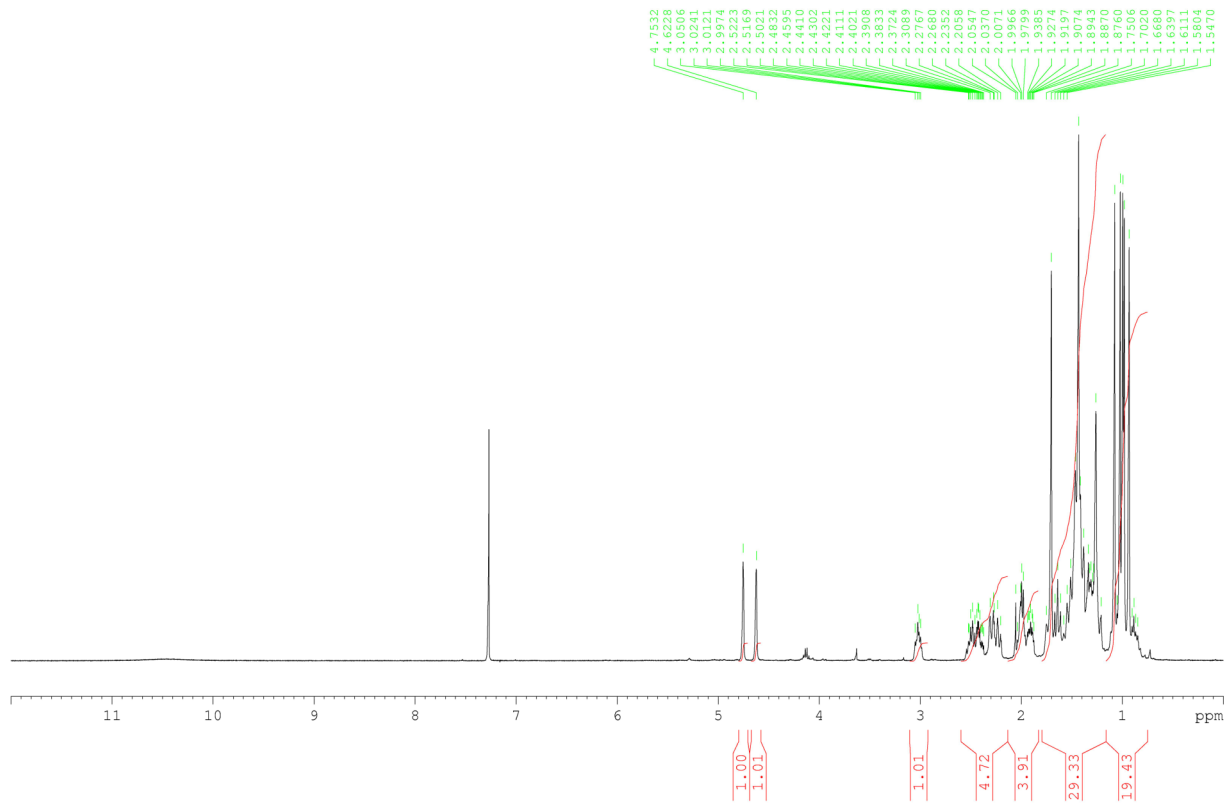
Position	1	5	6	7	8
	δ_{C}	δ_{C}	δ_{C}	δ_{C}	δ_{C}

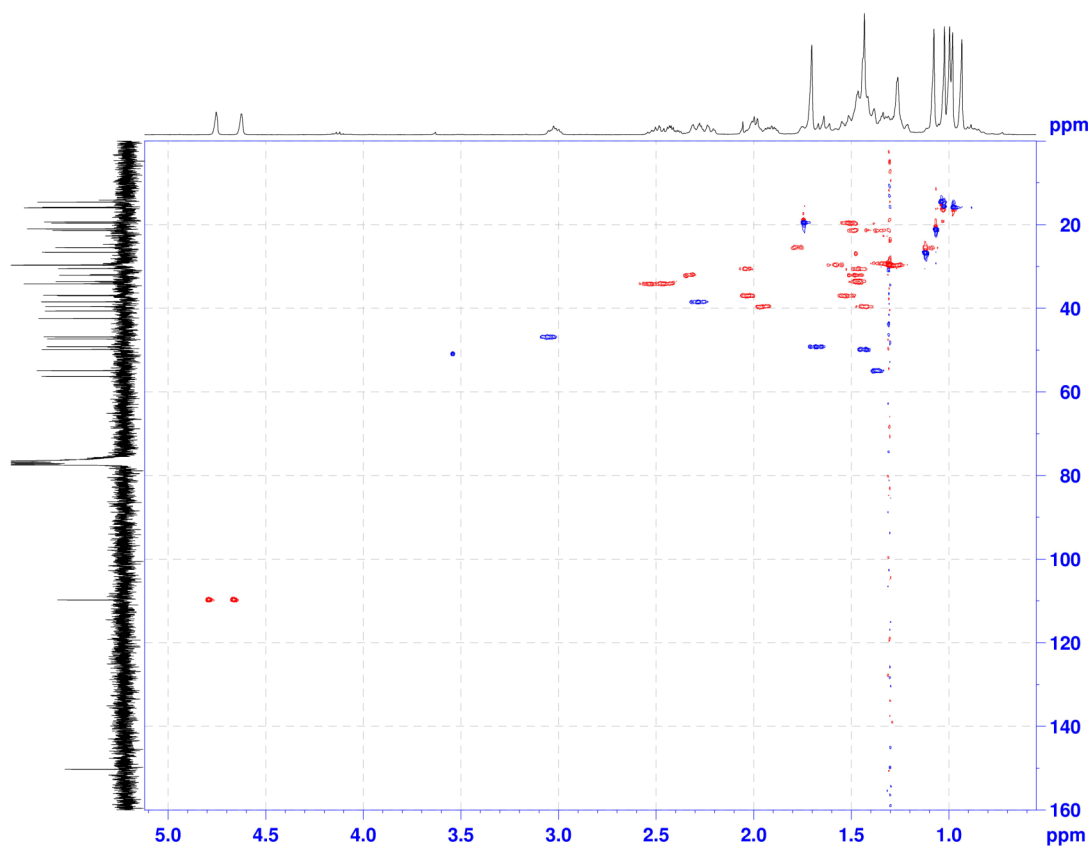
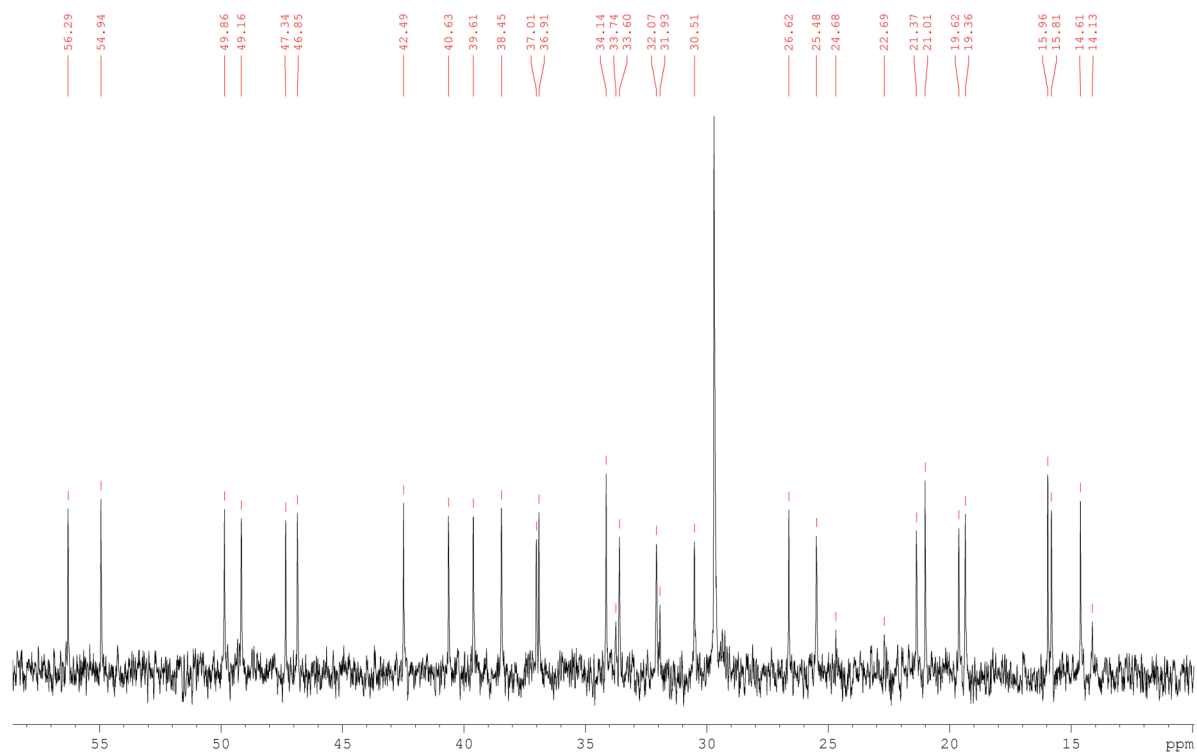
1	39.6	39.6	39.6	39.6	39.6
2	34.1	34.2	34.1	34.1	34.1
3	218.2	218.3	218.2	218.2	218.1
4	47.3	47.3	47.3	47.3	47.3
5	54.9	54.9	54.9	54.9	54.9
6	19.6	19.7	19.6	19.6	19.6
7	33.6	33.6	33.6	33.5	33.6
8	40.6	40.8	40.6	40.9	40.6
9	49.9	49.7	49.7	49.7	49.9
10	36.9	36.9	36.9	36.8	36.9
11	21.4	21.5	21.3	21.4	21.5
12	25.4	26.7	27.2	26.8	26.8
13	38.5	38.1	38.4	37.3	38.4
14	42.5	42.8	42.4	42.7	42.4
15	30.5	27.4	29.6	27.0	29.7
16	32.1	35.4	31.8	33.8	32.0
17	56.3	43.0	56.4	47.8	56.3
18	49.2	48.8	50.5	49.3	49.8
19	46.9	43.8	38.4	43.5	42.5
20	150.3	154.7	156.2	154.4	154.6
21	29.7	31.8	32.0	29.1	32.3
22	37.0	39.8	36.9	31.7	36.8
23	26.6	26.7	26.6	26.7	26.7
24	21.0	21.1	21.3	21.1	21.0
25	16.0	16.0	15.9	16.0	16.0
26	15.8	15.8	15.8	15.8	15.8
27	14.6	14.5	14.5	14.7	14.6
28	180.8	17.7	181.7	60.2	180.7
29	109.8	106.9	134.2	107.2	107.0
30	19.4	64.9	195.0	65.0	65.3

Original spectroscopic data:

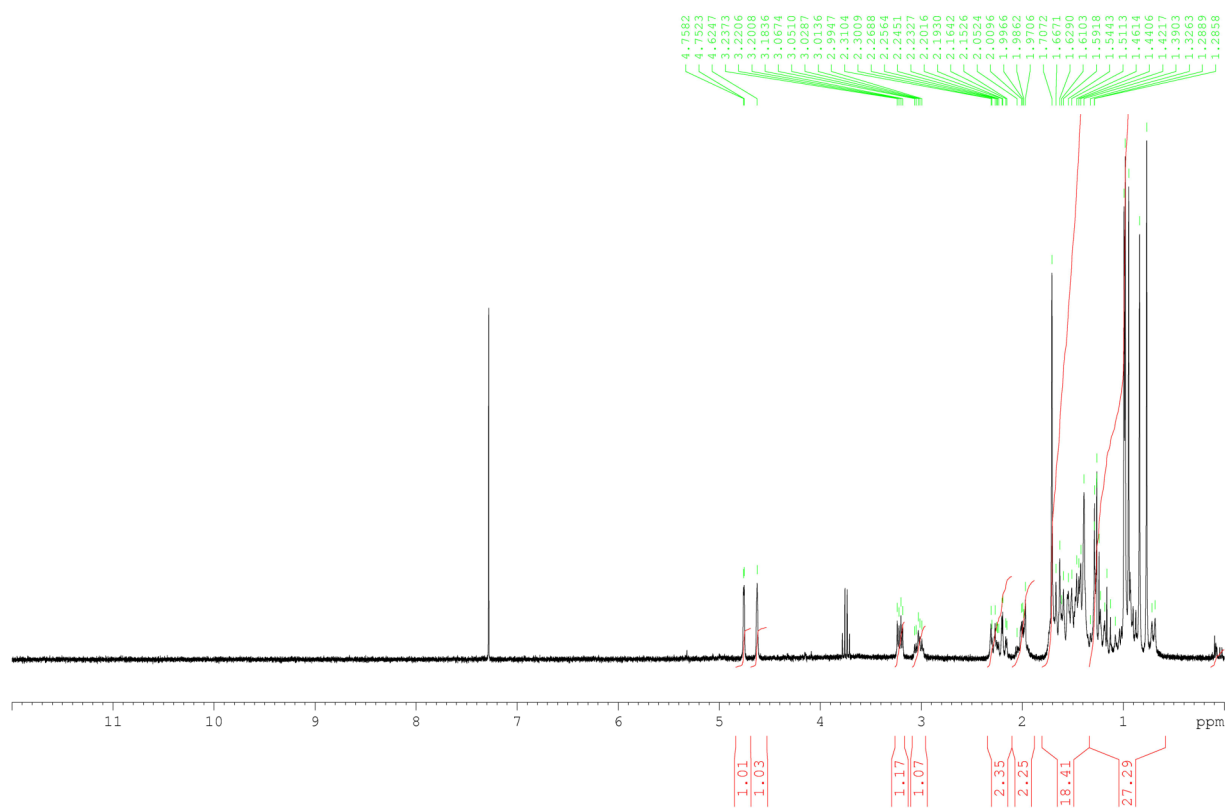
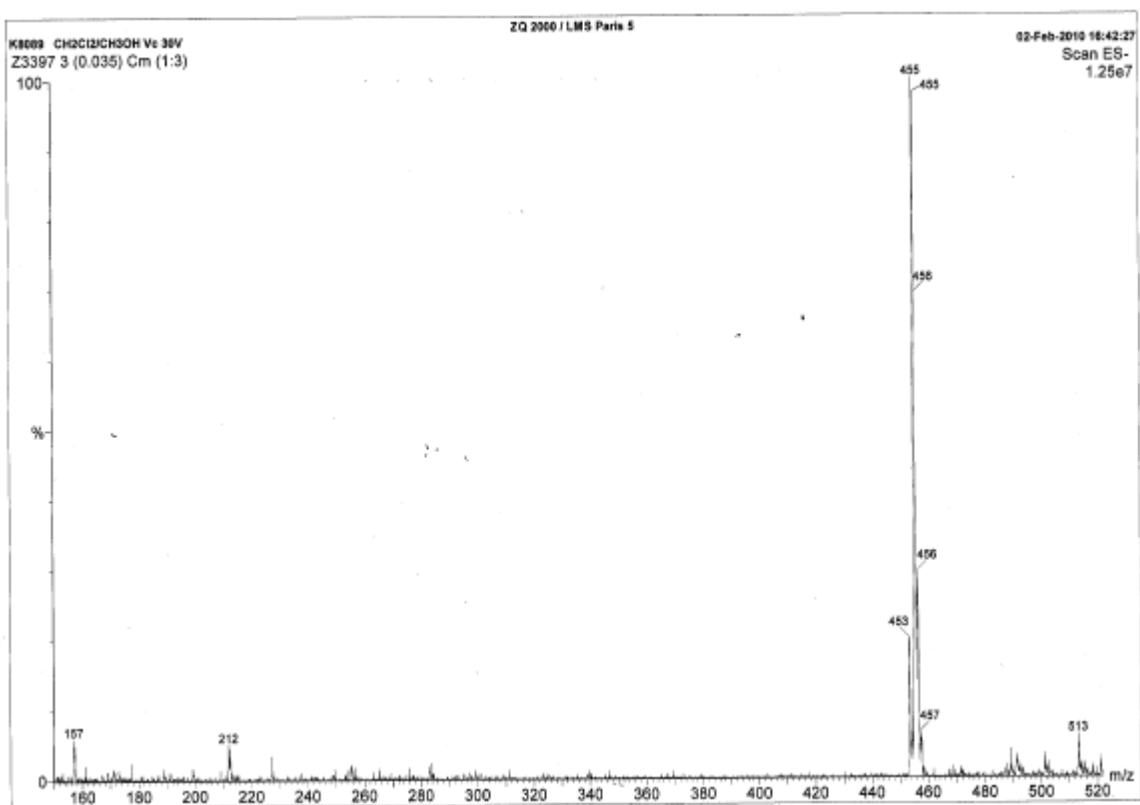
- Mass (ESI-: m/z 453 [M-H]⁻) and NMR (¹H+¹³C+HSQC edit) spectra of betulonic acid (C₃₀H₄₆O₃; MW 454) (1):

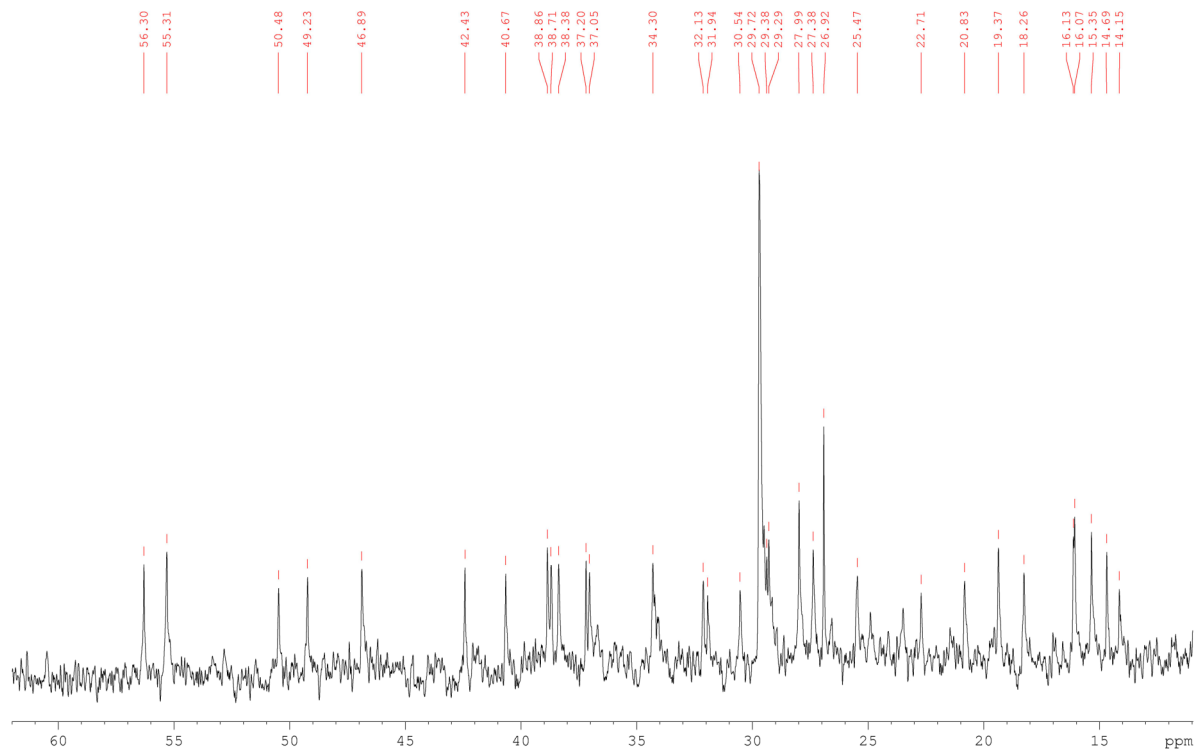
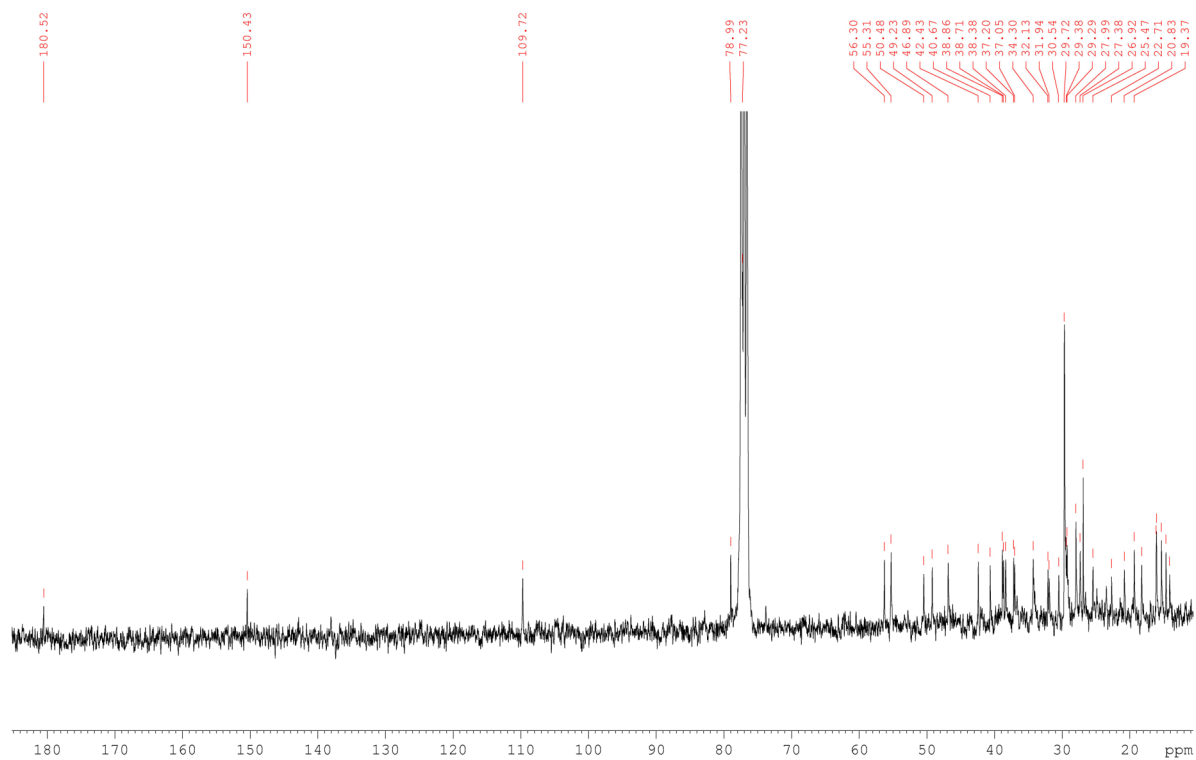




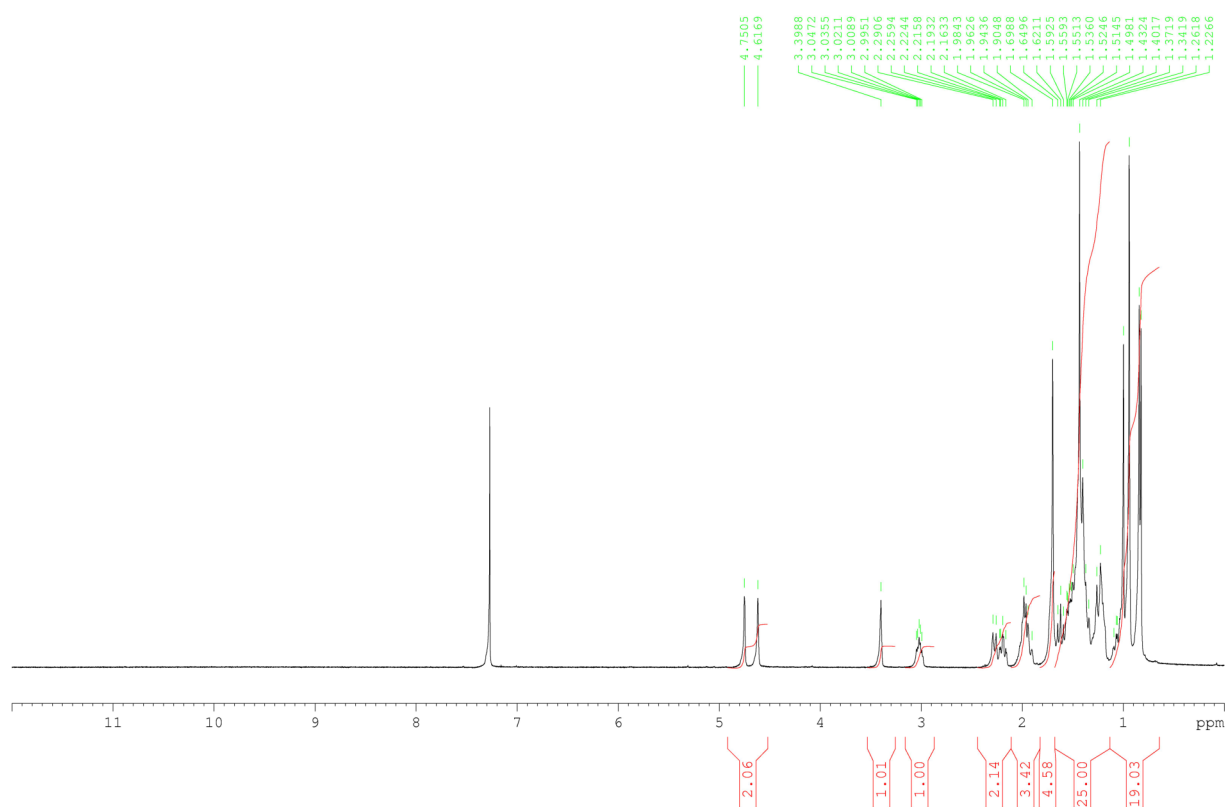
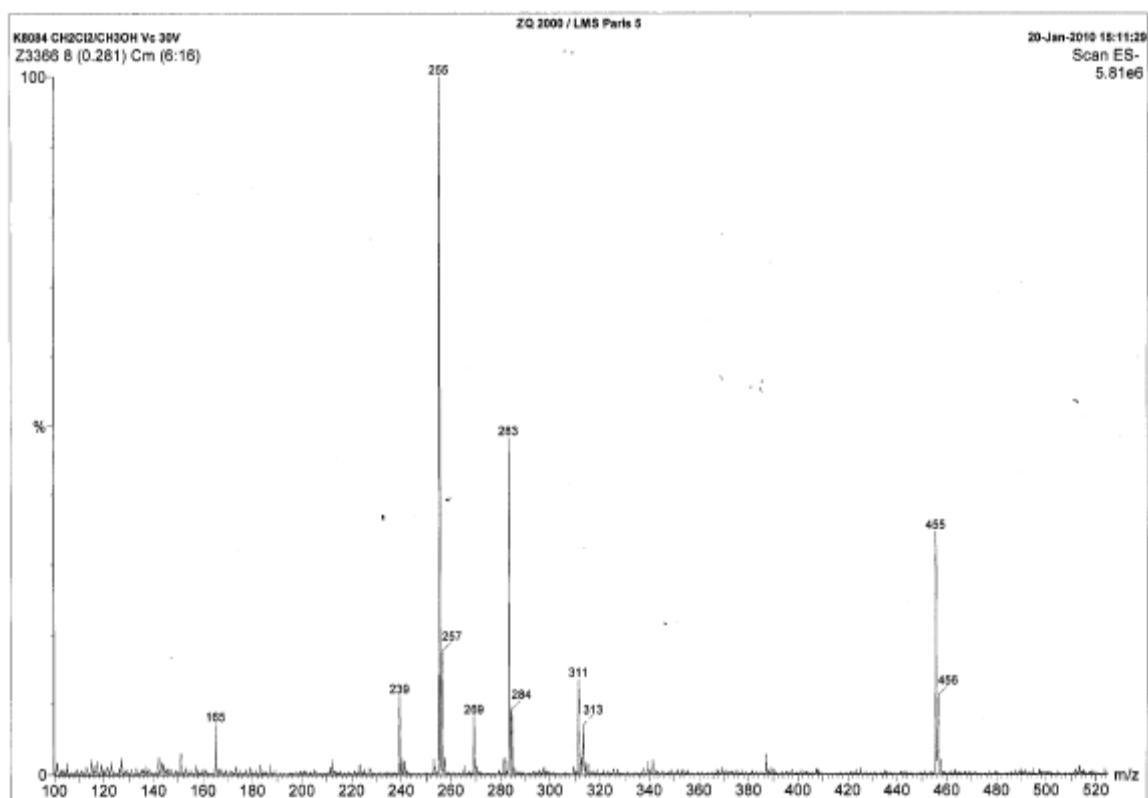


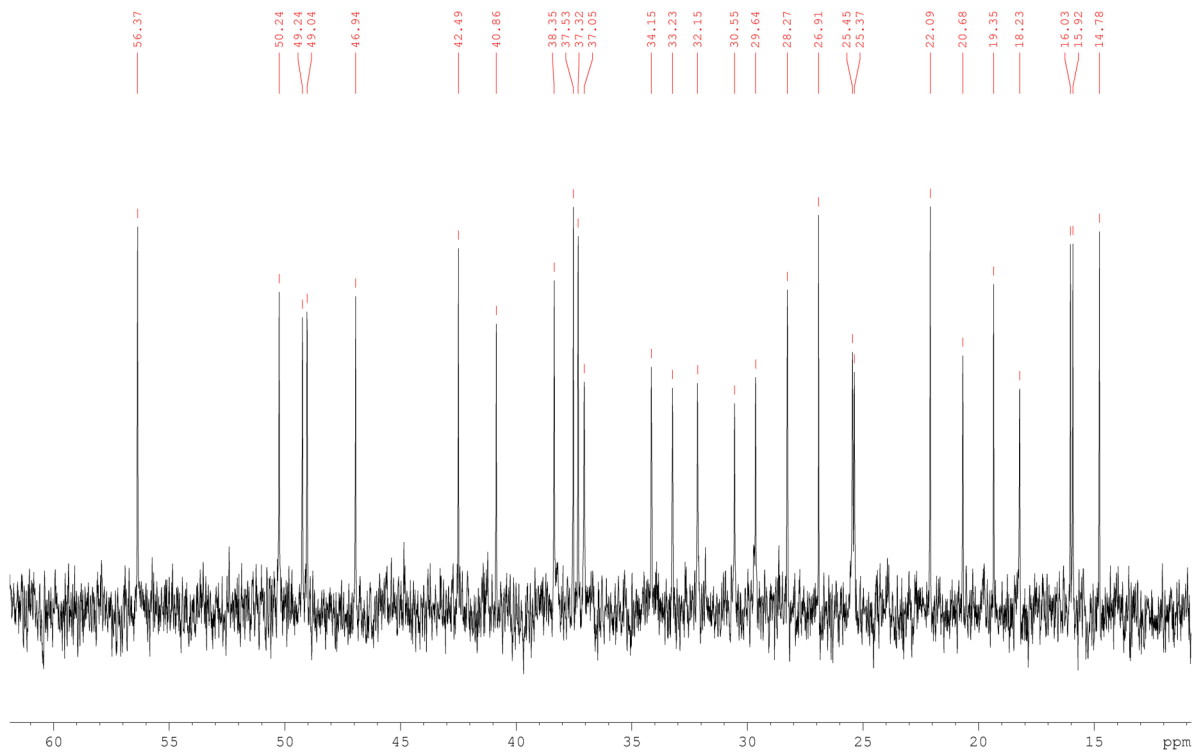
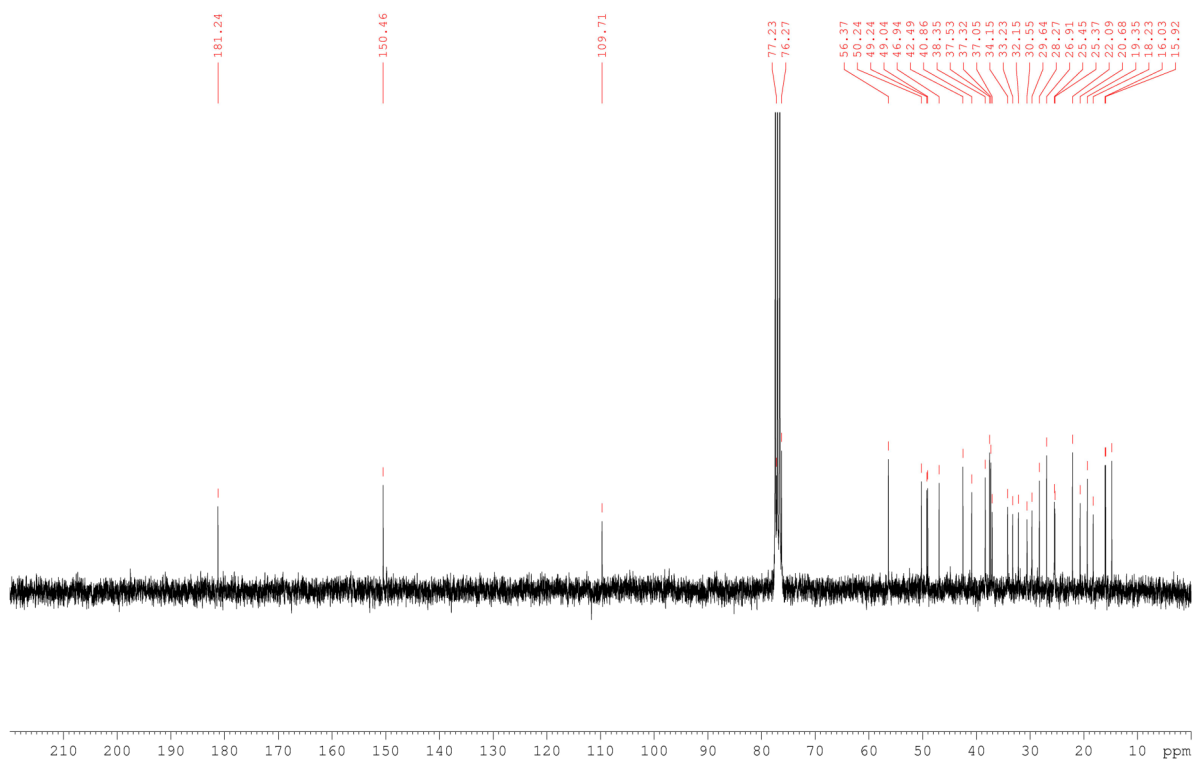
- Mass (ESI-: m/z 455 [M-H]⁻) and NMR (¹H+¹³C) spectra of betulinic acid (C₃₀H₄₈O₃; MW 456)
(2):



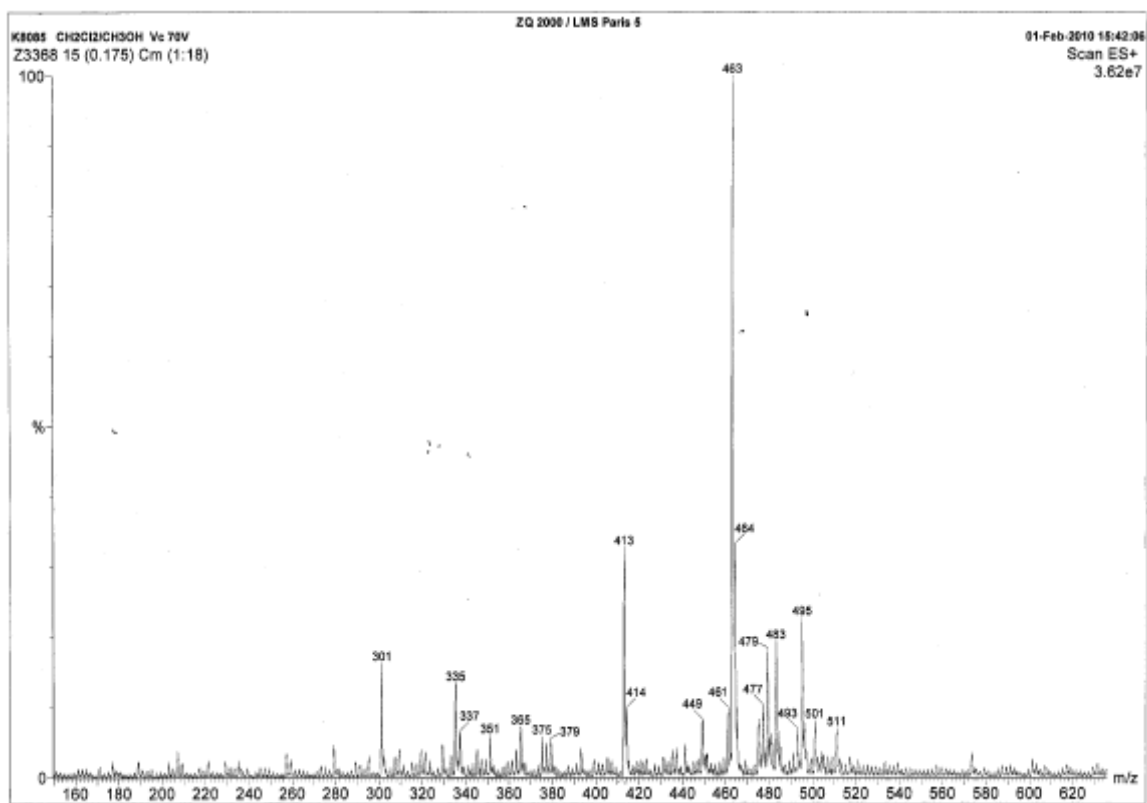


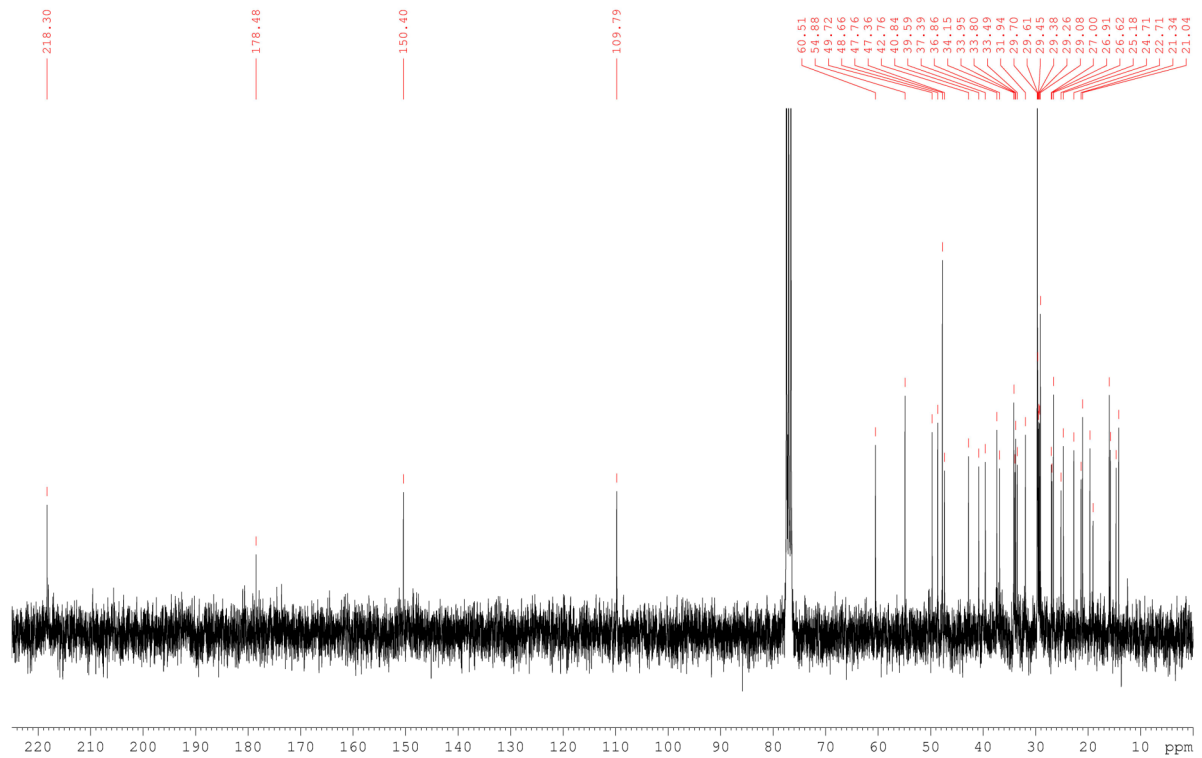
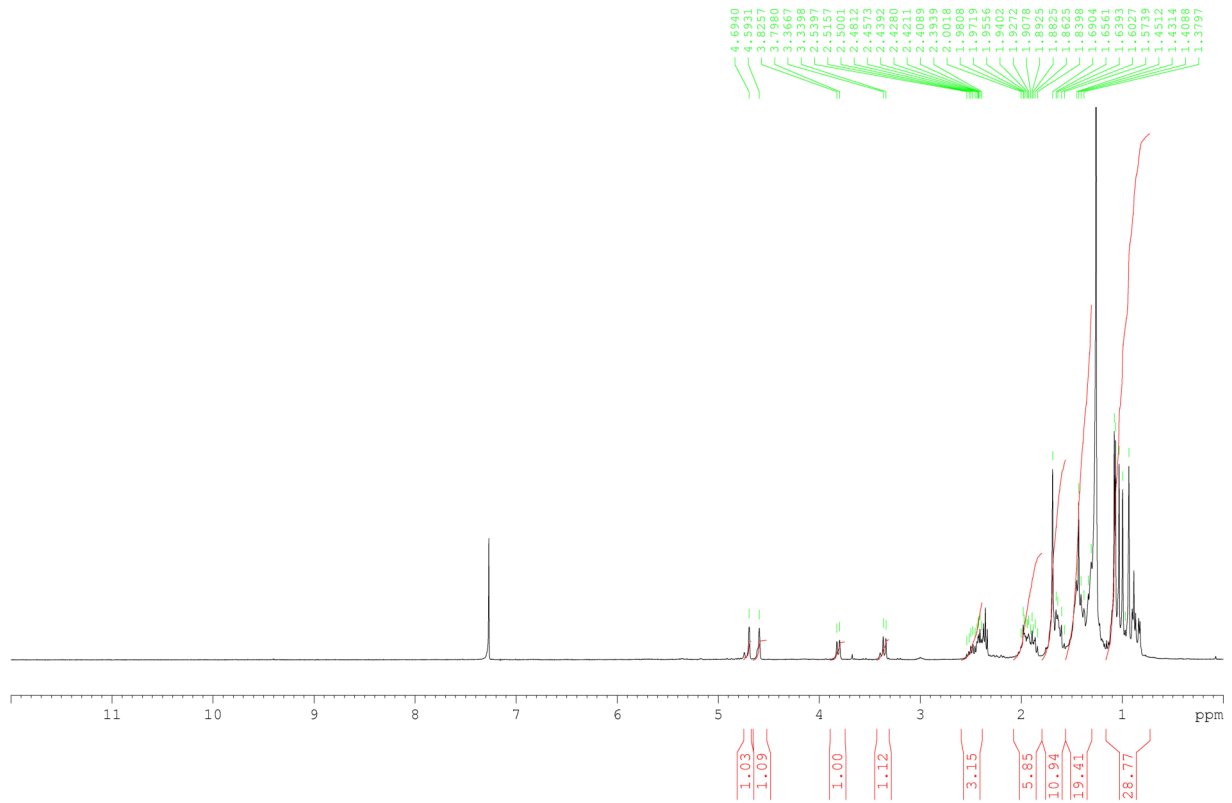
- Mass (ESI-: m/z 455 [M-H]⁻) and NMR (¹H+¹³C) spectra of epibetulinic acid (C₃₀H₄₈O₃; MW 456) (3):

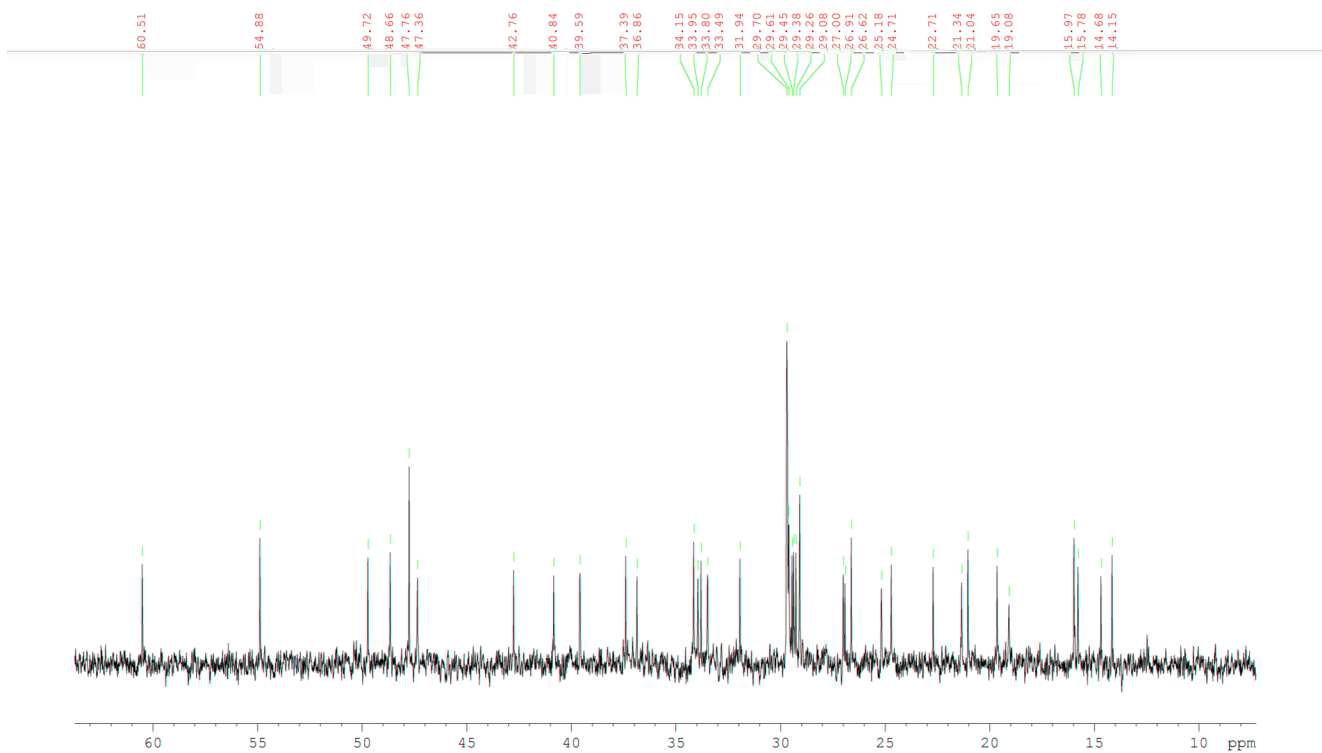




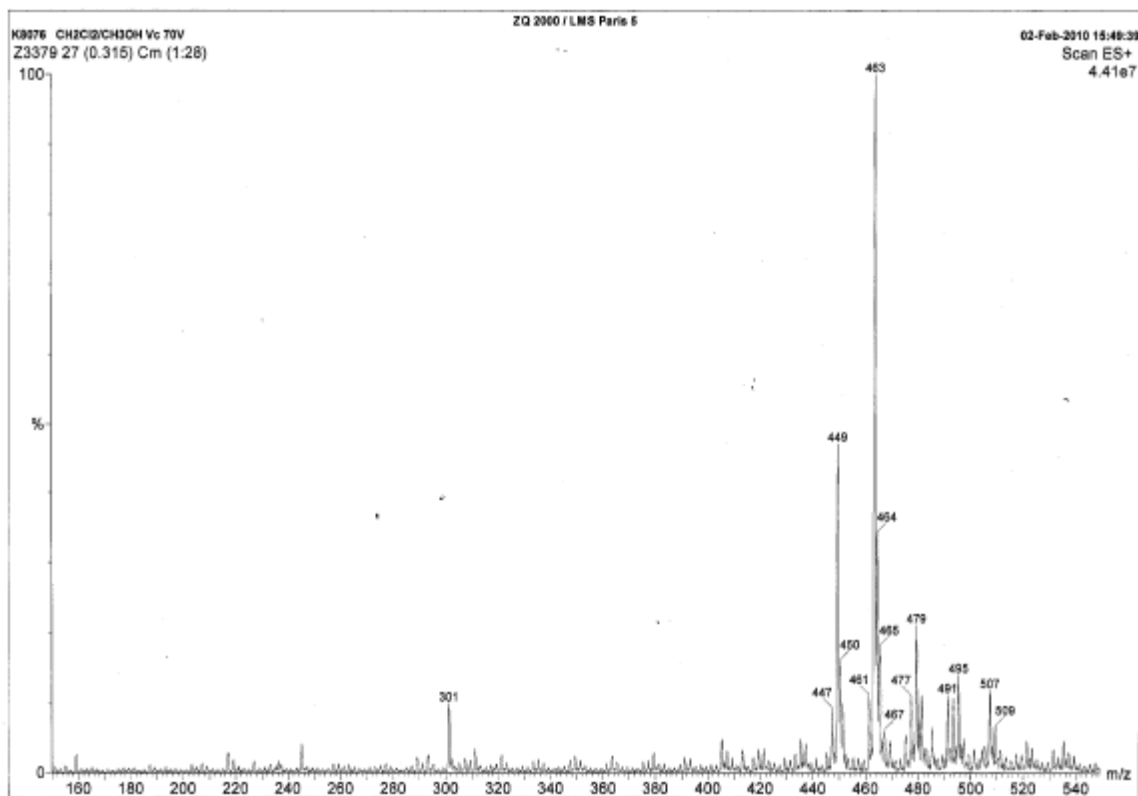
- Mass (ESI+: m/z 463 $[M+Na]^+$) and NMR (1H - ^{13}C) spectra of 28-hydroxylup-20(29)-en-3-one (= betulone) ($C_{30}H_{48}O_2$; MW 440) (4):

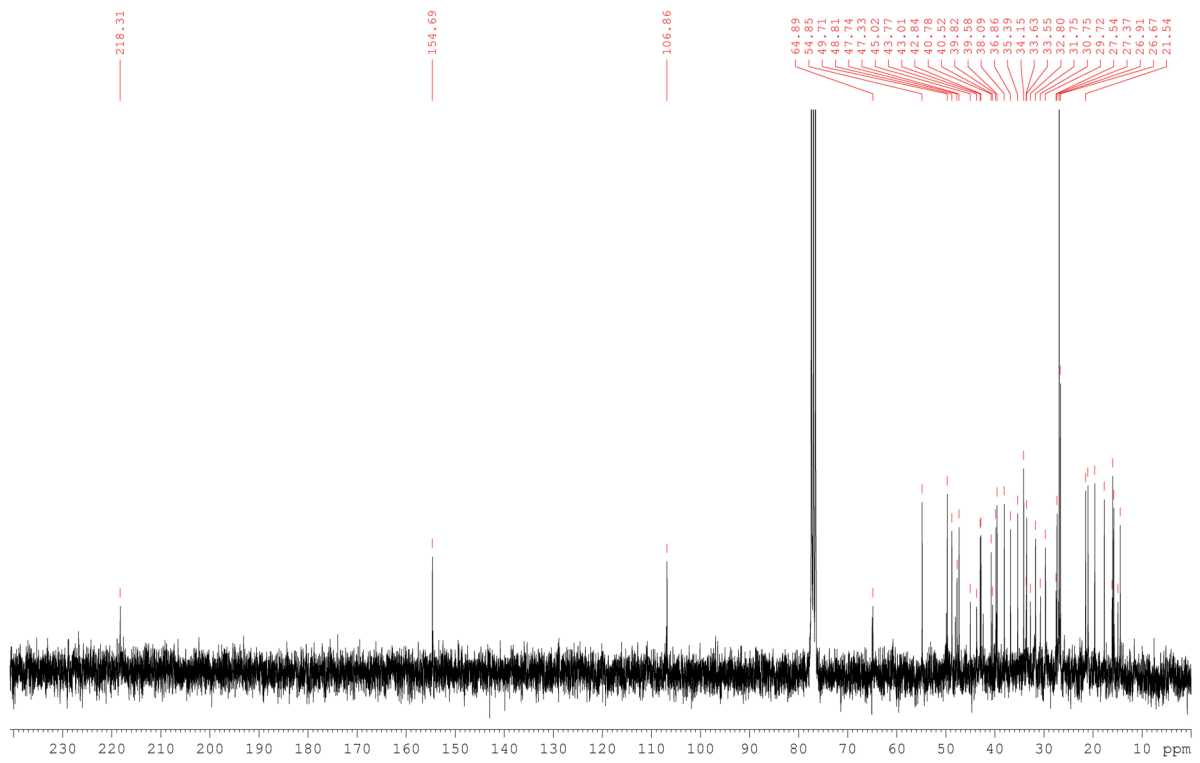
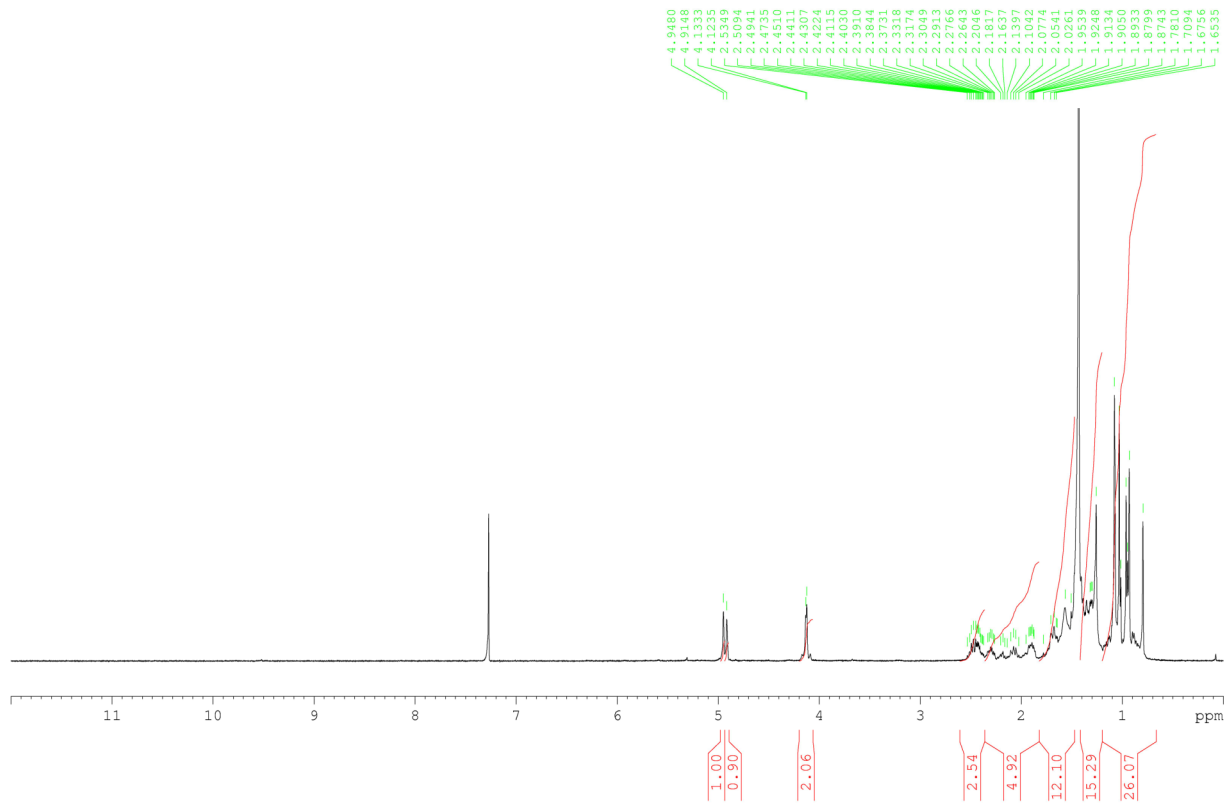


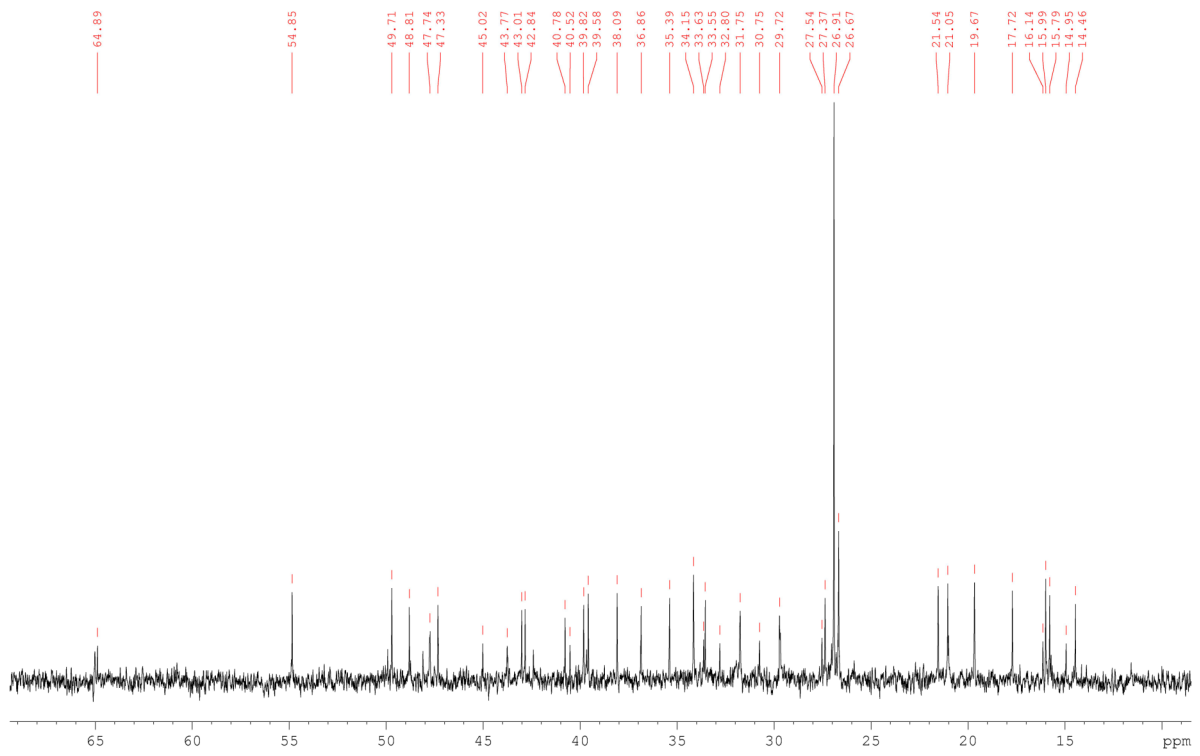




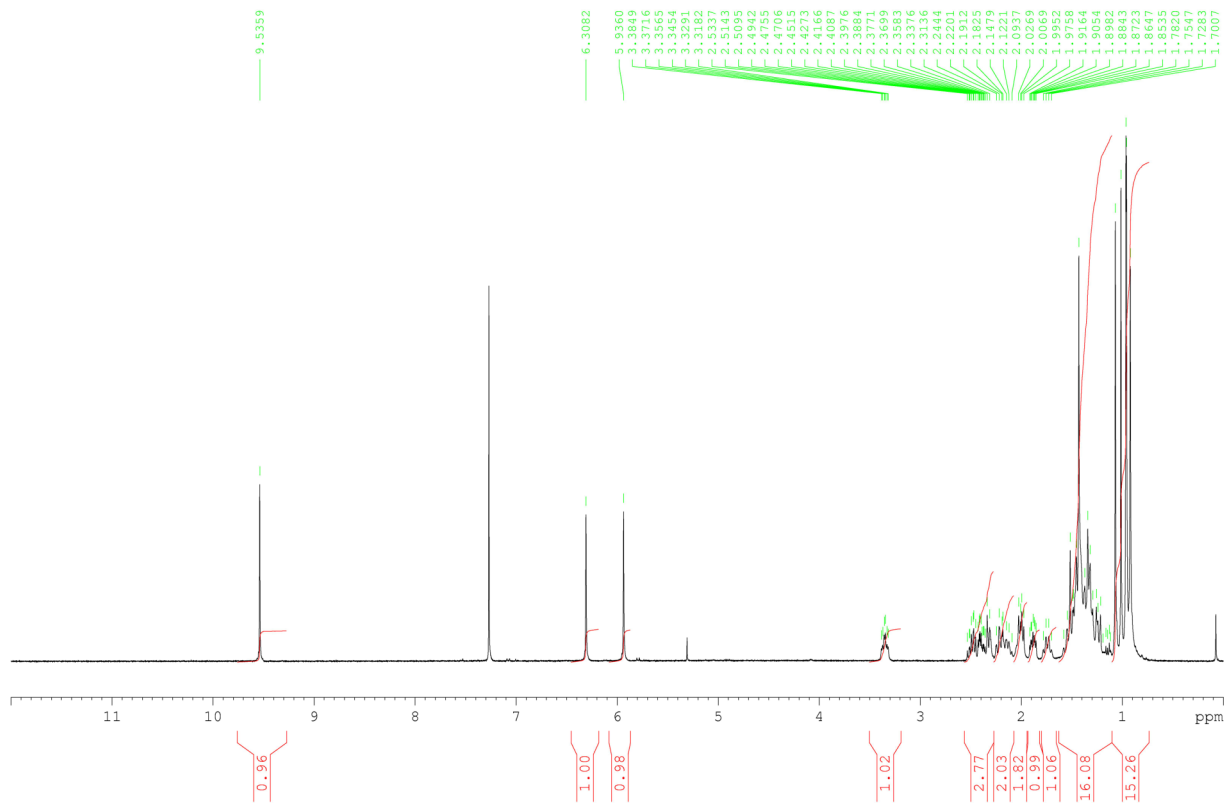
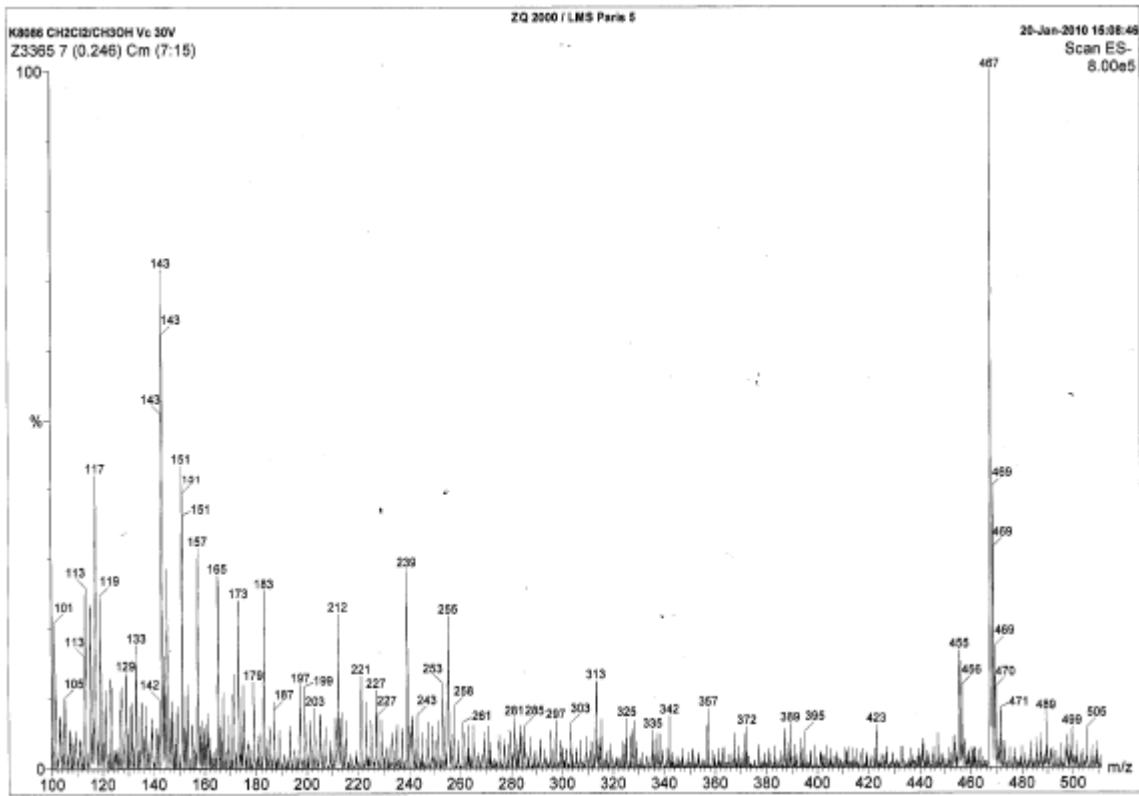
- Mass (ESI+: m/z 463 $[M+Na]^+$) and NMR ($^1H+^{13}C$) spectra of 30-hydroxy-3-lup-20(29)-ene ($C_{30}H_{48}O_2$; MW 440) (5):

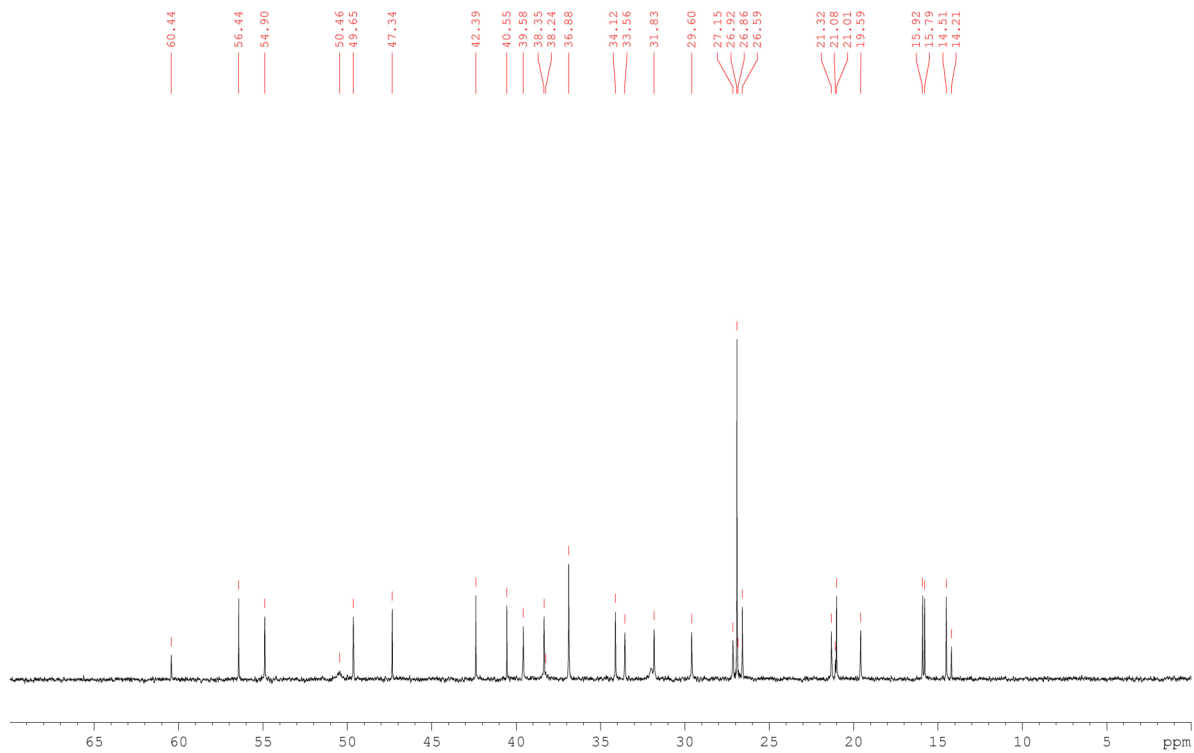
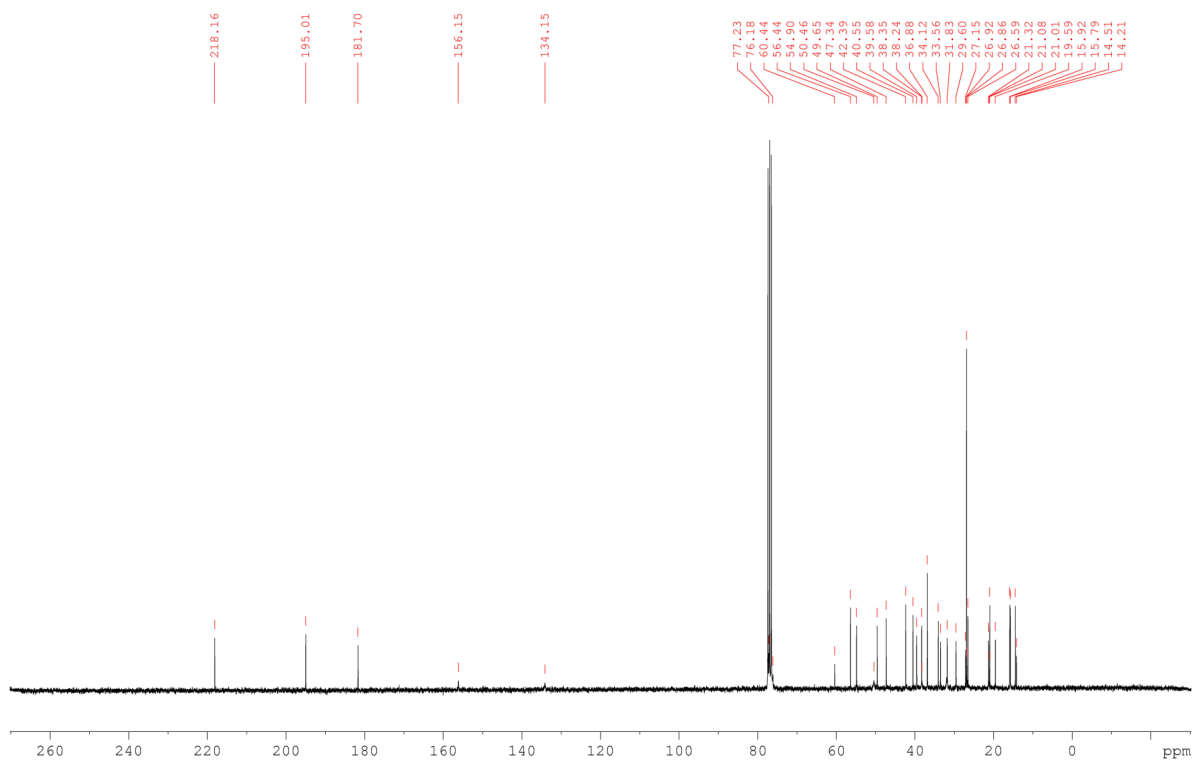


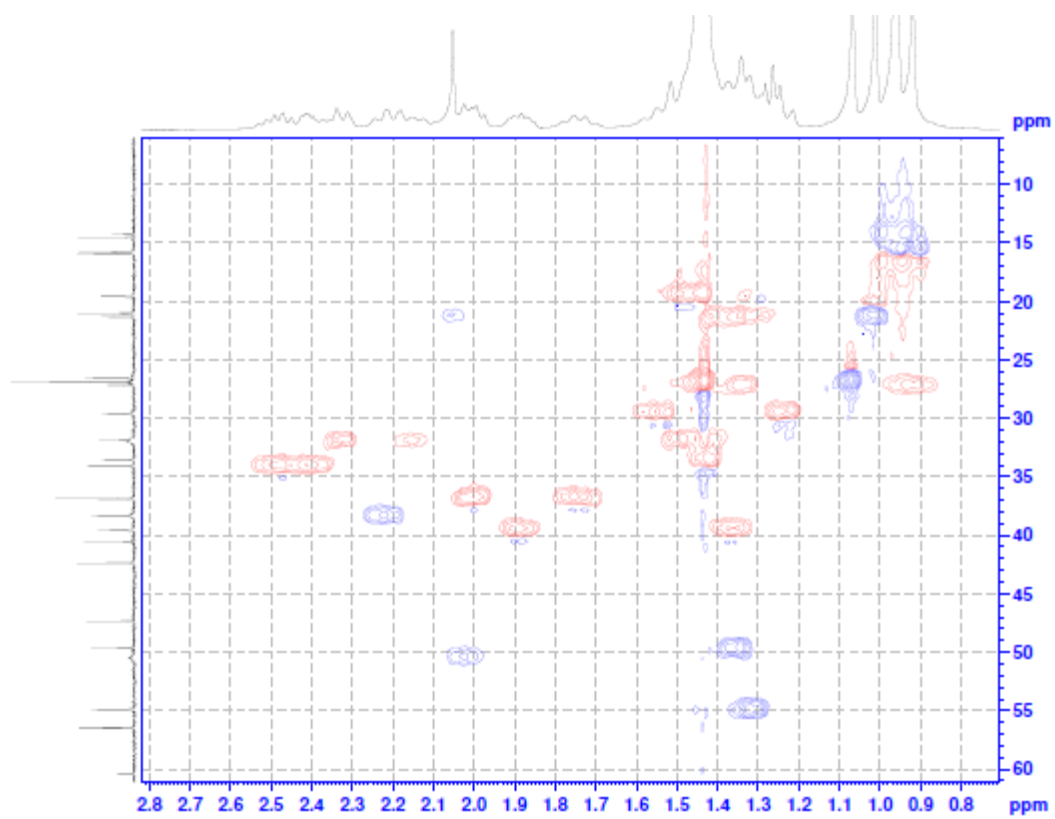
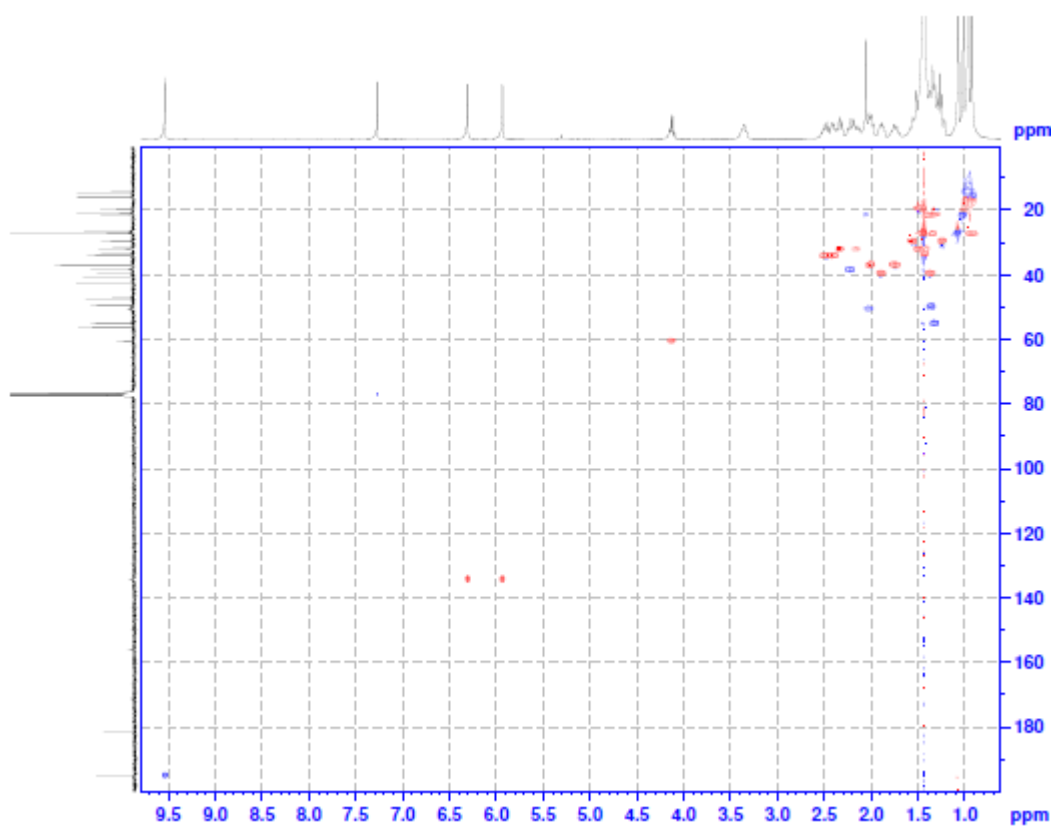


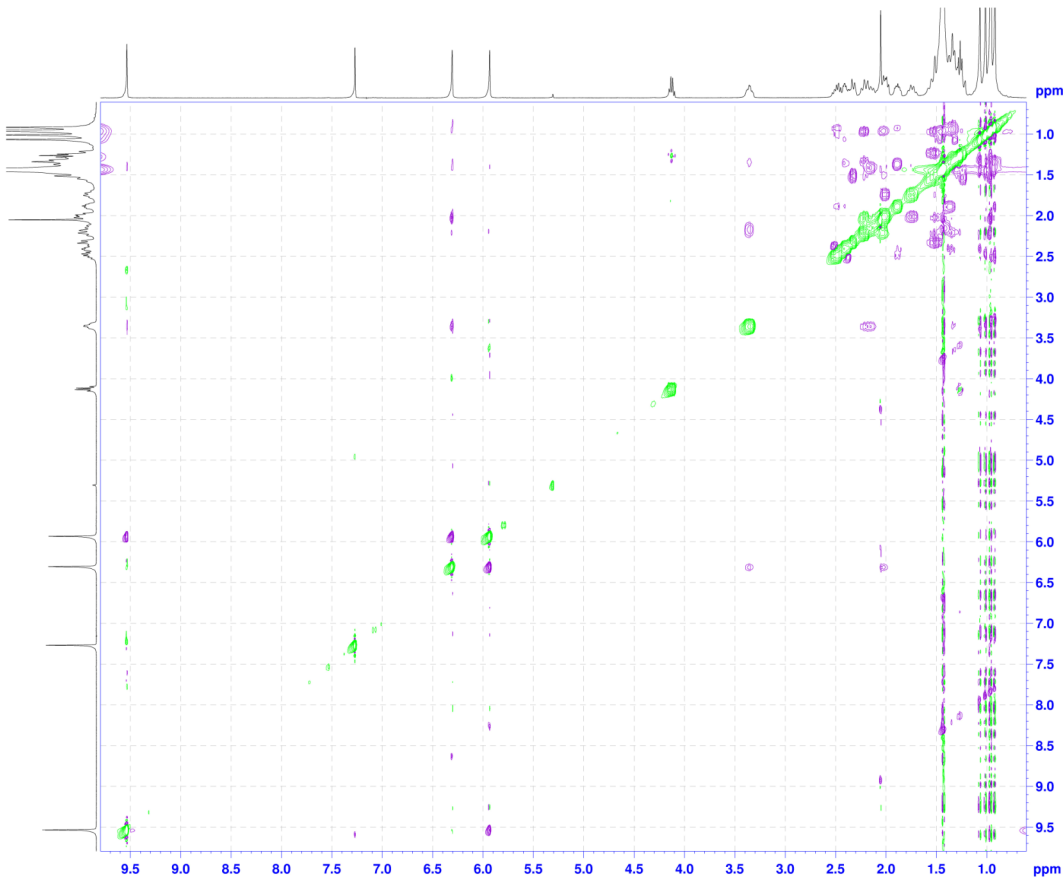
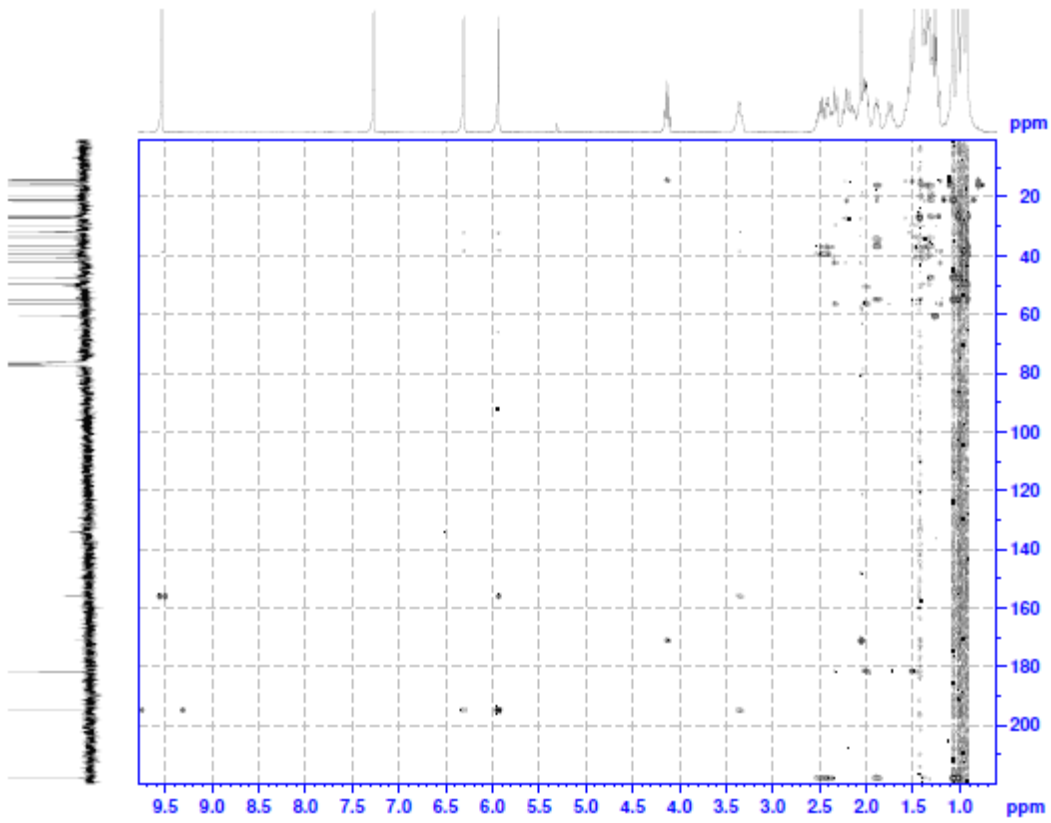


- Mass (ESI-: m/z 467 [M-H]⁻) and NMR (¹H + ¹³C + HSQC edit + HMBC + NOESY) spectra of 3,30-dioxolup-20(29)-en-28-oic acid (C₃₀H₄₄O₄; MW 468) (6):









```

Current Data Parameters
NAME      1-xc-mali-ho-ald-02
EXPNO     4
PROCNO    1

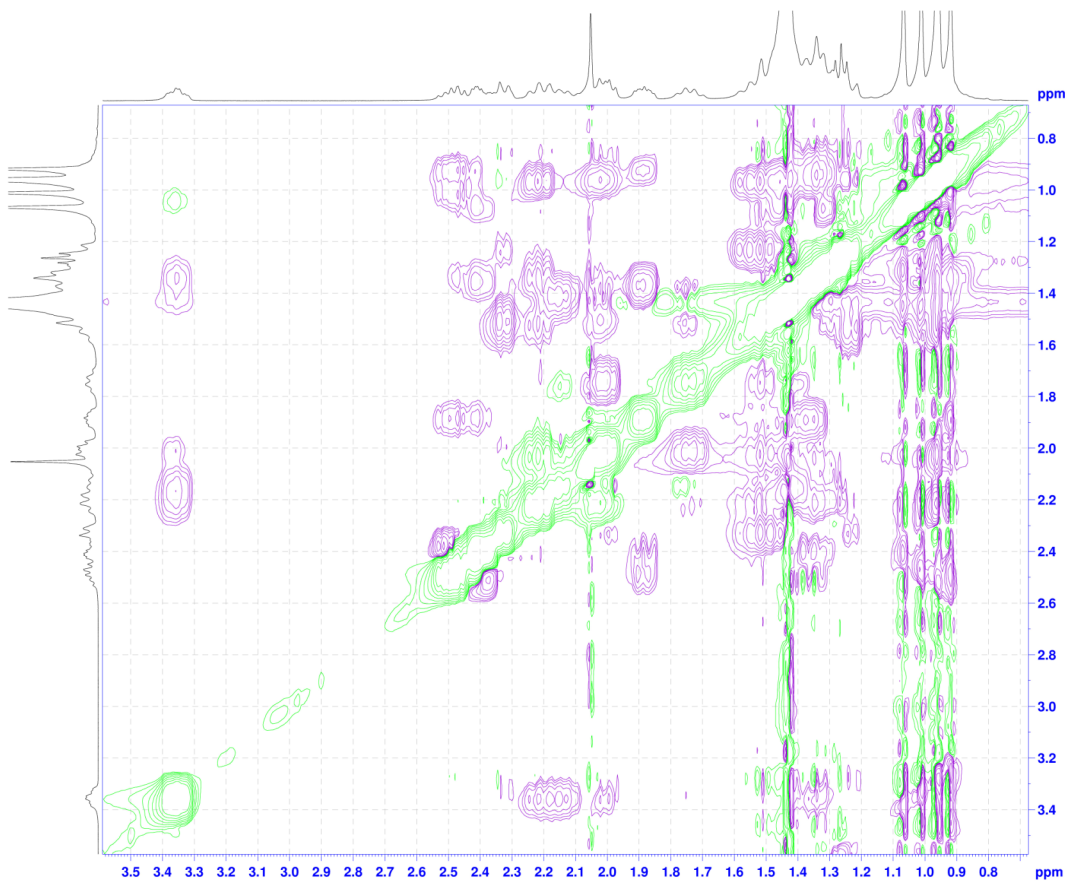
F2 - Acquisition Parameters
Date_     20100126
Time      17.39
INSTRUM   spect
PROBHD    5 mm Multinucl
PULPROG   noesyph
TD         2048
SOLVENT   CDCl3
NS         64
DS         16
SMH        3676.471 Hz
FIDRES     1.795152 Hz
AQ         0.2785760 sec
RG         64
DE         7.00 usec
TE         295.2 K
d0         0.00012862 sec
d1         2.00000000 sec
d2         0.69999999 sec
IN0        0.00027200 sec
STICNT    128

===== CHANNEL f1 =====
NUC1      1H
P1         5.80 usec
PL1        -1.00 dB
SFO1      400.1320965 MHz

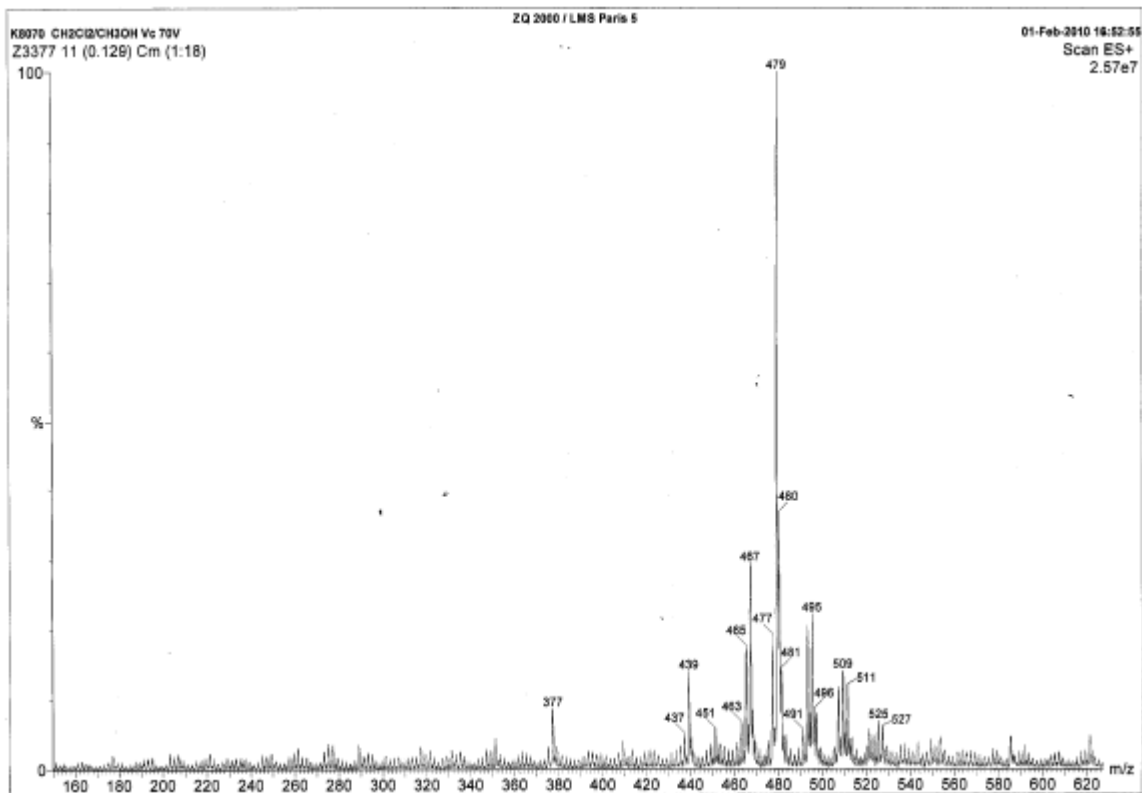
F1 - Acquisition parameters
ND0        1
TD         256
SFO1      400.1321 MHz
FIDRES     14.361214 Hz
SW         9.188 ppm
FwMODE     States-TPPI

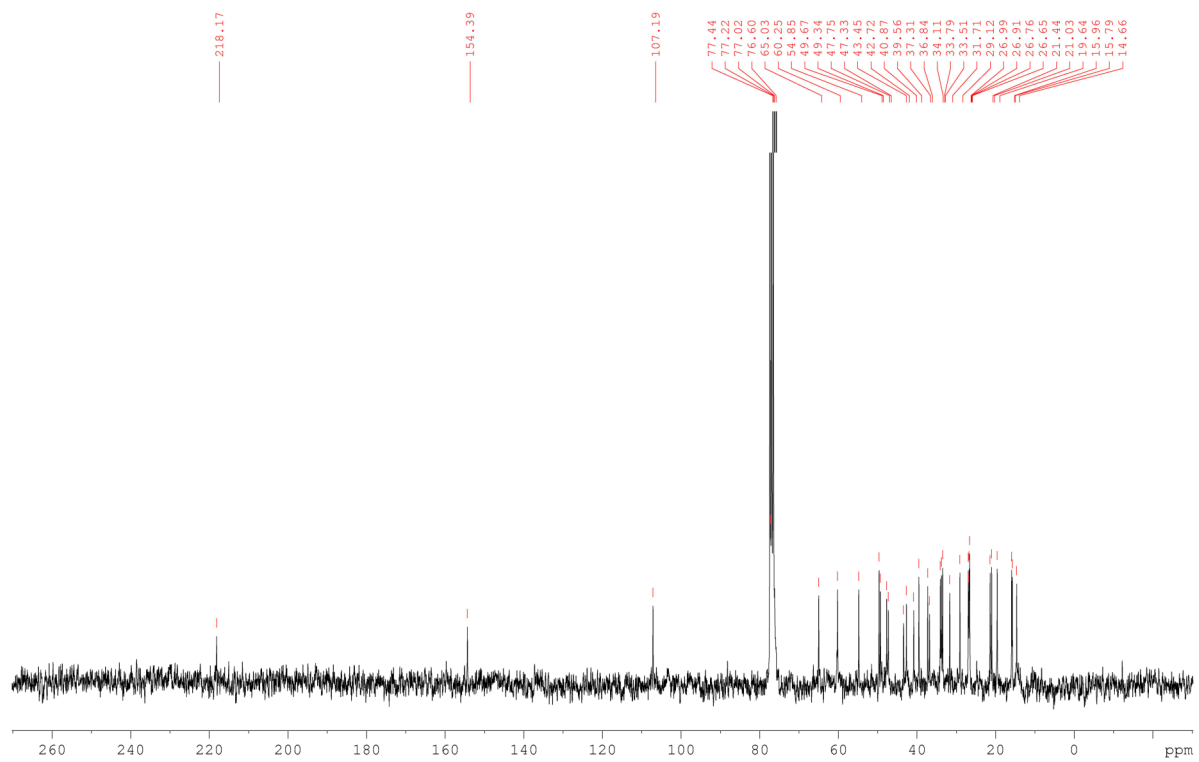
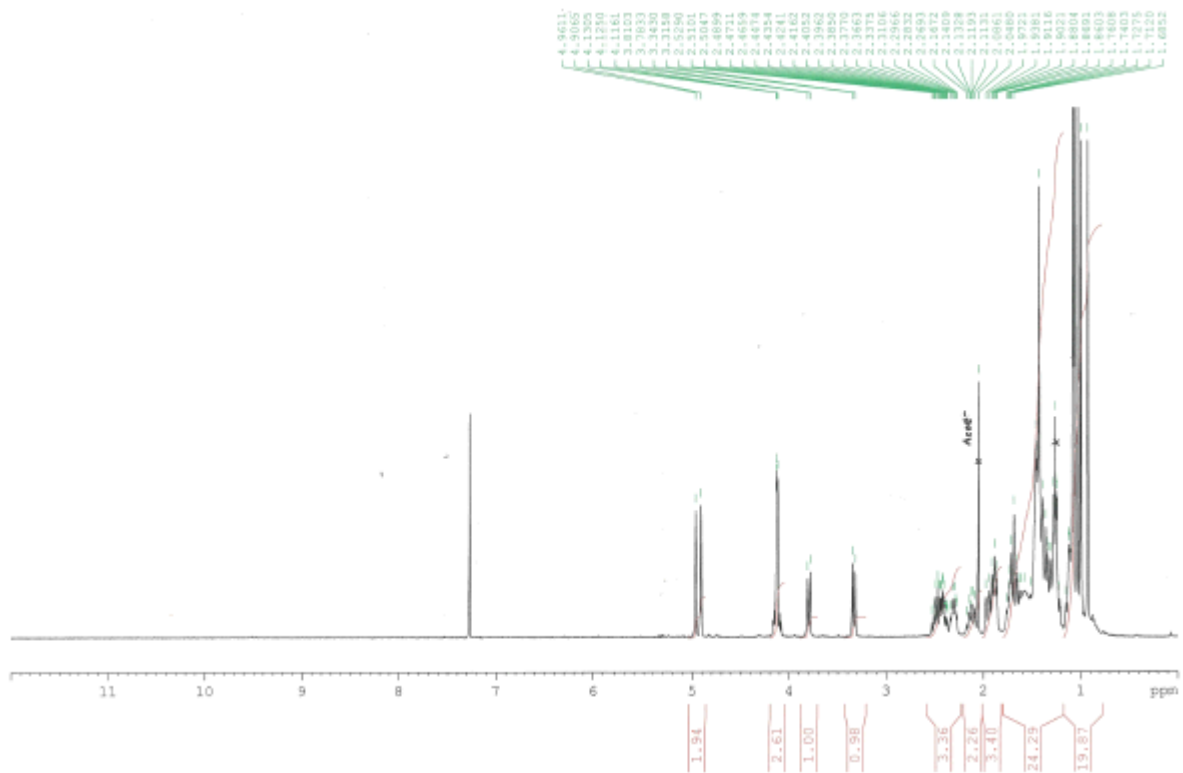
F2 - Processing parameters
SI         1024
SF         400.1300152 MHz
WDW        QSINE
SSB         2
LB         0.00 Hz
GB         0
PC         1.00

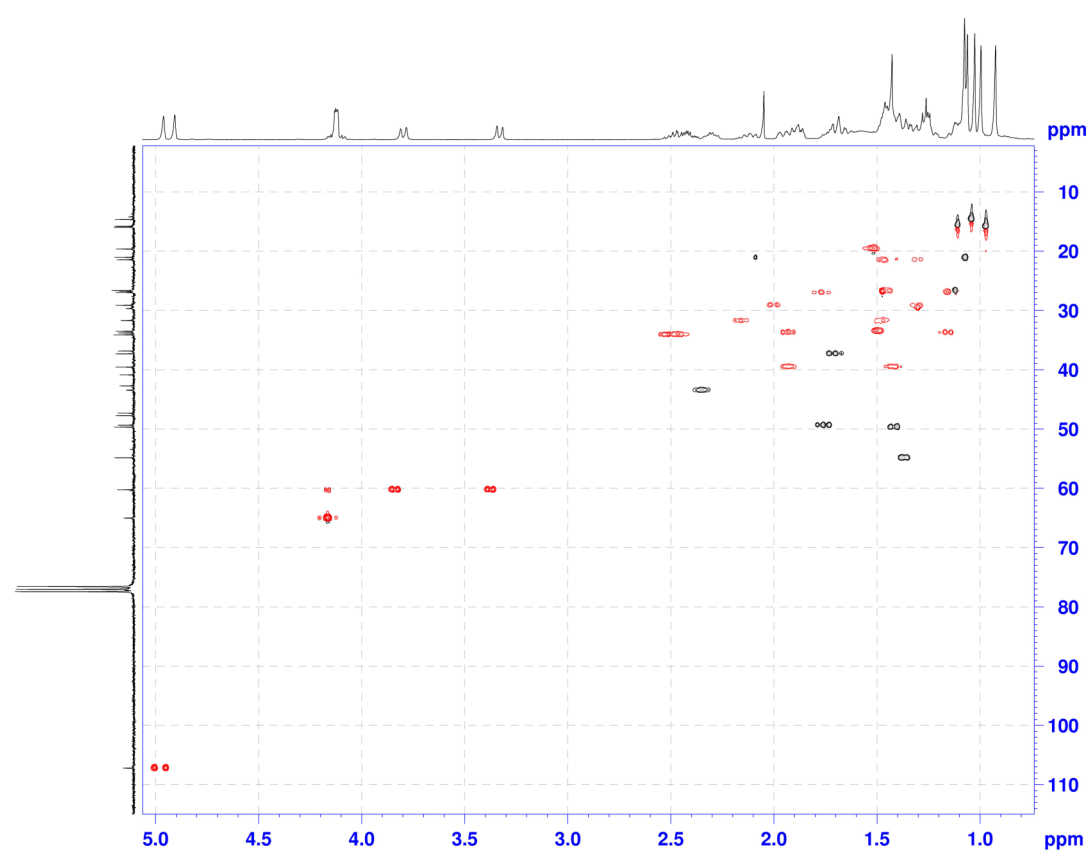
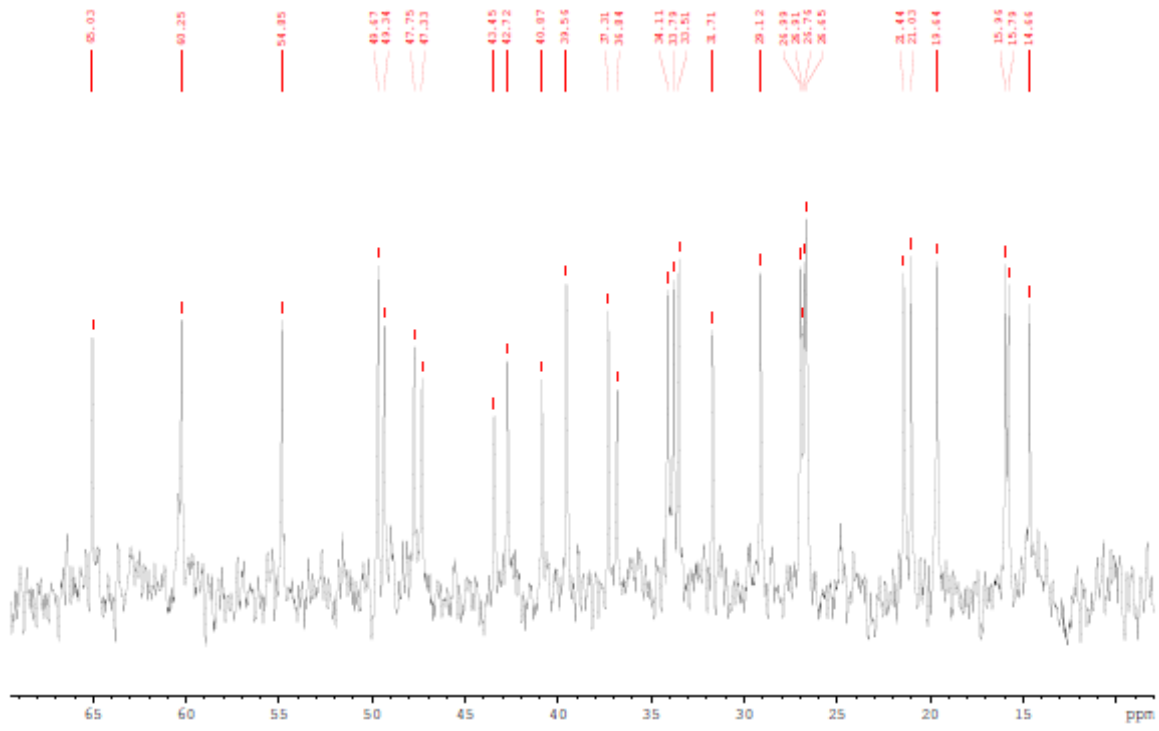
F1 - Processing parameters
SI         1024
SF         400.1300149 MHz
WDW        QSINE
SSB         2
LB         0.00 Hz
GB         0
  
```



- Mass (ESI+: m/z 479 $[M+Na]^+$) and NMR ($^1H + ^{13}C + HSQC$ edit + NOESY) spectra of 28,30-dihydroxy-3-oxolup-20(29)-ene ($C_{30}H_{48}O_3$; MW 456) (7):







Current Data Parameters
 NAME l-xc-mal-M10-38z
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20091114
 Time 11.29
 INSTRUM spect
 PROBP0 5 mm Multinuc1
 PULPROG zgpg30
 TD 2048
 ID 2048
 SOLVENT h2o
 SOLVENT2 ccc13
 NS 8
 DS 4
 SWH 1745.810 Hz
 FIDRES 0.852446 Hz
 AQ 0.3863972 sec
 RG 1890.4
 DW 286.400 usec
 DE 7.00 usec
 TE 296.2 K
 CHST2 145.000000 sec
 gd 0.00000300 sec
 d1 1.5000000 sec
 g4 0.00172414 sec
 d11 0.00000000 sec
 d13 0.00000400 sec
 D16 0.00000000 sec
 d21 0.00345000 sec
 DELTA 0.0023240 sec
 DELTA1 0.0007614 sec
 INO 0.00003105 sec
 STCNT 128
 ZGPTNS

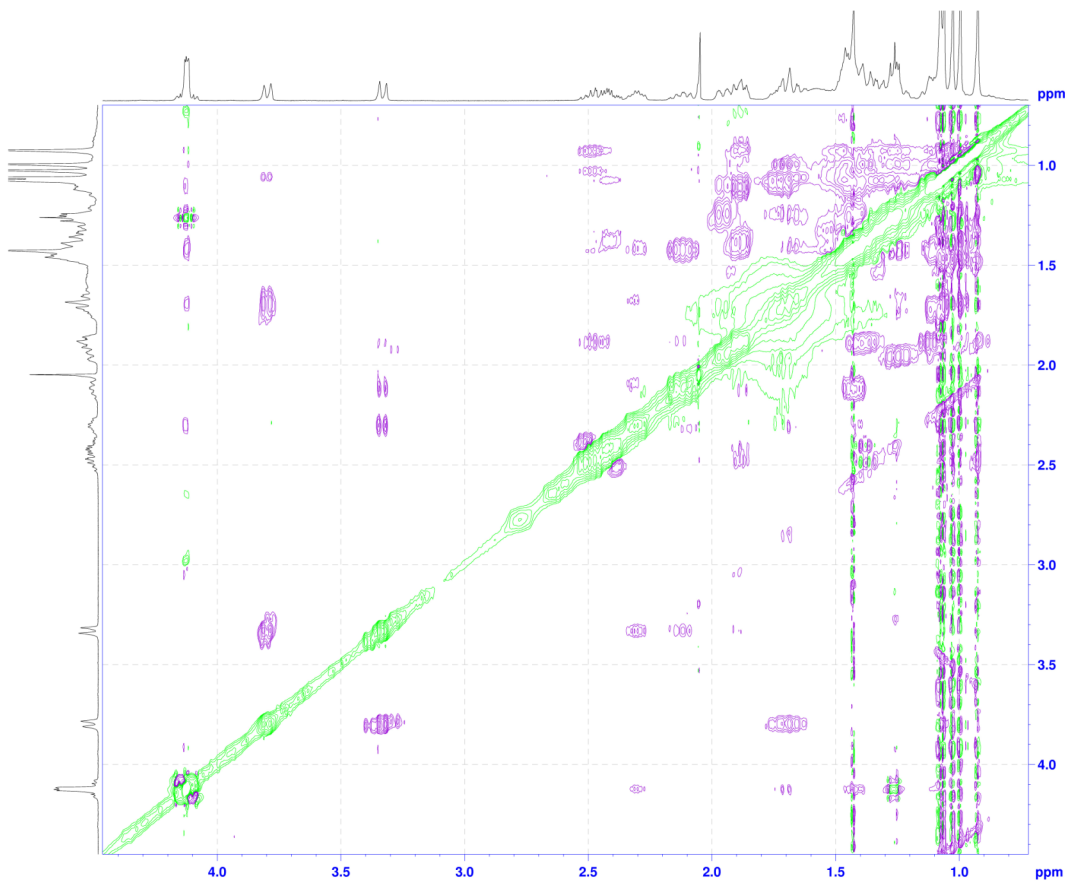
==== CHANNEL f1 =====
 NU01 1H
 P1 5.80 usec
 PC 11.40 usec
 P28 1000.00 usec
 PL1 -1.00 dB
 SFO1 400.1311561 MHz

==== CHANNEL f2 =====
 CPDPRG2 zgpg30
 NU02 13C
 P3 17.80 usec
 p4 35.60 usec
 PC02 60.00 usec
 PL2 -6.00 dB
 PL12 4.00 dB
 SFO2 100.6208180 MHz

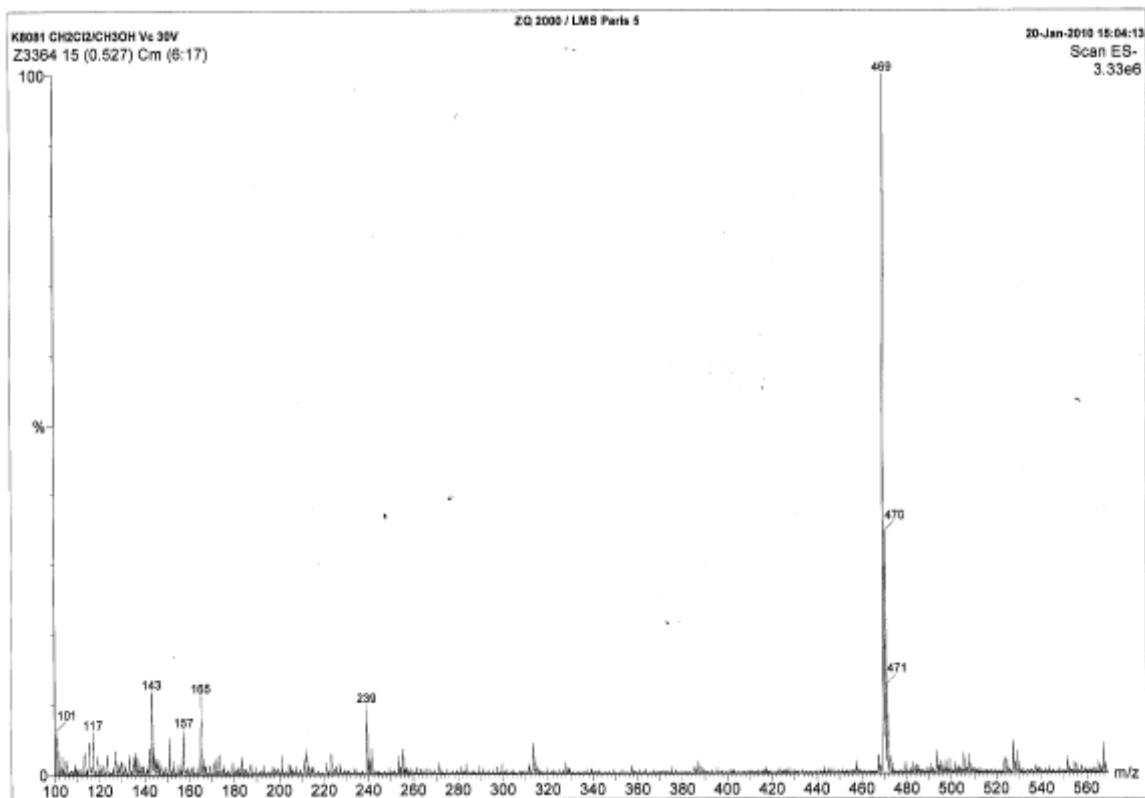
==== GRADIENT CHANNEL =====
 GPM01 SINE.100
 GPM02 SINE.100
 GP11 60.00 A
 GP22 20.10 A
 P16 1000.00 usec

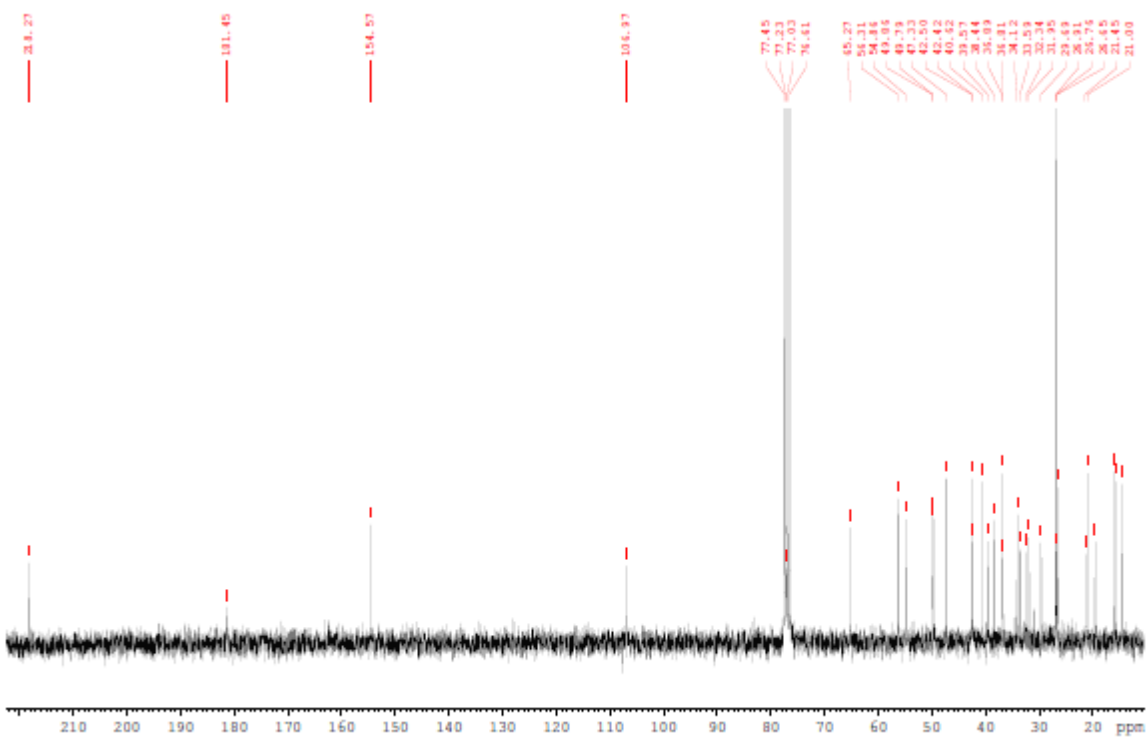
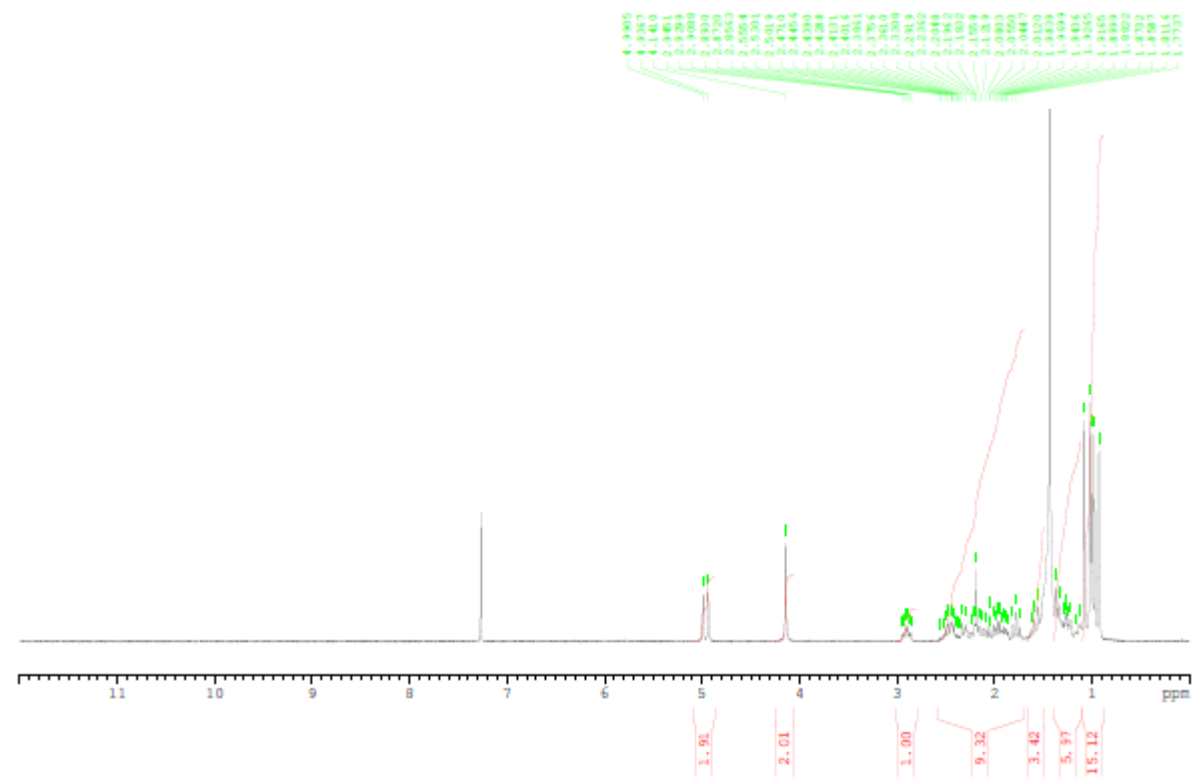
F1 - Acquisition parameters
 ID 2048
 SFO1 100.6208 MHz
 FIDRES 62.902576 Hz
 SW 145.037 ppm
 FWHM 0.852446 Hz
 F2 - Processing parameters
 S1 1024
 SF 400.1305000 MHz
 WDW EM
 SSB 0.00 Hz
 LB 0
 GB 1.40
 PC 0

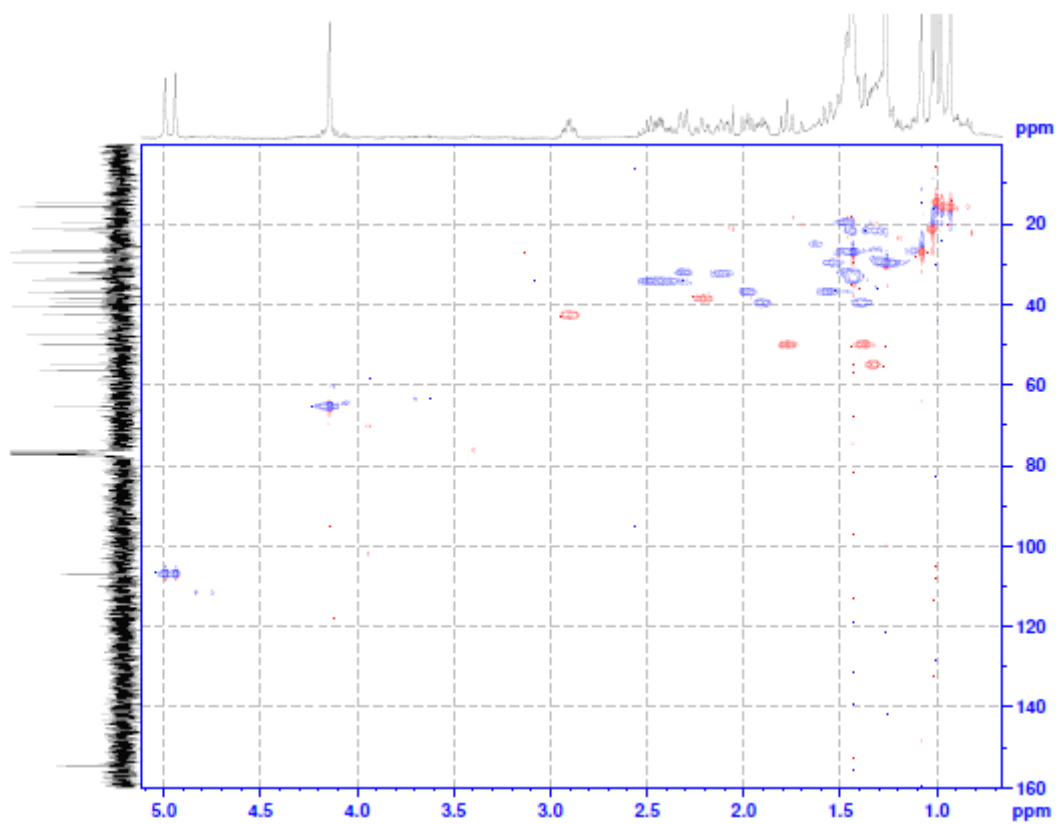
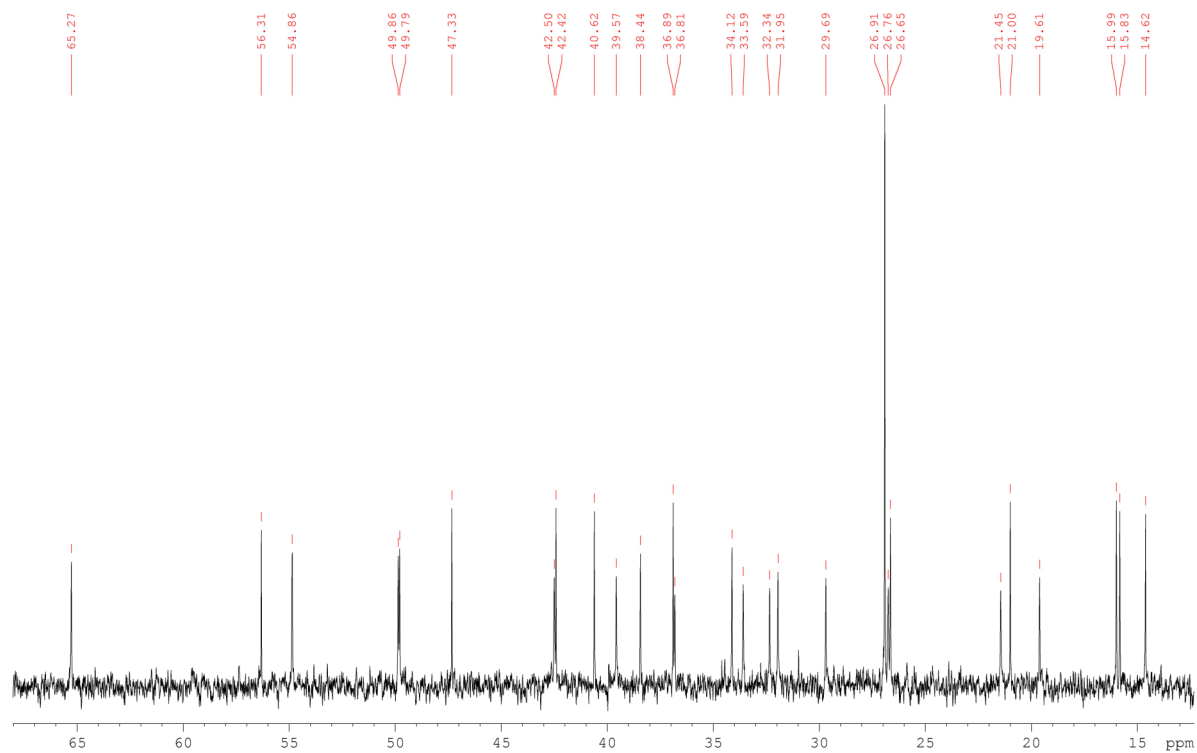
F1 - Processing parameters
 S1 1024
 MFC2 echo-antlecho
 SF 100.6127690 MHz
 WDW EM
 SSB 0.00 Hz
 LB 0



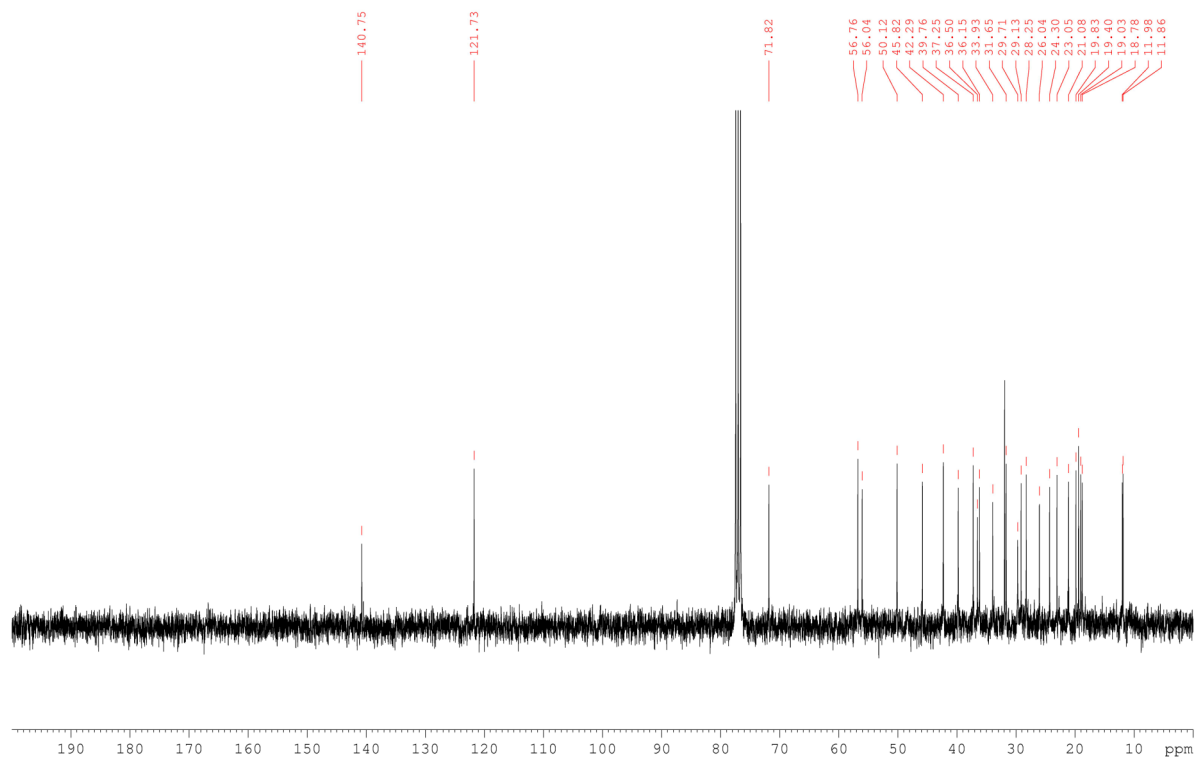
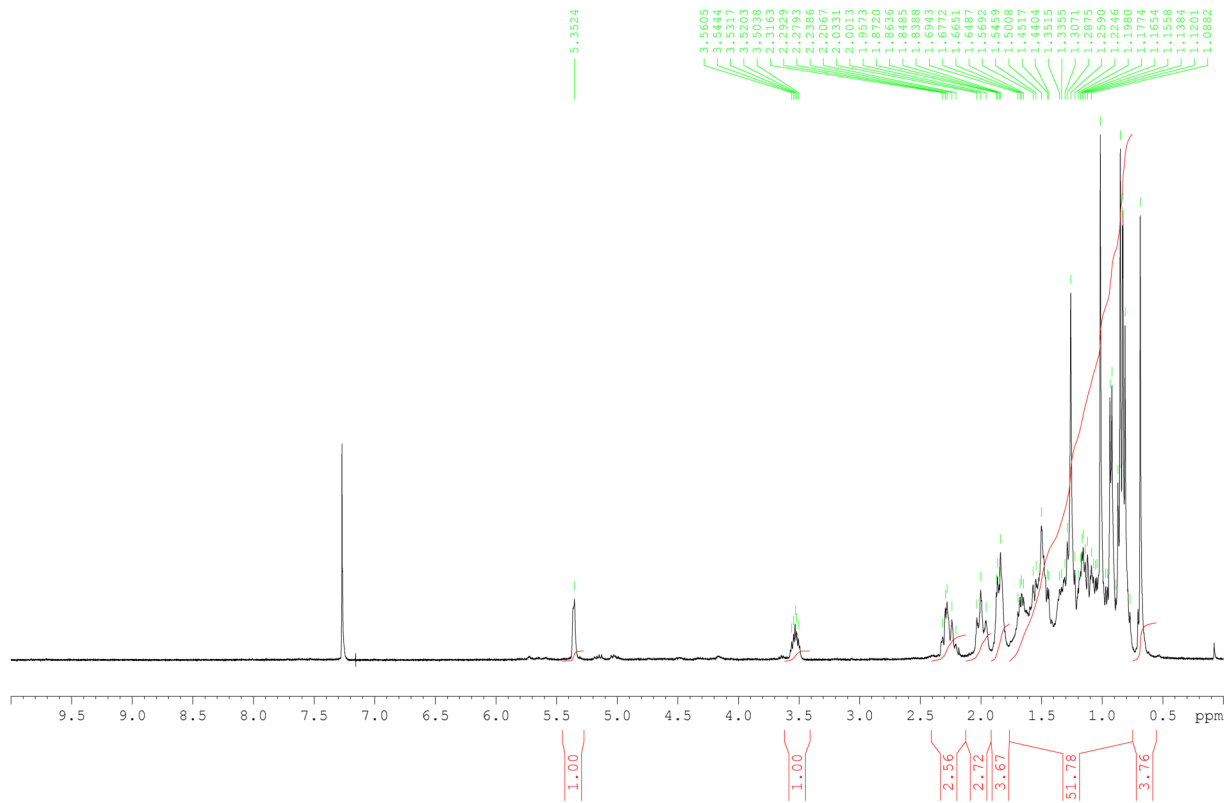
- Mass (ESI-: m/z 469 $[M-H]^-$) and NMR ($^1H + ^{13}C + HSQC$ edit) spectra of Messagenic acid G ($C_{30}H_{46}O_4$; MW 470) (8):

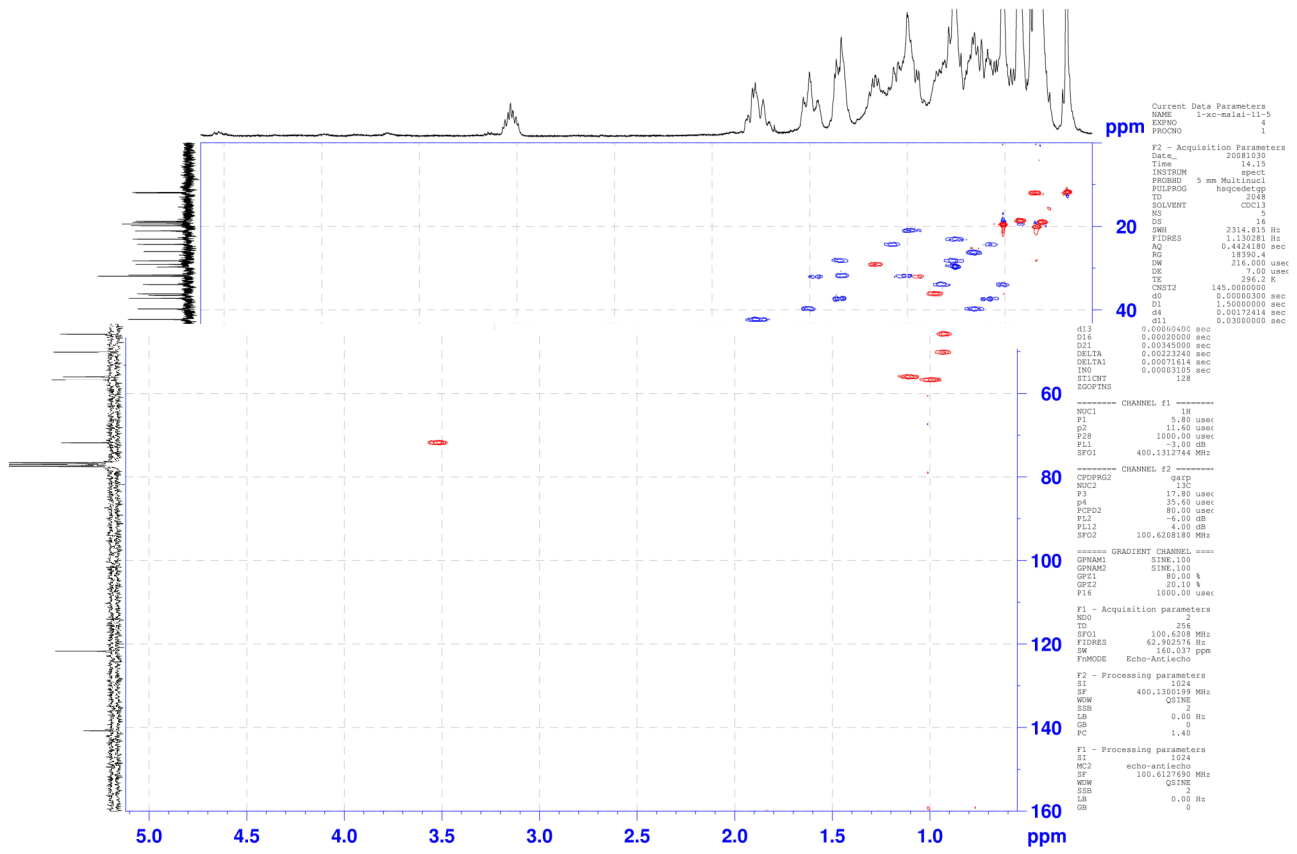






- Mass (EI: m/z 414 $[M]^+$) and NMR ($^1\text{H}+^{13}\text{C}$ +HSQC edit) spectra of β -sitosterol ($\text{C}_{29}\text{H}_{50}\text{O}$) (9):





```

Current Data Parameters
NAME      1-2c-m1a1-11-5
EXPNO     4
PROCNO    1

F2 - Acquisition Parameters
Date_     20081030
Time      14:15
INSTRUM   spect
PROBHD    5 mm Multispl1
PULPROG   hsqcsetep
TD         2688
SOLVENT   CDCl3
NS         3
DS         16
SWH        2314.815 Hz
FIDRES     1.1130281 Hz
AQ         0.4424180 sec
RG         18360.4
DM         216.000 usec
DE         7.50 usec
TE         296.2 K
CNS2      145.000000
d0         0.0000300 sec
D1         1.5000000 sec
d4         0.00172414 sec
d11        0.0300000 sec
d13        0.0000400 sec
D14        0.0000000 sec
D21        0.00345000 sec
DELTA     0.0003140 sec
DELTA1    0.00071614 sec
lD0        0.00003100 sec
STICHT    128
ZGPGTNS

===== CHANNEL f1 =====
NUC1       1H
P1         5.80 usec
p2         11.60 usec
P38        1800.00 usec
PL1        -3.00 dB
SFO1       400.1317444 MHz

===== CHANNEL f2 =====
CPDPRG2   gqip
NUC2       13C
P1         17.80 usec
p1         35.60 usec
P3P2       80.00 usec
PL2        -6.00 dB
PL12       4.00 dB
SFO2       100.6208180 MHz

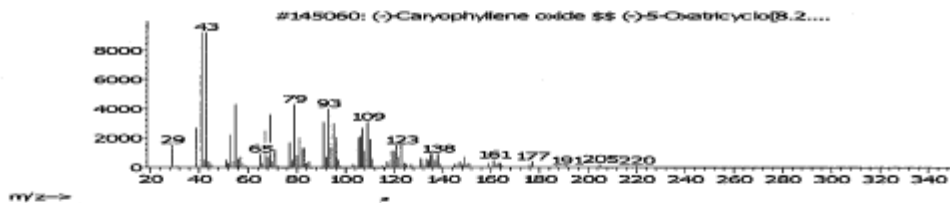
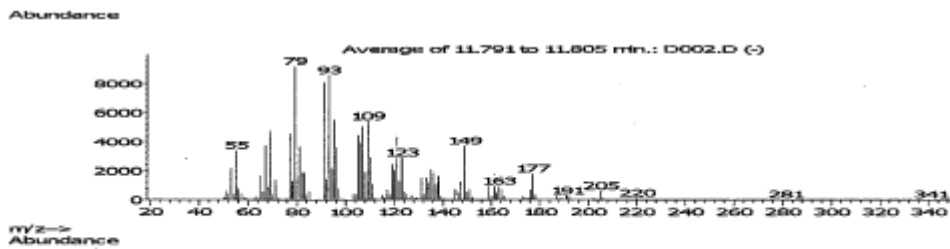
===== GRADIENT CHANNEL =====
GPRAM1    SINE.100
GPRAM2    SINE.100
GR1        80.00 %
GR2        20.10 %
P15        1800.00 usec

F1 - Acquisition parameters
ND0        2
TD         256
SFO1       100.6208 MHz
FIDRES     62.802076 Hz
SW         160.037 ppm
FWDOR      Echo-Antiecho

F2 - Processing parameters
SI         1024
SF         400.1300199 MHz
QSINSE    2
SIB        2
lB         0.00 Hz
GB         0
PC         1.40

F1 - Processing parameters
SI         1024
MC2        echo-antiecho
SF         100.6177690 MHz
QSINSE    2
SIB        2
lB         0.00 Hz
GB         0
  
```

- Mass (EI: m/z 220 $[M]^+$) and NMR ($^1H+^{13}C$) spectra of β -caryophyllene oxide ($C_{15}H_{24}O$; MW 220) (10):



Match Quality 92

Name (-)-Caryophyllene oxide \$\$ (-)-5-Oxatricyclo[8.2.0.0(4,6)]dodecane,, 12-trimethyl-9-methylene-, [1R-(1R*,4R*,6R*,10S*)]- (CAS) \$\$ (-)-.beta.-Caryophyllene epoxide \$\$
 Caryophyllene oxide \$\$ (-)caryophyllene oxide \$\$ 5-Oxatricyclo[8.2.0.0(4,6)-]dodecane, 4,1
 CAS Number 001139-30-6
 Molecular Formula $C_{15}H_{24}O$
 Molecular Weight 220.18

