

## ECOLOGY

## Naturally acidified habitat selects for ocean acidification–tolerant mussels

Jörn Thomsen,<sup>1\*</sup> Laura S. Stapp,<sup>2,3</sup> Kristin Haynert,<sup>1,4</sup> Hanna Schade,<sup>1</sup> Maria Danelli,<sup>1</sup> Gisela Lannig,<sup>2</sup> K. Mathias Wegner,<sup>5</sup> Frank Melzner<sup>1</sup>

Ocean acidification severely affects bivalves, especially their larval stages. Consequently, the fate of this ecologically and economically important group depends on the capacity and rate of evolutionary adaptation to altered ocean carbonate chemistry. We document successful settlement of wild mussel larvae (*Mytilus edulis*) in a periodically CO<sub>2</sub>-enriched habitat. The larval fitness of the population originating from the CO<sub>2</sub>-enriched habitat was compared to the response of a population from a nonenriched habitat in a common garden experiment. The high CO<sub>2</sub>-adapted population showed higher fitness under elevated P<sub>CO<sub>2</sub></sub> (partial pressure of CO<sub>2</sub>) than the non-adapted cohort, demonstrating, for the first time, an evolutionary response of a natural mussel population to ocean acidification. To assess the rate of adaptation, we performed a selection experiment over three generations. CO<sub>2</sub> tolerance differed substantially between the families within the F<sub>1</sub> generation, and survival was drastically decreased in the highest, yet realistic, P<sub>CO<sub>2</sub></sub> treatment. Selection of CO<sub>2</sub>-tolerant F<sub>1</sub> animals resulted in higher calcification performance of F<sub>2</sub> larvae during early shell formation but did not improve overall survival. Our results thus reveal significant short-term selective responses of traits directly affected by ocean acidification and long-term adaptation potential in a key bivalve species. Because immediate response to selection did not directly translate into increased fitness, multigenerational studies need to take into consideration the multivariate nature of selection acting in natural habitats. Combinations of short-term selection with long-term adaptation in populations from CO<sub>2</sub>-enriched versus nonenriched natural habitats represent promising approaches for estimating adaptive potential of organisms facing global change.

## INTRODUCTION

Ocean acidification, caused by rising atmospheric CO<sub>2</sub> concentrations due to excess fossil fuel burning, severely affects many marine organisms (1). Calcifying organisms are especially affected by the change of ocean chemistry because their ability to form calcified structures is reduced. Bivalves are among the most vulnerable taxonomic groups because their CaCO<sub>3</sub>-containing shells protect the animal from predation. In particular, their larval stages suffer from substantial reductions in growth and survival under elevated P<sub>CO<sub>2</sub></sub> (partial pressure of CO<sub>2</sub>) (2–5). This is likely a consequence of the high calcification rates during the formation of the first larval shell (6, 7). Although the benthic life stage is able to compensate for the negative impact even of highly elevated P<sub>CO<sub>2</sub></sub> (~3000 μatm) when food supply is abundant, early larval development is completely fueled by the limited energy provided by the egg and thus represents an important ecological bottleneck (6, 7). Bivalves of the genus *Mytilus* have important ecological roles in boreal, benthic ecosystems and can contribute by up to 90% to epibenthic biomass in coastal habitats (8). In addition, their high economic value for aquaculture has stimulated a number of recent studies to estimate their adaptation potential to future ocean conditions (4, 9–11). However, the relatively long generation time of bivalve species complicates multigenerational (MG) studies. Consequently, most studies until now have estimated evolutionary potential by quantifying variation of fitness-relevant traits such as growth during early development within and between locally adapted populations (10, 12, 13) or assessed transgenerational phenotypic plasticity (10). These studies did not provide a uniform picture on the potential of bivalves to

adapt to ocean acidification. Modeling the rate of adaptation in a single population based on larval shell size variations within the first 60 hours of development under elevated P<sub>CO<sub>2</sub></sub> (1000 μatm) suggested a low potential for adaptation when extrapolated over 50 generations (8). To capture longer time periods, comparison of populations naturally experiencing differing carbonate system conditions offers useful proxies for estimating adaptation [space-for-time substitution (14)]. In one study, the growth response of field-collected juvenile mytilid mussels originating from two populations differed under elevated P<sub>CO<sub>2</sub></sub>, indicating local adaptation (12). In another, larval shell development of two *Mytilus* species was similar during exposure to varying carbonate system treatments, although one species originated from a habitat that encounters upwelling events associated with elevated P<sub>CO<sub>2</sub></sub> (13). Although these studies suggested that bivalves can potentially adapt to rising P<sub>CO<sub>2</sub></sub>, they lack a formal estimation of genetic versus nongenetic sources of variation. For example, transgenerational acclimation to elevated P<sub>CO<sub>2</sub></sub> can substantially modulate fitness of offspring as observed in fish (15). In the oyster *Saccostrea glomerata*, a 5-week exposure of parental animals to elevated P<sub>CO<sub>2</sub></sub> during gametogenesis enhanced the development and growth rates of F<sub>1</sub> and even F<sub>2</sub> offspring under acidified conditions (9, 16). Selective breeding for aquaculture purposes substantially increased the productivity of *S. glomerata* and resulted in 25% improved growth within two to four generations (17). The higher developmental rates of this breeding line were also maintained under elevated P<sub>CO<sub>2</sub></sub> compared to wild-type oysters (10). Therefore, rapid evolutionary responses in bivalves may enable adaptation to ocean acidification, but these MG selection studies using continuously elevated P<sub>CO<sub>2</sub></sub> as selective agent are lacking so far.

To fill this gap, we performed a 3-year MG experiment to test whether the blue mussel *Mytilus edulis* can successfully adapt to ocean acidification and to estimate which mechanisms contribute to rapid evolutionary responses. The tested population inhabits the seasonally acidified Kiel Fjord, western Baltic Sea, which is characterized by low pH and elevated P<sub>CO<sub>2</sub></sub> levels during the reproductive period of the species

<sup>1</sup>Helmholtz Centre for Ocean Research Kiel (GEOMAR), 24105 Kiel, Germany. <sup>2</sup>Integrative Ecophysiology, Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research, 27570 Bremerhaven, Germany. <sup>3</sup>University of Bremen, 28359 Bremen, Germany. <sup>4</sup>Marine Research Department, Senckenberg am Meer, 26382 Wilhelmshaven, Germany. <sup>5</sup>Coastal Ecology, Wadden Sea Station Sylt, Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research, 25992 List/Sylt, Germany.

\*Corresponding author. Email: jthomsen@geomar.de

(18, 19). This experiment was supported by field monitoring of carbonate chemistry variation in relation to mussel settlement patterns. In addition, we compared the Baltic population in a common garden experiment to the response of mussels from the North Sea, which is characterized by less variable pH conditions and higher seawater alkalinity due to higher salinity (20).

To investigate the time scale of adaptation to ocean acidification, we conducted two experiments to compare long-term adaptation between populations and processes of short-term adaptation within a population. We hypothesized that Baltic mussels have already adapted to high- $\text{CO}_2$  seawater and would better tolerate simulated ocean acidification than North Sea mussels. Furthermore, we hypothesized that selection for ocean acidification-tolerant specimens would increase the fitness of their offspring when exposed to acidified conditions.

## RESULTS

### Field carbonate chemistry monitoring and larval settlement (Baltic Sea)

Monitoring of pH in Sylt and Kiel Fjord revealed higher and more stable pH in the North Sea habitat compared to the habitat of the Baltic population (fig. S1). Monthly mean pH in Sylt remained above 8, with maximum values recorded during spring bloom in April (Fig. 1A). In contrast, mean pH values declined to about 7.7 during the upwelling period in summer and autumn in Kiel Fjord. Our monitoring of mussel settlement on weekly deployed panels and continuously logged seawater  $P_{\text{CO}_2}$  revealed that bivalve larvae survived and settled in Kiel Fjord, which is characterized by elevated and fluctuating  $P_{\text{CO}_2}$ . The hourly averaged  $P_{\text{CO}_2}$  was  $1087 \pm 537 \mu\text{atm}$  and ranged between 266 and 2861  $\mu\text{atm}$  over the whole monitoring period from mid-July to mid-September 2012 (Fig. 1B). Despite such high and fluctuating environmental  $P_{\text{CO}_2}$ , Baltic mussels settled successfully, with a peak of more than 1000 larvae settled per panel in early August (Fig. 1C) at elevated  $P_{\text{CO}_2}$  levels similar to those predicted for the average surface ocean of 2100. Because  $P_{\text{CO}_2}$  fluctuated rapidly due to upwelling events in Kiel Fjord, environmental conditions experienced by different larval cohorts differed significantly (fig. S2). Earlier settlers of the July cohort experienced only moderately elevated  $P_{\text{CO}_2}$  because larvae avoided the first pronounced upwelling peak at the beginning of August (Fig. 1C) (mean  $P_{\text{CO}_2}$ , 826  $\mu\text{atm}$ ; range, 266 to 1502  $\mu\text{atm}$ ). In contrast, larvae that settled at the end of August experienced  $P_{\text{CO}_2}$  levels between 443 and 2861  $\mu\text{atm}$  (mean, 1191  $\mu\text{atm}$ ) during a calculated 27-day larval phase (Fig. 1C). Larvae settling in mid-September were again exposed to lower and more stable  $P_{\text{CO}_2}$  levels (larval phase, 25 days; mean, 859  $\mu\text{atm}$ ; range, 427 to 2225  $\mu\text{atm}$ ) (Fig. 1C). The number of days August and September cohorts were exposed to daily mean  $P_{\text{CO}_2}$  values above 1000  $\mu\text{atm}$  differed, with 17 and 5 days corresponding to 63 and 20% of their estimated whole planktonic life phase, respectively.

### Population comparison experiment (Baltic Sea versus North Sea)

The formation of the first larval shell [prodissoconch I (PD I)] (fig. S3) (21) was strongly delayed in both North Sea and Baltic Sea populations at high  $P_{\text{CO}_2}$ , which resulted in significantly reduced shell length compared to larvae from the control  $P_{\text{CO}_2}$  [two-way analysis of variance (ANOVA): population:  $F = 1.6$ ,  $P > 0.05$ ;  $P_{\text{CO}_2}$ :  $F = 112.1$ ,  $P < 0.01$ ]. However, Baltic mussel larvae were less affected and showed a smaller shell length reduction compared to North Sea larvae at elevated  $P_{\text{CO}_2}$  [−24% (Baltic Sea) versus −38% (North Sea) shell length

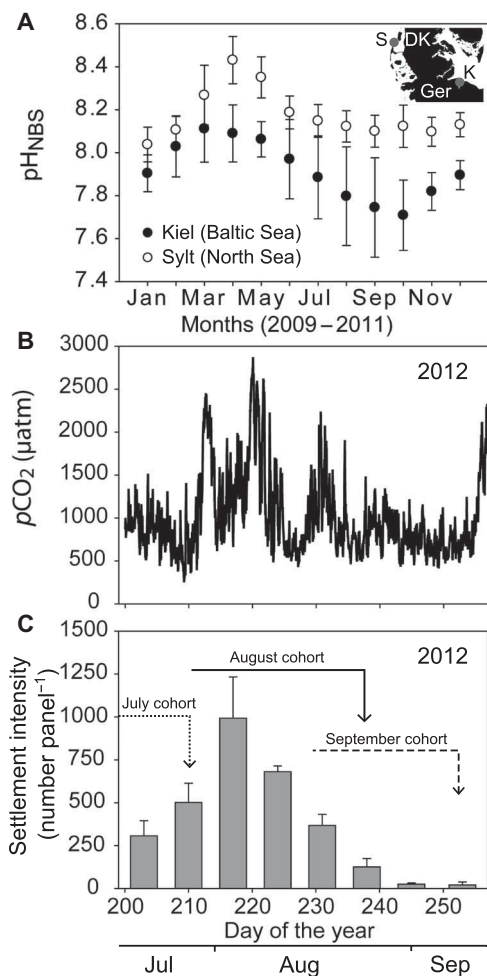
compared to respective controls; population  $\times P_{\text{CO}_2}$ :  $F = 6.5$ ,  $P < 0.05$ ] (Fig. 2A).

Growth patterns translated well into observed survival. Here, survival at 390- $\mu\text{atm}$   $P_{\text{CO}_2}$  did not differ between the two populations, but Baltic larval survival was higher at elevated  $P_{\text{CO}_2}$  (two-way ANOVA: population:  $F = 0.8$ ,  $P > 0.05$ ;  $P_{\text{CO}_2}$ :  $F = 10.9$ ,  $P < 0.01$ ; population  $\times P_{\text{CO}_2}$ :  $F = 6.9$ ,  $P < 0.05$ ) (Fig. 2C and table S1). Subsequent shell growth rates were similar in Baltic larvae exposed to 390- and 2400- $\mu\text{atm}$   $P_{\text{CO}_2}$  (Fig. 2B; no data for North Sea larvae at 2400- $\mu\text{atm}$   $P_{\text{CO}_2}$  because of high mortality).

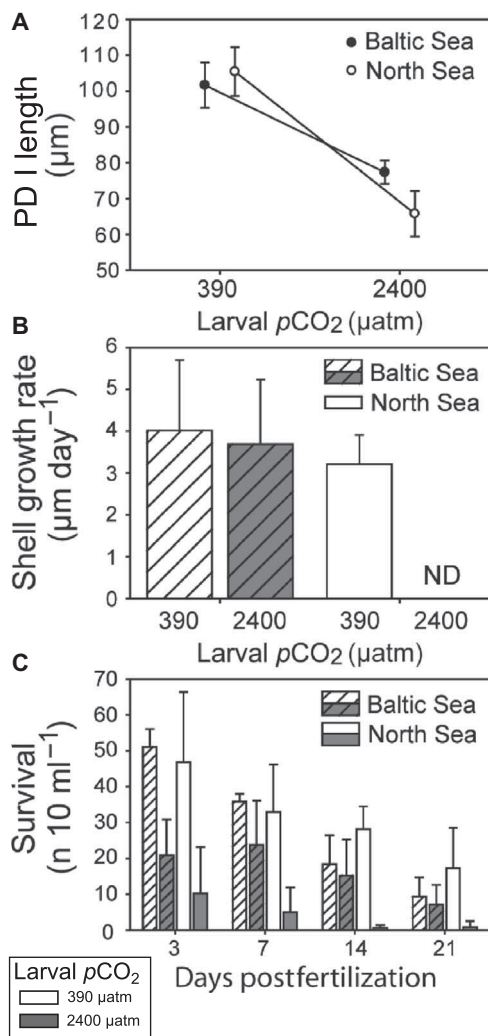
### Three-year MG experiment (Baltic Sea)

#### $F_0$ and $F_1$ generation (2012).

The MG experiment utilized controlled genetic crosses of mussels collected from the more  $P_{\text{CO}_2}$ -tolerant Baltic Sea population to select for  $\text{CO}_2$ -tolerant and  $\text{CO}_2$ -sensitive families, which were used to elucidate the relative contribution of genetic and nongenetic environmental factors enabling adaptation to ocean acidification (Fig. 3B). PD I size was strongly reduced in  $F_1$  larvae exposed to elevated  $P_{\text{CO}_2}$  and declined

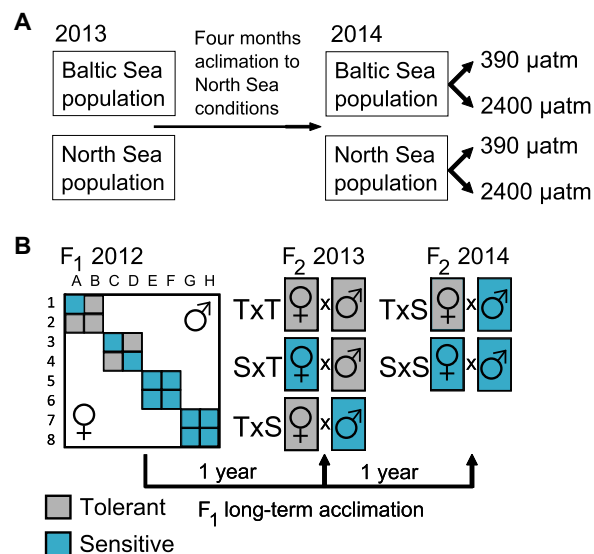


**Fig. 1. Habitat carbonate system variability and juvenile settlement.** (A) Averaged monthly pH values recorded from 2009 to 2011 in the habitats of the two tested populations. The inset depicts the location of the two habitats in North Sea and Baltic Sea (S, Sylt; K, Kiel; DK, Denmark; Ger, Germany). (B) Continuously measured seawater  $P_{\text{CO}_2}$  in summer 2012 in Kiel Fjord. (C) Mussel larval settlement intensity on panels between July and September. The arrows correspond to the estimated planktonic phases of larvae settled in July, August, and September. Values are means  $\pm$  SD.



**Fig. 2. Larval performance of Baltic Sea and North Sea populations exposed to elevated P<sub>CO<sub>2</sub></sub>.** (A) PD I length of both populations declined at high P<sub>CO<sub>2</sub></sub>, but Baltic larval size was less affected ( $n = 22$  to  $73$ ). (B) Daily shell growth was similar for both populations and P<sub>CO<sub>2</sub></sub> treatments [no data (ND) for North Sea larvae at high P<sub>CO<sub>2</sub></sub> due to low survival on days 14 and 21]. (C) Survival rapidly declined in North Sea larvae exposed to elevated P<sub>CO<sub>2</sub></sub>, whereas Baltic Sea larvae were less affected by elevated P<sub>CO<sub>2</sub></sub> from day 7 forward. Values are means  $\pm$  SD; numbers in bracket state the number of measured individuals per P<sub>CO<sub>2</sub></sub> treatment.

from  $112 \pm 6 \mu\text{m}$  at  $390 \mu\text{atm}$  to  $94 \pm 7 \mu\text{m}$  and  $78 \pm 8 \mu\text{m}$  at  $1120$  and  $2400 \mu\text{atm}$ , respectively (ANOVA:  $P_{\text{CO}_2}$ :  $F = 165.1$ ,  $P < 0.001$ ) (Fig. 4A). Family-specific PD I shell length varied substantially, and the calculated heritability for this trait was  $0.56$  [confidence interval (CI),  $0.27$  to  $0.81$ ],  $0.47$  (CI,  $0.24$  to  $0.81$ ), and  $0.53$  (CI,  $0.23$  to  $0.83$ ) at  $390$ -,  $1120$ -, and  $2400$ - $\mu\text{atm}$  P<sub>CO<sub>2</sub></sub>, respectively. Subsequently, larvae from all P<sub>CO<sub>2</sub></sub> treatments grew at comparable rates and thus reached similar sizes at the end of the planktonic phase (Fig. 4C). Because of the large variance in final larval survival between families, our study showed no significant difference in larval survival between  $390$ - and  $1120$ - $\mu\text{atm}$  P<sub>CO<sub>2</sub></sub>, but did reveal a drastic reduction at  $2400 \mu\text{atm}$  (Fig. 4E and table S2). Similarly, larvae from all families successfully settled at the two lower P<sub>CO<sub>2</sub></sub> levels, but only the offspring of five families (classified as “tolerant” families A2, B1, B2, C4, and D3) metamorphosed into juveniles at  $2400 \mu\text{atm}$  (Fig.



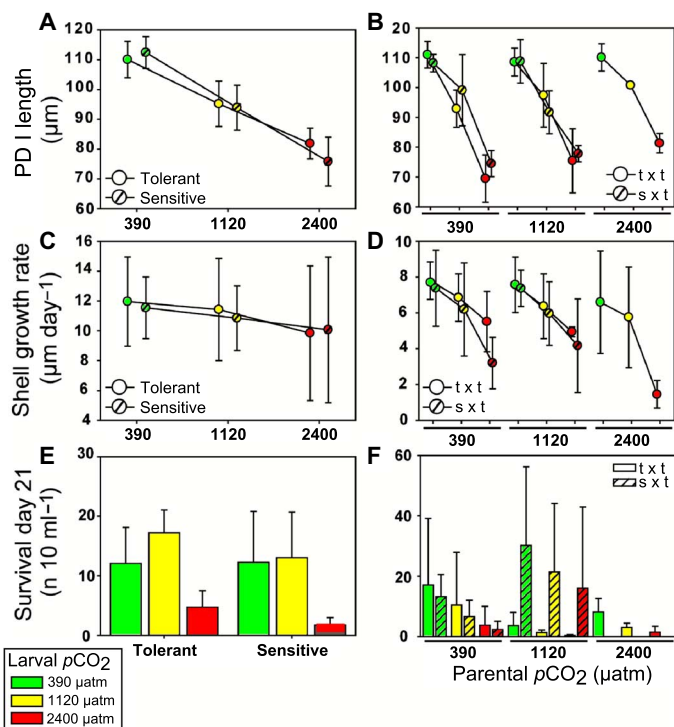
**Fig. 3. Flow chart of the experimental approach.** (A) Collection of mussels from North Sea and Baltic Sea population, acclimation to North Sea conditions from December 2013 to April 2014, and subsequent transfer, spawning, and exposure to two P<sub>CO<sub>2</sub></sub> treatments. (B) Crossing schemes for F<sub>1</sub> and F<sub>2</sub> generation. In 2012, F<sub>0</sub> animals (eight dams and eight sires) were crossed pairwise to generate 16 full-sib F<sub>1</sub> families nested within four half-sib groups. Settled mussels were subsequently acclimated for 1 and 2 years. In 2013, F<sub>2</sub> was generated by performing single-specimen crosses using pure F<sub>1</sub> tolerant lines as well as tolerant and sensitive F<sub>1</sub> families selected in, and long-term acclimated to,  $390$ -,  $1120$ -, and  $2400$ - $\mu\text{atm}$  (tolerant only) P<sub>CO<sub>2</sub></sub>. In 2014, F<sub>2</sub> was generated with three dams from four families of each line (tolerant from  $390$ - and  $2400$ - $\mu\text{atm}$  P<sub>CO<sub>2</sub></sub> acclimation and sensitive from  $390$ - $\mu\text{atm}$  P<sub>CO<sub>2</sub></sub> acclimation only) crossed with one male from the tolerant family E6.

3B). Successful metamorphosis and thus tolerance correlated positively with PD I size because shell length was slightly larger in tolerant compared to sensitive families at  $2400 \mu\text{atm}$  ( $82 \pm 5 \mu\text{m}$  versus  $76 \pm 8 \mu\text{m}$ ) (two-way ANOVA: sensitive versus tolerant:  $F = 1.5$ ,  $P > 0.05$ ; P<sub>CO<sub>2</sub></sub>:  $F = 365.7$ ,  $P < 0.001$ ; sensitive versus tolerant  $\times$  P<sub>CO<sub>2</sub></sub>:  $F = 5.5$ ,  $P < 0.05$ ) (Fig. 4A).

Following settlement, juveniles of all families were transferred into a flow-through experimental system and raised for 1 year (2012–2013) at the respective P<sub>CO<sub>2</sub></sub> until the next spawning season (Table 1). During that time, no mortality was observed and F<sub>1</sub> juveniles from all families grew to shell sizes of about  $25 \text{ mm}$  within 1 year irrespective of P<sub>CO<sub>2</sub></sub> treatment or family type [ $390 \mu\text{atm}$ ,  $24.7 \pm 3.7 \text{ mm}$ ;  $1120 \mu\text{atm}$ ,  $26.0 \pm 2.7 \text{ mm}$ ; and  $2400 \mu\text{atm}$ ,  $24.8 \pm 3.0 \text{ mm}$  (tolerant families only)] (ANOVA:  $F = 0.216$ ,  $P > 0.05$ ).

**First F<sub>2</sub> generation (2013).**

Crosses of F<sub>1</sub> specimens were carried out to test (i) whether developmental acclimation of F<sub>1</sub> families conferred environment-specific benefits in relation to offspring P<sub>CO<sub>2</sub></sub> (that is, transgenerational plasticity) and (ii) whether tolerance has a genetic component that could be crossed into the genetic background of sensitive families (Fig. 3B). Maternal investment, measured as egg production of F<sub>1</sub> dams and egg diameter, did not change under elevated P<sub>CO<sub>2</sub></sub> levels (fig. S4). Fertilization success was not significantly affected by P<sub>CO<sub>2</sub></sub> levels, irrespective of whether it was assayed in tolerant or sensitive families. In crosses between tolerant F<sub>1</sub> parents (T $\times$ T), PD I size of F<sub>2</sub> larvae was similar at control P<sub>CO<sub>2</sub></sub>, irrespective of parental rearing history. However, at high P<sub>CO<sub>2</sub></sub>, PD I sizes were larger for offspring from tolerant F<sub>1</sub> families raised at elevated P<sub>CO<sub>2</sub></sub> compared to larvae from control P<sub>CO<sub>2</sub></sub>-treated parents (see parental P<sub>CO<sub>2</sub></sub>  $\times$  offspring P<sub>CO<sub>2</sub></sub> interaction effects in table S2). Shell size of offspring from F<sub>1</sub> dams selected



**Fig. 4. Larval performance of F<sub>1</sub> and F<sub>2</sub> animals exposed to elevated  $P_{CO_2}$ .** (A) PD I length declined with increasing  $P_{CO_2}$  but remained larger in tolerant F<sub>1</sub> families ( $n = 314$  to 620) and (B) partly recovered in tolerant (T×T) but not sensitive (S×T) F<sub>2</sub> offspring from high  $P_{CO_2}$ -treated parents ( $n = 902$  to 1314). (C) Larval daily shell growth at elevated  $P_{CO_2}$  was not affected in F<sub>1</sub> offspring (D) but decreased in F<sub>2</sub> larvae and further deteriorated with high- $CO_2$  exposure of parental animals. (E) Larval survival decreased at high  $P_{CO_2}$  but was higher in tolerant families compared to sensitive F<sub>1</sub> families. (F) Parental selection and long-term acclimation at high  $P_{CO_2}$  did not improve survival in the F<sub>2</sub> generation. Values are means  $\pm$  SD; numbers in brackets state the number of measured individuals per  $P_{CO_2}$  treatment.

and raised under 2400- $\mu\text{atm}$   $P_{CO_2}$  increased by 11.9  $\mu\text{m}$  or 17% compared to that of offspring of 390- $\mu\text{atm}$  acclimated F<sub>1</sub> dams (Fig. 4B and table S2). In contrast to the F<sub>1</sub> generation, F<sub>2</sub> larval growth rates in general were slower and particularly reduced at high  $P_{CO_2}$  and additionally declined when raised from F<sub>1</sub> exposed to elevated  $P_{CO_2}$  during long-term acclimation (Fig. 4D and table S2). Furthermore, larger PD I size of F<sub>2</sub> larvae at 2400- $\mu\text{atm}$   $P_{CO_2}$  had no positive effect on larval survival, in contrast to observations in the F<sub>1</sub> generation (Fig. 4F and table S2). Selection of tolerant phenotypes in the F<sub>1</sub> generation thus only had a positive transgenerational effect on PD I size but did not improve the mean population fitness of their F<sub>2</sub> offspring. Rather, offspring from mothers acclimated at control  $P_{CO_2}$  conditions (390  $\mu\text{atm}$ ) showed higher survival rates, indicating no positive effects of parental acclimation to high  $P_{CO_2}$  on offspring survival (table S2). In contrast to the results for crosses between tolerant families (T×T), crosses between tolerant mothers and sensitive fathers (S×T) resulted in similar PD I sizes of F<sub>2</sub> larvae, irrespective of parental  $P_{CO_2}$  treatment (Fig. 4B and table S2). Crosses between tolerant mothers and sensitive fathers showed an increased survival when compared to T×T crosses, especially at 1120- $\mu\text{atm}$   $P_{CO_2}$  (Fig. 4F and table S2).

#### Second F<sub>2</sub> generation (2014).

The observed responses for fecundity, PD I, larval growth rates, and survival were largely confirmed when generating the second F<sub>2</sub> genera-

tion from the same F<sub>1</sub> animals in the subsequent year (Fig. 3B and fig. S5). When kept for another year in the experimental system at their respective  $P_{CO_2}$  treatment (2013–2014), fecundity and egg sizes were affected neither by parental  $P_{CO_2}$  treatment nor by family type (sensitive/tolerant F<sub>1</sub> families; figs. S4, B and D, and S5A). F<sub>2</sub> offspring from high  $P_{CO_2}$ -selected parental F<sub>1</sub> animals showed a nonsignificant trend toward larger PD I size (fig. S5B and table S3). Shell growth rates were reduced in all high  $P_{CO_2}$ -treated larvae but were again affected neither by parental acclimation  $P_{CO_2}$  treatment nor by family type (fig. S5C). Similarly, survival of larvae was negatively affected by elevated  $P_{CO_2}$  and not improved by selection of tolerant F<sub>1</sub> parents (T×T) or the prolonged high- $P_{CO_2}$  acclimation of F<sub>1</sub> animals (fig. S5D and table S3).

## DISCUSSION

The high sensitivity of bivalve larvae to elevated  $P_{CO_2}$  (3, 4) suggests that selective pressures should be strong and populations should rapidly adapt to the prevailing local  $P_{CO_2}$  levels. However, evidence for this hypothesis is circumstantial (9, 12, 13). We used two different approaches to study the adaptation potential of mussels to ocean acidification. First, we performed a population comparison (PC) experiment to test for existing differences in tolerance to ocean acidification. Second, we assessed how rapidly tolerance can be acquired by selection of tolerant phenotypes or transgenerational plasticity in an MG experiment.

### Population comparison experiment (Baltic Sea versus North Sea)

In an experimental common garden approach, larval performance of North Sea mussels under low and elevated  $P_{CO_2}$  was compared to that of larvae from the Baltic Sea population. On the phenotypic level, adaptation to elevated  $P_{CO_2}$  in Baltic mussels was indicated by increased survival under elevated  $P_{CO_2}$  and higher capacity to maintain PD I formation rates compared to the more sensitive North Sea mussels. The experiment revealed that naturally and locally deviating ocean carbonate chemistry characteristics influence the responses of blue mussel populations to experimental ocean acidification, most likely reflecting local adaptation to prevailing environmental conditions on longer time scales (22–24). In general, larval calcification was strongly impaired by elevated  $P_{CO_2}$ ; this impairment was even more pronounced than reported at comparable  $P_{CO_2}$  for fully marine populations (5, 25). This reflects our choice of the highly selective environment in the brackish Baltic Sea, where conditions for calcification are less favorable (7). Calcification of bivalve larvae is not directly affected by  $P_{CO_2}$ , but is sensitive to lowered pH and availability of inorganic carbon ( $\text{HCO}_3^-$ ;  $C_T$ ) as a substrate for calcification, which correlates with seawater  $\Omega$  [calcium carbonate saturation state (5, 7, 26)]. The low alkalinity and thus low  $C_T$  concentrations of the Baltic Sea result in lowered carbon availability and  $\Omega$  and therefore synergistically enhance the negative effects of elevated  $P_{CO_2}$  on larval calcification (7). As a result of this intensified selection pressure, Baltic mytilid mussels have successfully adapted to adverse conditions for calcification.

Calcification of PD I coincides with the highest relative calcification rates of all bivalve life stages, which makes this ontogenetic stage most vulnerable to external carbonate system perturbations (5–7). The correlation of higher calcification rates and survival of tolerant Baltic mussels suggests that PD I calcification is mechanistically linked to survival and therefore directly to fitness. PD I sizes were similar in both populations under control  $P_{CO_2}$  when the external carbonate chemistry did not limit calcification; thus, growth and development were potentially limited by

**Table 1. Carbonate chemistry during the larval experiments and the long-term acclimation.** pH on total scale and  $C_T$  were measured ( $n = 163$ ), and  $A_T$ ,  $P_{CO_2}$ ,  $[CO_3^{2-}]$ , and  $\Omega_{Aragonite}$  were calculated using CO2SYS. NBS, National Bureau of Standards.

	Temperature (°C)	Salinity (g kg <sup>-1</sup> )	$P_{CO_2}$ treatment (μatm)	$C_T$ (μmol kg <sup>-1</sup> )	pH (total scale)	Measured pH (NBS scale)	$A_T$ (μmol kg <sup>-1</sup> )	$P_{CO_2}$ (μatm)	$[CO_3^{2-}]$ (μmol kg <sup>-1</sup> )	$\Omega_{Aragonite}$
F <sub>1</sub> larvae 2012	17.7 ± 0.1	15.5 ± 0.3	390	1802 ± 46	7.97 ± 0.02	8.17 ± 0.07	1884 ± 51	508 ± 14	76.6 ± 4.9	1.23 ± 0.08
			1120	1889 ± 78	7.65 ± 0.01	7.75 ± 0.06	1897 ± 76	1128 ± 82	38.6 ± 1.2	0.62 ± 0.02
			2400	1995 ± 50	7.39 ± 0.03	7.46 ± 0.05	1944 ± 51	2114 ± 108	22.6 ± 1.4	0.36 ± 0.02
F <sub>2</sub> larvae 2013	17.1 ± 0.2	16.0 ± 0.4	390	1916 ± 86	8.03 ± 0.20	8.16 ± 0.07	2026 ± 90	476 ± 181	99.8 ± 14.8	1.60 ± 0.24
F <sub>2</sub> larvae 2014	18.8 ± 0.1	16.0 ± 0.2	1120	2056 ± 54	7.64 ± 0.15	7.69 ± 0.01	2063 ± 87	1264 ± 167	44.1 ± 33.5	0.71 ± 0.53
			2400	2078 ± 27	7.43 ± 0.15	7.43 ± 0.02	2032 ± 28	2093 ± 379	28.7 ± 15.2	0.46 ± 0.24
			390	1875 ± 12	8.05 ± 0.03	8.19 ± 0.03	1989 ± 17	440 ± 24	99.6 ± 5.3	1.62 ± 0.09
Long-term acclimation (2012–2014)	11.4 ± 4.3	15.1 ± 2.1	2400	2039 ± 25	7.40 ± 0.07	7.55 ± 0.07	1991 ± 13	2160 ± 358	25.0 ± 4.3	0.41 ± 0.07
			1120	2108 ± 118	7.57 ± 0.05	7.71 ± 0.07	2068 ± 118	1381 ± 136	25.7 ± 6.7	0.40 ± 0.11
PC	15.5 ± 0.1	28.5 ± 0.2	2400	2258 ± 257	7.33 ± 0.06	7.44 ± 0.09	2146 ± 253	2515 ± 382	15.7 ± 4.6	0.24 ± 0.07
			390	2160 ± 14	8.02 ± 0.02	8.16 ± 0.01	2334 ± 14	462 ± 26	136.0 ± 6.0	2.14 ± 0.09
			2400	2411 ± 30	7.33 ± 0.01	7.46 ± 0.01	2357 ± 29	2588 ± 45	31.4 ± 0.6	0.49 ± 0.01

other physiological processes. Because PD I is formed before development of the larval feeding apparatus, a substantial fraction of the limited energy stored in the bivalve egg is needed for shell formation even under favorable carbonate system conditions (6, 27). Because calcification generates protons, which need to be excreted by means of an active transport process, disproportional up-regulation of shell formation could challenge the larval energy budget. A more efficient energy allocation into PD I formation may thus explain larger and maintained PD I size under elevated  $P_{CO_2}$  in the adapted Baltic compared to North Sea mussels.

### Three-year MG experiment (Baltic Sea)

Although common garden experiments offer a means to test for the existence of local adaptation, only experiments performed over multiple generations can give insights into the rate and mechanistic basis of the adaptation process. Earlier studies using oysters as model organisms revealed that parental preexposure to elevated  $P_{CO_2}$  resulted in faster growth and development of larvae under high  $P_{CO_2}$  when compared to larvae generated from parents that were acclimated to control conditions (9). Therefore, transgenerational phenotypic plasticity needs to be considered as an important factor that can modulate the response of bivalves to ocean acidification (28).

In our MG experiment performed with the Baltic Sea population, we observed a large variance in response to elevated  $P_{CO_2}$  among the  $CO_2$ -sensitive and  $CO_2$ -tolerant families. Our high-resolution environmental  $P_{CO_2}$  monitoring in the habitat of the population revealed rapidly fluctuating  $P_{CO_2}$ . Thus, different cohorts of larvae can be exposed to either high or low  $P_{CO_2}$  during the sensitive planktonic larval phase, indicating that not all individuals from the Kiel Fjord population were selected in a high- $P_{CO_2}$  environment. It is likely that this environmental heterogeneity selects for maintenance of variance of  $CO_2$  tolerance and genetic diversity in this population. The role of temporal heterogeneity of selection pressures for maintaining genetic diversity has historically been underestimated, although the scaling of phenotypic change with time strongly suggests that fluctuating selection pressures are the rule rather than the exception (29). Especially when generations overlap and selection pressures vary across life stages, fixation of alleles by selective sweeps becomes unlikely (30). The high sensitivity of larvae to elevated  $P_{CO_2}$  compared to adult mussels along with several yearly cohorts observed in the fjord fits this condition for maintaining genetic diversity. The low predictability of selective environmental  $P_{CO_2}$  levels in the Baltic population makes tracking of these fluctuations by heritable trait changes unlikely and should rather select for bet hedging (31) or mechanisms of

plasticity, particularly for transgenerational plasticity with the early life stages that are affected here (10, 32).

In both experiments (the MG and PC experiments), selection for tolerance to high  $P_{CO_2}$  correlated with a higher capacity to reach larger PD I sizes in the  $F_1$  generation. Calculated heritabilities for this trait (0.23 to 0.83) were within the range of values previously reported for mytilid larvae [0.09 to 0.9 (8, 33)]. The relatively larger PD I size of tolerant compared to sensitive families was also passed on to the  $F_2$  generation, thus showing a heritable component. This suggests that the ability to form the PD I shell even under adverse environmental conditions can be an important fitness trait. Although transgenerational plasticity could partly compensate the negative effects of elevated  $P_{CO_2}$  levels for PD I formation rates within one generation (Fig. 4B), the absence of an effect on  $F_2$  survival implies that PD I size alone cannot be used as a trait for reliable modeling of the evolutionary response of population mean fitness (9). Similar results were obtained for oysters when selection of larvae under 856- $\mu\text{atm}$   $P_{CO_2}$  did not improve the survival of their  $F_2$  offspring under the same  $P_{CO_2}$  treatment (16). Increased performance observed in marine organisms under moderately elevated  $P_{CO_2}$  can probably be attributed to transgenerational phenotypic plasticity (TGP). TGP has been suggested to function as a short-term buffering mechanism to alleviate the effects of adverse environments before genetic adaptation can fill the fitness deficit. TGP has been shown to even persist over several generations in a range of species (10, 15, 34, 35). TGP can manifest itself in altered animal performance with beneficial effects on growth and fecundity (34–37) or, in the case of bivalves, via modification of shell formation processes (38). More specifically, TGP can modulate, for example, respiratory capacity (aerobic scope) by acting upon mitochondrial properties. TGP thereby enables animals to adjust crucial physiological processes to the changed environment (35–37, 39). Maternal effects can play a central role in passing TGP from one generation to the next (35, 37). The rapid recovery of the PD I size of offspring from high  $P_{CO_2}$ -treated dams under acidified conditions (Fig. 4A) could result from such maternally driven TGP as well. In contrast, the absence of a positive effect on  $F_2$  survival suggests that  $F_1$  larval fitness is dependent on specific combinations of genotypes and nonheritable components.

Although significant adaptive responses may not necessarily be detectable on the whole-organism level within the three generations investigated in this study, they likely have contributed to the higher fitness of the Kiel Fjord population compared to North Sea mussels over longer time scales. Although high- $CO_2$  fluctuations in this habitat have increased only within recent decades as a result of eutrophication, adverse conditions for calcification due to lower alkalinity compared to the North Sea have prevailed for thousands of years (20, 40). The high mortality of bivalves during the sensitive larval phase and the very high effective population size of mussels in the Baltic Sea (41) should have efficiently selected for beneficial mutations that increased population fitness. In support of this view, changed allele frequencies in response to elevated  $P_{CO_2}$  have been observed in sea urchin larvae within only 7 days of exposure (24). In our study, selection of tolerant  $F_1$  specimens did not improve  $F_2$  survival, which corresponds to findings obtained with oysters (16). However, selective breeding of high-yield oysters for aquaculture purposes resulted in significantly improved ocean acidification tolerance as a side effect within just four generations (10, 17). The absence of a beneficial effect of selection in our study could be due to the small number of individuals used for the genetic crosses, which reduced the standing genetic variation present in the  $F_1$  generation. However, a large standing variation is needed as a prerequisite for selection (24). Consequently, future experiments would need to use a larger number of indi-

viduals or families to lower the risk of detrimental genetic drift to more closely resemble the genetic variability present in populations, enabling rapid adaptation (42). This is particularly important for coastal habitats such as Kiel Fjord, which are characterized by large abiotic variability that could lead to high genetic variation within a single population.

In conclusion, several lines of evidence suggest a potential of *Mytilus* populations to adapt to elevated  $CO_2$ . This conclusion is supported by (i) the different sensitivity of Baltic Sea and North Sea populations in response to a natural  $P_{CO_2}$  gradient and (ii) a heritable component of calcification performance in early larval development observed in the MG experiment. Mussel larvae from the Baltic were characterized by higher  $CO_2$  tolerance that correlated with higher ability to form the PD I shell under  $CO_2$  stress. In concurrence with these data, our MG experiment revealed that selection for settlement in high- $P_{CO_2}$  environments correlated with retention of PD I formation capabilities in  $F_1$  animals. However, selection of tolerant  $F_1$  phenotypes and long-term acclimation of  $F_1$  specimens in our MG study did not significantly improve  $F_2$  offspring survival. Consequently, prediction of adaptation potential based on short-term experiments and single traits within a population and generation appears to be highly speculative. Future experiments need to be performed over multiple generations to obtain a detailed understanding of the rate of adaptation and the underlying mechanisms to predict whether adaptation will enable marine organisms to overcome the constraints of ocean acidification.

## MATERIALS AND METHODS

Kiel Fjord seawater  $P_{CO_2}$  was continuously monitored using a HydroC  $CO_2$  sensor [Kongsberg Maritime AS (43)] mounted on a floating platform in about 1-m water depths. Abundance of settled bivalve larvae was assessed weekly on 5 cm  $\times$  5 cm manually roughed, replicated polyvinyl chloride panels ( $n = 4$ ) suspended in the fjord in about 50-cm water depth.

For the PC experiment, *M. edulis* from Kiel Fjord (Baltic Sea) were transferred to List/Sylt (North Sea) and suspended in net cages along with North Sea specimens to acclimate to North Sea conditions. Acclimation lasted from December 2013 to April 2014 when all specimens were transferred back to Kiel and used for spawning the next day.

For the MG experiment, adult *M. edulis* were collected in Kiel Fjord in 2012 and kept overnight in a flow-through seawater setup under control conditions. Spawning was induced by a moderate heat shock (5°C) using heaters. Parental ( $F_0$ ) animals (eight dams, A to H; eight sires, 1 to 8) were crossed pairwise in a reduced North Carolina I cross under control conditions to generate 16 full-sib families within four half-sib groups. Embryos were transferred into three experimental  $P_{CO_2}$  levels (390, 1120, or 2400  $\mu\text{atm}$ ). All families with successful settlement at 2400- $\mu\text{atm}$   $P_{CO_2}$  were considered as tolerant (5 of 16), and the remaining families (11 of 16) were termed “sensitive.” Juveniles were transferred to a flow-through setup under constant  $P_{CO_2}$  until the next spawning season. The setup consisted of a header tank, which steadily supplied the experimental aquaria with seawater from Kiel Fjord. A *Rhodomonas* suspension was pumped into the header using a peristaltic pump and provided food to the experimental aquaria. Each aquarium was separately aerated with pressurized air with a  $P_{CO_2}$  of either 390, 1120, or 2400  $\mu\text{atm}$ . Animals grew to average sizes of about 25 mm and sexual maturity within 1 year.

In 2013, individual crosses of  $F_1$  specimens were carried out within tolerant families (dams: A2, B1, C4  $\times$  sires: D3) and between tolerant and sensitive families (tolerant dams: A2, B1, C4  $\times$  sensitive sires: E6; sensitive dams: F5, G7, H8  $\times$  tolerant sires: D3). The sex bias in mussels

(44) resulted in only 4 of the 16 families with male offspring and 12 exclusively female families, which reduced the number of potential crosses.  $P_{CO_2}$  during fertilization and the larval phase corresponded to tolerant sires' acclimation  $P_{CO_2}$  (exception tolerant dams  $\times$  sensitive sires), and larvae were exposed to all  $P_{CO_2}$  levels in a fully crossed experimental design. In 2014, the response of 390- or 2400- $\mu\text{atm}$  acclimated tolerant families and 390- $\mu\text{atm}$  acclimated sensitive families was compared. For this purpose, eggs from three individuals from each of four tolerant dam families (A2, B1, B2, and C4) acclimated to 390- or 2400- $\mu\text{atm}$   $P_{CO_2}$  or females from the sensitive dam families (F5, G7, H7, and H8) acclimated to 390- $\mu\text{atm}$   $P_{CO_2}$  were pooled in equal numbers. Eggs were fertilized at 390- or 2400- $\mu\text{atm}$   $P_{CO_2}$ , corresponding to the treatment  $P_{CO_2}$  during the larval phase, with the sperm of one sire of family E6 acclimated to 1120  $\mu\text{atm}$ .

Larval experiments were performed in a constant-temperature room at 19°C (population experiment, 16°C) at the Helmholtz Centre for Ocean Research Kiel. For fertilization, excess sperm was added, which does not allow testing for effects of  $P_{CO_2}$  on fertilization success. After assessing fertilization success in replicated measurements, embryos were transferred into the experimental units at an initial density of 10 larvae  $\text{ml}^{-1}$ . Experimental units were filled with 0.2- $\mu\text{m}$  filtered seawater from Kiel Fjord. Weekly, 60% of the water volume was exchanged. In 2012 and 2013, larvae were fed daily with *Isochrysis* and *Rhodomonas* (days 7 to 21) or with *Rhodomonas* only (2014) cultured in F/2 or Provasoli enriched seawater (PES) medium, respectively. Larval survival and shell growth were assessed on days 2, 7, 14, and 21 after fertilization. Samples were immediately fixed, or living specimens were collected individually using a pipette, counted and subsequently fixed in 4% paraformaldehyde, and buffered with 4 mM  $\text{NaHCO}_3$ . Pictures of eggs and larvae were taken under a stereomicroscope with a MicroPublisher 3.3 RTV camera and analyzed for shell length using the Image-Pro Plus 5.0.1 software.

Monitoring of  $\text{pH}_{\text{NBS}}$ , salinity, and temperature was carried out twice or once a week in the larval experiments and the juvenile long-term acclimation, respectively. Weekly, water samples were analyzed for  $C_T$  using an AIRICA CT analyzer (Marianda) or for  $A_T$  using a Metrohm 862 Compact Titrosampler. Sample pH was determined on NBS or total scale using seawater buffers. Carbonate system speciation was calculated using the CO2SYS program using published  $\text{KHSO}_4$ ,  $K_1$ , and  $K_2$  dissociation constants (45–47).

Statistical tests are based on replicate means and were analyzed by ANOVA using R. Genetic variation of size was based on individual measurements and was analyzed by generalized linear mixed models (GLMMs) containing animal and replicate tank as random effects. See the Supplementary Materials for a detailed method description.

## SUPPLEMENTARY MATERIALS

Supplementary material for this article is available at <http://advances.sciencemag.org/cgi/content/full/3/4/e1602411/DC1>

### Supplementary Materials and Methods

fig. S1. Geographic origin of the two tested populations from Kiel Fjord in the Baltic Sea and the island of Sylt in the North Sea.

fig. S2. Analysis of the  $P_{CO_2}$  data from Fig. 1B on  $P_{CO_2}$  levels experienced by larvae settling in July, August, and September in Kiel Fjord (54°19.8'N; 10°9.0'E).

fig. S3. Picture of an *M. edulis* larva, with an approximate shell length of 120  $\mu\text{m}$ , at the PD I stage 2 days after fertilization.

fig. S4. Egg diameter and fecundity of  $F_0$  and  $F_1$  dams.

fig. S5.  $F_1$  egg diameter and  $F_2$  larval performance in 2014.

table S1. Statistical analyses of population experiment.

table S2. Main effect contrasts from Bayesian GLMMs.

table S3. Statistical analyses of the transgenerational experiment in 2014.

References (48–54)

## REFERENCES AND NOTES

- K. J. Kroeker, R. L. Kordas, R. Crim, I. E. Hendriks, L. Ramajo, G. S. Singh, C. M. Duarte, J.-P. Gattuso, Impacts of ocean acidification on marine organisms: Quantifying sensitivities and interaction with warming. *Glob. Chang. Biol.* **19**, 1884–1896 (2013).
- H. Kurihara, T. Asai, S. Kato, A. Ishimatsu, Effects of elevated  $p\text{CO}_2$  on early development in the mussel *Mytilus galloprovincialis*. *Aquat. Biol.* **4**, 225–223 (2008).
- S. C. Talmage, C. J. Gobler, Effects of past, present, and future ocean carbon dioxide concentrations on the growth and survival of larval shellfish. *Proc. Natl. Acad. Sci. U.S.A.* **107**, 17246–17251 (2010).
- F. Gazeau, L. M. Parker, S. Cormeau, J.-P. Gattuso, W. A. O'Connor, S. Martin, H.-O. Pörtner, P. M. Ross, Impacts of ocean acidification on marine shelled molluscs. *Mar. Biol.* **160**, 2207–2245 (2013).
- G.G. Waldbusser, B. Hales, C. J. Langdon, B. A. Haley, P. Schrader, E. L. Brunner, M. W. Gray, C. A. Miller, I. Gimenez, Saturation-state sensitivity of marine bivalve larvae to ocean acidification. *Nat. Clim. Change* **5**, 273–280 (2014).
- G. G. Waldbusser, E. L. Brunner, B. A. Haley, B. Hales, C. J. Langdon, F. G. Prahl, A developmental and energetic basis linking larval oyster shell formation to acidification sensitivity. *Geophys. Res. Lett.* **40**, 2171–2176 (2013).
- J. Thomsen, K. Hayner, K. M. Wegner, F. Melzner, Impact of seawater carbonate chemistry on the calcification of marine bivalves. *Biogeosciences* **12**, 4209–4220 (2015).
- P. Enderlein, M. Wahl Dominance of blue mussels versus consumer-mediated enhancement of benthic diversity. *J. Sea Res.* **51**, 145–155 (2004).
- J. M. Sunday, R. N. Crim, C. D. G. Harley, M. W. Hart, Quantifying rates of evolutionary adaptation in response to ocean acidification. *PLOS ONE* **6**, e22881 (2011).
- L. M. Parker, P. M. Ross, W. A. O'Connor, L. Borysko, D. A. Raftos, H.-O. Pörtner, Adult exposure influences offspring response to ocean acidification in oysters. *Glob. Chang. Biol.* **18**, 82–92 (2012).
- J. M. Sunday, P. Calosi, S. Dupont, P. L. Munday, J. H. Stillman, T. B. H. Reusch, Evolution in an acidifying ocean. *Trends Ecol. Evol.* **29**, 117–125 (2014).
- C. Duarte, J. M. Navarro, K. Acuña, R. Torres, P. H. Manríquez, M. A. Lardies, C. A. Vargas, N. A. Lagos, V. Aguilera, Intraspecific variability in the response of the edible mussel *Mytilus chilensis* (Hupe) to ocean acidification. *Estuaries Coasts* **38**, 590–598 (2015).
- G.G. Waldbusser, B. Hales, C. J. Langdon, B. A. Haley, P. Schrader, E. L. Brunner, M. W. Gray, C. A. Miller, I. Gimenez, G. Hutchinson, Ocean acidification has multiple modes of action in bivalve larvae. *PLOS ONE* **10**, e0128376 (2015).
- T. B. H. Reusch, Climate change in the oceans: Evolutionary versus phenotypically plastic responses of marine animals and plants. *Evol. Appl.* **7**, 104–122 (2014).
- G.M. Miller, S.-A. Watson, J. M. Donelson, M. I. McCormick, P. L. Munday, Parental environment mediates impacts of increased carbon dioxide on a coral reef fish. *Nat. Clim. Change* **2**, 858–861 (2012).
- L. M. Parker, W. A. O'Connor, D. A. Raftos, H.-O. Pörtner, P. M. Ross, Persistence of positive carryover effects in the oyster, *Saccostrea glomerata*, following transgenerational exposure to ocean acidification. *PLOS ONE* **10**, e0132276 (2015).
- J. A. Nell, B. Perkins, Evaluation of progeny of fourth generation Sydney rock oyster *Saccostrea glomerata* (Gould, 1850) breeding lines. *Aquacult. Res.* **36**, 753–757 (2005).
- J. Thomsen, M. A. Gutowska, J. Saphörster, A. Heinemann, K. Trübenbach, J. Fietzke, C. Hiebenthal, A. Eisenhauer, A. Körtzinger, M. Wahl, F. Melzner, Calcifying invertebrates succeed in a naturally  $\text{CO}_2$ -rich coastal habitat but are threatened by high levels of future acidification. *Biogeosciences* **7**, 3879–3891 (2010).
- C. Pansch, I. Schaub, J. Havenhand, M. Wahl, Habitat traits and food availability determine the response of marine invertebrates to ocean acidification. *Glob. Chang. Biol.* **20**, 765–777 (2014).
- J. Beldowski, A. Löffler, B. Schneider, L. Joensuu, Distribution and biogeochemical control of total  $\text{CO}_2$  and total alkalinity in the Baltic Sea. *J. Mar. Syst.* **81**, 252–259 (2010).
- T. R. Waller, Functional morphology and development of veliger larvae of the European oyster, *Ostrea edulis* Linné. *Smithson. Contrib. Zool.* **328**, 1–70 (1981).
- P. Calosi, S. P. S. Rastrick, C. Lombardi, H. J. de Guzman, L. Davidson, M. Jahnke, A. Giangrande, J. D. Hardege, A. Schulze, J. I. Spicer, M.-C. Gambi, Adaptation and acclimatization to ocean acidification in marine ectotherms: An in situ transplant experiment with polychaetes at a shallow  $\text{CO}_2$  vent system. *Philos. Trans. R. Soc. B* **368**, 20120444 (2013).
- M. W. Kelly, J. L. Padilla-Gamiño, G. E. Hofmann, Natural variation and the capacity to adapt to ocean acidification in the keystone sea urchin *Strongylocentrotus purpuratus*. *Glob. Chang. Biol.* **19**, 2536–2546 (2013).
- S. R. Palumbi, D. J. Barshis, N. Tylor-Knowles, R. A. Bay, Mechanisms of reef coral resistance to future climate change. *Science* **344**, 895–898 (2014).
- R. K. Bechmann, I. C. Taban, S. Westerlund, B. F. Godal, M. Anberg, S. Vingen, A. Ingvarsdottir, T. Baussant, Effects of ocean acidification on early life stages of shrimp

- (*Pandalus borealis*) and mussel (*Mytilus edulis*). *J. Toxicol. Environ. Health A* **74**, 424–438 (2011).
26. L. T. Bach, Reconsidering the role of carbonate ion concentration in calcification by marine organisms. *Biogeosciences* **12**, 4939–4951 (2015).
  27. A. Lucas, C. Rangel, Detection of the first larval feeding in *Crassostrea gigas*, using the epifluorescence microscope. *Aquaculture* **30**, 369–374 (1983).
  28. S. A. Foo, M. Byrne, Acclimatization and adaptive capacity of marine species in a changing ocean. *Adv. Mar. Biol.* **74**, 69–116 (2016).
  29. P. W. Messer, S. P. Ellner, N. G. Hairston Jr., Can population genetics adapt to rapid evolution? *Trends Genet.* **32**, 408–418 (2016).
  30. S. Ellner, N. G. Hairston Jr., Role of overlapping generations in maintaining genetic variation in a fluctuating environment. *Am. Nat.* **143**, 403–417 (1994).
  31. L. N. S. Shama, Bet hedging in a warming ocean: Predictability of maternal environment shapes offspring size variation in marine sticklebacks. *Glob. Chang. Biol.* **21**, 4387–4400 (2015).
  32. C. K. Ghilambor, J. K. McKay, S. P. Carroll, D. N. Reznick, Adaptive versus non-adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Funct. Ecol.* **21**, 394–407 (2007).
  33. J. E. Toro, L. I. Paredes, Heritability estimates of larval shell length in the Chilean blue mussel *Mytilus chilensis*, under different food densities. *Aquat. Living Resour.* **9**, 347–350 (1996).
  34. A. Rodriguez-Romero, M. D. Jarrold, G. Massamba-N'Siala, J. I. Spicer, P. Calosi, Multi-generational responses of a marine polychaete to a rapid change in seawater pCO<sub>2</sub>. *Evol. Appl.* **9**, 1082–1095 (2015).
  35. L. N. S. Shama, A. Strobel, F. C. Mark, K. M. Wegner, Transgenerational plasticity in marine sticklebacks: Maternal effects mediate impacts of a warming ocean. *Funct. Ecol.* **28**, 1482–1493 (2014).
  36. P. Thor, S. Dupont, Transgenerational effects alleviate severe fecundity loss during ocean acidification in a ubiquitous planktonic copepod. *Glob. Chang. Biol.* **21**, 2261–2271 (2015).
  37. L. N. S. Shama, F. C. Mark, A. Strobel, A. Lokmer, U. John, K. M. Wegner, Transgenerational effects persist down the maternal line in marine sticklebacks: Gene expression matches physiology in a warming ocean. *Evol. Appl.* **9**, 1096–1111 (2016).
  38. S. C. Fitzner, M. Cusack, V. R. Phoenix, N. A. Kamenos, Ocean acidification reduces the crystallographic control in juvenile mussel shells. *J. Struct. Biol.* **188**, 39–45 (2014).
  39. J. M. Donelson, P. L. Munday, M. I. McCormick, C. R. Pitcher, Rapid transgenerational acclimation of a tropical reef fish to climate change. *Nat. Clim. Change* **2**, 30–32 (2012).
  40. J. Thomsen, I. Casties, C. Pansch, A. Körtzinger, F. Melzner, Food availability outweighs ocean acidification effects in juvenile *Mytilus edulis*: Laboratory and field experiments. *Glob. Chang. Biol.* **19**, 1017–1027 (2013).
  41. K. Johannesson, C. André, Life on the margin: Genetic isolation and diversity loss in a peripheral marine ecosystem, the Baltic Sea. *Mol. Ecol.* **15**, 2013–2029 (2006).
  42. R. D. H. Barrett, D. Schluter, Adaptation from standing genetic variation. *Trends Ecol. Evol.* **23**, 38–44 (2008).
  43. P. Fietzek, B. Fiedler, T. Steinhoff, A. Körtzinger, In situ quality assessment of a novel underwater pCO<sub>2</sub> sensor based on membrane equilibration and NDIR spectrometry. *J. Atmos. Oceanic Tech.* **31**, 181–196 (2014).
  44. E. Kenchington, B. MacDonald, L. Cao, D. Tsagkarakis, E. Zouros, Genetics of mother-dependent sex ratio in blue mussels (*Mytilus spp.*) and implications for doubly uniparental inheritance of mitochondrial DNA. *Genetics* **161**, 1579–1588 (2002).
  45. A. G. Dickson, F. J. Millero, A comparison of the equilibrium constants for the dissociation of carbonic acid in seawater media. *Deep-Sea Res.* **34**, 1733–1743 (1987).
  46. A. G. Dickson, Standard potential of the reaction: AgCl(s) + 1/2H<sub>2</sub>(g) = Ag(s) + HCl(aq), and the standard acidity constant of the ion HSO<sub>4</sub><sup>-</sup> in synthetic sea water from 273.15 to 318.15 K. *J. Chem. Thermodyn.* **22**, 113–127 (1990).
  47. C. Mehrbach, C. H. Culbertson, J. E. Hawley, R. M. Pytkowic, Measurement of apparent dissociation-constants of carbonic acid in seawater at atmospheric-pressure. *Limnol. Oceanogr.* **18**, 897–907 (1973).
  48. N. Rayssac, F. Pernet, O. Lacasse, R. Tremblay, Temperature effect on survival, growth and triacylglycerol content during the early ontogeny of *Mytilus edulis* and *M. trossulus*. *Mar. Ecol. Prog. Ser.* **417**, 183–191 (2010).
  49. M. Sprung, Physiological energetics of mussel larvae (*Mytilus edulis*). I. Shell growth and biomass. *Mar. Ecol. Prog. Ser.* **17**, 283–293 (1984).
  50. H. Stuckas, K. Stoof, H. Quesada, R. Tiedemann, Evolutionary implications of discordant clines across the Baltic *Mytilus* hybrid zone (*Mytilus edulis* and *Mytilus trossulus*). *Heredity* **103**, 146–156 (2009).
  51. S. S. Mathiesen, J. Thyrring, J. Hemmer-Hansen, J. Berge, A. Sukhotin, P. Leopold, M. Bekaert, M. K. Sejr, E. E. Nielsen, Genetic diversity and connectivity within *Mytilus* spp. in the subarctic and Arctic. *Evol. Appl.* **10**, 39–55 (2017).
  52. F. Melzner, J. Thomsen, W. Koeve, A. Oeschlies, M. A. Gutowska, H. W. Bange, H. P. Hansen, A. Körtzinger, Future ocean acidification will be amplified by hypoxia in coastal habitats. *Mar. Biol.* **160**, 1875–1888 (2013).
  53. A. G. Dickson, C. L. Sabine, J. R. Christian, *Guide to Best Practices for Ocean CO<sub>2</sub> Measurements* (PICES Special Publications, 2007), vol. 3, 191 pp.
  54. J. D. Hadfield, MCMC methods for multi-response generalized linear mixed models: The MCMCglmm R package. *J. Stat. Softw.* **33**, 1–22 (2010).

**Acknowledgments:** We would like to thank U. Panknin for algae culturing and monitoring of mussel cultures during the 3-year experimental period; A. Resteu and I. Podbielski for assisting during the larval experiment; and R. Asmus (AWI Sylt), V. Saderne, and C. Hiebenthal [Kiel Marine Organism Culture Centre (KIMOCC)] for providing data and supporting carbonate system monitoring. T. Reusch is acknowledged for his comments on an earlier version of the manuscript. **Funding:** This work was supported by the German Federal Ministry of Education and Research (BMBF)-funded project BIOACID II [subproject 3.7 (FKZ03F0655B) and subproject 3.4 (FKZ 03F0655A)] and is a contribution to the PACES (Polar regions and coasts in a changing earth system) research programme of the Alfred Wegener Institute, Helmholtz Centre for Polar and Marine Research. **Author contributions:** J.T., K.M.W., and F.M. conceived the study, analyzed the data, and wrote the manuscript with the help of all coauthors. J.T., L.S.S., K.H., H.S., and M.D. conducted the experiments. **Competing interests:** The authors declare that they have no competing interests. **Data and materials availability:** All data needed to evaluate the conclusions in the paper are present in the paper and/or the Supplementary Materials. Additional data related to this paper may be requested from the authors.

Submitted 30 September 2016

Accepted 28 February 2017

Published 26 April 2017

10.1126/sciadv.1602411

**Citation:** J. Thomsen, L. S. Stapp, K. Haynert, H. Schade, M. Danelli, G. Lannig, K. M. Wegner, F. Melzner, Naturally acidified habitat selects for ocean acidification-tolerant mussels. *Sci. Adv.* **3**, e1602411 (2017).



This article is published under a Creative Commons license. The specific license under which this article is published is noted on the first page.

For articles published under **CC BY** licenses, you may freely distribute, adapt, or reuse the article, including for commercial purposes, provided you give proper attribution.

For articles published under **CC BY-NC** licenses, you may distribute, adapt, or reuse the article for non-commercial purposes. Commercial use requires prior permission from the American Association for the Advancement of Science (AAAS). You may request permission by clicking [here](#).

***The following resources related to this article are available online at <http://advances.sciencemag.org>. (This information is current as of April 26, 2017):***

**Updated information and services**, including high-resolution figures, can be found in the online version of this article at:

<http://advances.sciencemag.org/content/3/4/e1602411.full>

**Supporting Online Material** can be found at:

<http://advances.sciencemag.org/content/suppl/2017/04/24/3.4.e1602411.DC1>

This article **cites 53 articles**, 4 of which you can access for free at:

<http://advances.sciencemag.org/content/3/4/e1602411#BIBL>

*Science Advances* (ISSN 2375-2548) publishes new articles weekly. The journal is published by the American Association for the Advancement of Science (AAAS), 1200 New York Avenue NW, Washington, DC 20005. Copyright is held by the Authors unless stated otherwise. AAAS is the exclusive licensee. The title *Science Advances* is a registered trademark of AAAS