

Bromosceptrin, an Alkaloid from the Marine Sponge *Agelas conifera*

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Sponges, *Agelas*, Bromopyrrole Alkaloids

Six dimeric bromopyrrole alkaloids (1–6) were isolated from a Florida Keys specimen of *Agelas conifera*. One of the constituents was identified as a new bromopyrrole metabolite, bromosceptrin (1). The structure of 1 was established from MS spectrometry and 1D and 2D NMR spectroscopy.

Introduction

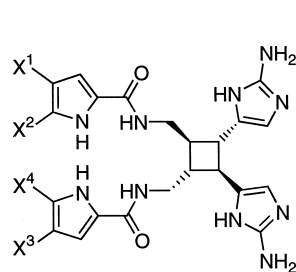
Bromopyrrole alkaloids are well known in marine sponges of the genus *Agelas* (Braekman *et al.*, 1992). In our search for bioactive substances from marine organisms, a series of brominated dimeric pyrrole alkaloids have been isolated from a specimen of the sponge *Agelas conifera* collected off the coast of the Florida Keys, Florida, USA. Examination of the dichloromethane/methanol extract of this sponge resulted in isolation of the known alkaloids sceptrin (2), dibromosceptrin (5), ageliferin (3), bromoageliferin (4), and dibromoageliferin (6) which have been previously isolated from *Agelas* sponges (Walker *et al.*, 1981; Kobayashi *et al.*, 1990; Keifer *et al.*, 1991) as well as of the new bromopyrrole alkaloid, bromosceptrin (1). In this communication we describe the isolation and structure elucidation of 1. Due to the isolation of

bromosceptrin (1) the family of the sceptrins is now completed.

Materials and Methods

The marine sponge *Agelas conifera* (Schmidt, 1870) (order Agelasida, family Agelasidae) employed in this study was collected in May 1998 at Elbow Reef by SCUBA diving (19 m depth) off the coast of the Florida Keys, Florida, USA. The growth form of the specimen is repent-ramose with volcano-shaped oscules, colour in life is brownish, consistency is tough, spongy, firm and almost incompressible. A voucher fragment of the sponge has been deposited under registration no. ZMA POR. 16866 in the Zoologisch Museum, Amsterdam, The Netherlands.

Samples of *Agelas conifera* were immediately frozen after collection and kept at -20°C until

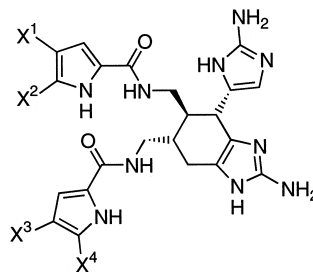


Sceptrin-Skeleton

Sceptrin (2): $X^1 = \text{Br}$, $X^2 = \text{H}$, $X^3 = \text{Br}$, $X^4 = \text{H}$

Bromosceptrin (1): $X^1 - X^3 = \text{Br}$, $X^4 = \text{H}$

Dibromosceptrin (5): $X^1 - X^4 = \text{Br}$



Ageliferin-Skeleton

Ageliferin (3): $X^1 = \text{Br}$, $X^2 = \text{H}$, $X^3 = \text{Br}$, $X^4 = \text{H}$

Bromoageliferin (4): $X^1 - X^3 = \text{Br}$, $X^4 = \text{H}$

Dibromoageliferin (6): $X^1 - X^4 = \text{Br}$

extraction. For bulk extraction followed by isolation of brominated secondary metabolites, freeze-dried sponge tissue (269 g \approx 1360 ml) was extracted 3 times in MeOH, twice in a 1:1-mixture of dichloromethane/MeOH, and once in dichloromethane at room temperature each. The organic extracts were combined and evaporated to dryness. The obtained crude extract was partitioned between *n*-hexane (3 \times 500 ml) and MeOH (150 ml). The remaining MeOH phase was concentrated and the residue (10.7 g) was purified by gel chromatography on Sephadex LH-20 (Pharmacia) using MeOH as mobile phase. A part of the fraction containing sceptrins and ageliferins (2.03 g, see Fig. 1) was finally purified by preparative RP₁₈ HPLC (conditions: 5 min A, 45 min 45% B; A: 10% MeCN/H₂O + 0.1% TFA; B: MeCN + 0.1% TFA). The following compound proportions approximating those found in the sponge tissue by HPLC quantification can be given: bromosceptrin (1) (0.05 mg/ml corresponds to 0.03% of dry weight), sceptrin (2) (1.02 mg/ml \approx 0.52%), dibromosceptrin (5) (0.13 mg/ml \approx 0.07%), ageliferin (3) (0.07 mg/ml \approx 0.04%), bromoageliferin (4)

(0.12 mg/ml \approx 0.06%), and dibromoageliferin (6) (0.09 mg/ml \approx 0.05%).

¹H NMR and ¹³C NMR spectra were recorded on a Bruker DRX600 NMR spectrometer. All NMR experiments were measured at 300 K. The 2D experiments (¹H,¹H-COSY, ¹H,¹³C-HSQC, ¹H,¹³C-HMBC, ¹H,¹⁵N-HSQC and ¹H,¹⁵N-HMBC) were carried out using standard parameters. Mass spectral analysis (HRFABMS) was performed on a JEOL JMS-700 sector-field mass spectrometer with 3-nitrobenzyl alcohol (NBA) as matrix or using a Fison VG Platform II for ESIMS. HPLC analysis and quantification was carried out as previously reported (Assmann *et al.*, 1999; Assmann *et al.*, 2000). IR (KBr) spectra were recorded on a Perkin Elmer 1600 Series FT-IR spectrometer. UV/VIS spectra were obtained using a Perkin Elmer UV/VIS spectrometer Lambda 16.

Results and Discussion

The compounds 1–6 could be isolated from a Florida Keys specimen of the marine sponge *Agelas conifera*. The brominated alkaloids sceptrin (2), dibromosceptrin (5), ageliferin (3), bromoageliferin (4), and dibromoageliferin (6) were identified by comparison of mass spectrometry and NMR data with those previously reported (Walker *et al.*, 1981 \rightarrow 2; Kobayashi *et al.*, 1990 \rightarrow 3, 4, 6; Keifer *et al.*, 1991 \rightarrow 2–6). The ESI mass spectrum (negative ion mode) of 1 showed prominent pseudomolecular ion peaks at *m/z* 697, 699, 701, 703 in the ratio 1:2:2:1, suggesting the presence of three bromine atoms.

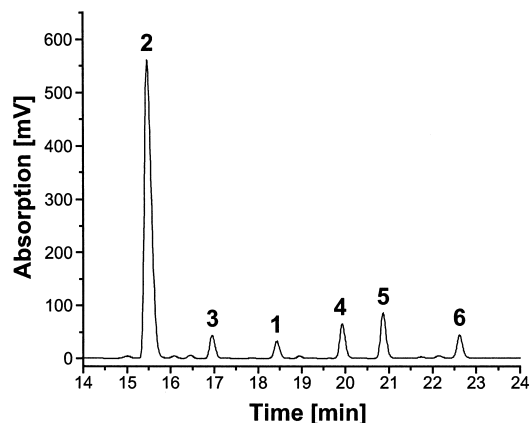
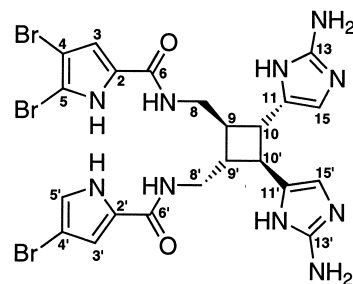


Fig. 1. HPLC profile of a fraction from the *n*-BuOH phase of *Agelas conifera* which has been purified by Sephadex LH-20 chromatography. This fraction contains only dimeric pyrrole alkaloids with the sceptrin and ageliferin skeleton. The retention times are: sceptrin (2) t_R = 15.46 min, ageliferin (3) t_R = 16.95 min, bromosceptrin (1) t_R = 18.43 min, bromoageliferin (4) t_R = 19.93 min, dibromosceptrin (5) t_R = 20.87 min, and dibromoageliferin (6) t_R = 22.62 min. HPLC conditions: column Kromasil RP₁₈, 4.6 \times 250 mm, 5 μ m; gradient 20 to 60% MeCN/H₂O + 0.1% TFA in 40 min; flow rate 1 ml/min.



From this data it cannot be distinguished between the two different skeleton of the dimeric bromopyrrole alkaloids (ageliferin or sceptrin). Due to the cyclisation of oroidin type compounds to the dimeric forms new aliphatic protons are generated which can be used as fingerprint region.

The ageliferin skeleton has three methine and one methylene group in the region between 2.0 and 3.0 ppm whereas the sceptrin skeleton has four methine signals. Another criterion is the olefinic region since the sceptrin skeleton shows one signal more (H15'). The molecular formula of **1** was established as $C_{22}H_{24}^{79}Br_3N_{10}O_2$ by HRFABMS (m/z 696.9634, $[M+H]^+$, Δ +4.1 mmu), which is in accordance with the 1H and ^{13}C NMR data of the new compound bromosceptrin (**1**) (see Table I). The absolute configuration of **1** was obtained by comparison of the CD spectral data ($c = 43 \mu\text{mol/l}$, MeOH, $[\theta]_{232} -1320$) with the values published in the literature (Walker *et al.*, 1981; Kobayashi *et al.*, 1990; Keifer *et al.*, 1991; Shen *et al.*, 1998). Due to the isolation of bromosceptrin (**1**), which contains 3 bromines, the family of the sceptrins is completed. The members differ in the degree of bromination. Debromosceptrin (none bromine, Shen *et al.*, 1998), monobromosceptrin (one bromine, Keifer *et al.*, 1991), sceptrin (**2**, two bromines, Walker *et al.*, 1981) and dibromosceptrin (**5**, four bromines, Keifer *et al.*, 1991) were already described in the literature.

Bromopyrrole alkaloids are known to be feeding deterrent against predatory reef fishes (Chanas *et al.*, 1996; Wilson *et al.*, 1999; Lindel *et al.*, 2000; Assmann *et al.*, 2000; Assmann *et al.*, 2001). The

dimeric compounds show a higher activity in comparison to the monomeric counterparts. Interestingly, the activity does not increase by the degree of bromination as known from the monomeric compounds (Assmann *et al.*, 2000). Only Sceptrin (**2**) appears in such concentration in the sponge tissue, which are above the required concentration of a single compound for feeding deterrence.

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Table I. 1H , ^{13}C and ^{15}N NMR chemical shifts (δ) of **1** in DMSO- d_6 .^a

Position		$\delta(^{13}C)/\delta(^{15}N)^b$	$\delta(^1H)^c$
1 (1')	NH	166 (161)	12.65 (11.78)
2 (2')	C	127.7 (126.4)	–
3 (3')	CH	112.5 (111.4)	6.88 (6.80)
4 (4')	C	97.7 (94.8)	–
5 (5')	C/CH	104.5 (121.2)	(6.98)
6 (6')	C	159.0 (159.7)	–
7 (7')	NH	105 (105)	8.20 (8.16)
8 (8')	CH ₂	40.7 (40.7)	3.39 (3.39)
9 (9')	CH	41.8 (41.8)	2.27 (2.27)
10 (10')	CH	37.1 (37.1)	2.94 (2.94)
11 (11')	C	126.8 (126.8)	–
12 (12')	NH	137 (137)	12.20 (12.20)
13 (13')	C	146.8 (146.8)	–
14 (14')	NH	134 (134)	11.73 (11.73)
15 (15')	CH	108.9 (108.9)	6.60 (6.60)
16 (16')	NH ₂	58 (58)	7.37 (7.37)

^a The structure of bromosceptrin (**1**) was already published (Assmann *et al.*, 2000), but without any analytical data.

^b ^{13}C chemical shifts are given in [ppm] and are referenced to the DMSO- d_6 signal (39.5 ppm). ^{15}N chemical shifts are given in [ppm] and are calibrated according to the Bruker frequency, which is set to 0 ppm for NH₃, the accuracy is about 1 to 2 ppm.

^c 1H chemical shifts are given in [ppm] and are referenced to the DMSO- d_6 signal (2.50 ppm).

- Assmann M., Lichte E., Pawlik J. R. and Köck M. (2000), Chemical defenses of the Caribbean sponges *Agelas wiedenmayeri* and *Agelas conifera*. *Mar. Ecol. Prog. Ser.* **207**, 255–262.
- Assmann M., Lichte E., van Soest R. W. M. and Köck M. (1999), New bromopyrrole alkaloid from the marine sponge *Agelas wiedenmayeri*. *Org. Lett.* **1**, 455–457.
- Assmann M., van Soest R. W. M. and Köck M. (2001), New antifeedant bromopyrrole alkaloid from the Caribbean sponge *Stylissa caribica*. *J. Nat. Prod.* **64**, 1345–1347.
- Braekman J. C., Daloz D., Stoller C. and van Soest R. W. M. (1992), Chemotaxonomy of *Agelas* (Porifera: Demospongiae). *Biochem. Syst. Ecol.* **20**, 417–431.
- Chanas B., Pawlik J. R., Lindel T. and Fenical W. (1996), Chemical defense of the Caribbean sponge *Agelas clathrodes* (Schmidt). *J. Exp. Mar. Biol. Ecol.* **208**, 185–196.
- Keifer P. A., Schwartz R. E., Koker M. E. S., Hughes R. G. Jr., Rittschof D. and Rinehart K. L. (1991), Bioactive bromopyrrole metabolites from the Caribbean sponge *Agelas conifera*. *J. Org. Chem.* **56**, 2965–2975, errata 5736, 6728.
- Kobayashi J., Tsuda M., Murayama T., Nakamura H., Ohizumi Y., Ishibashi M., Iwamura M., Ohta T. and Nozoe S. (1990), Ageliferins, potent actomyosin AT-Pase activator from the Okinawan marine sponge *Agelas* sp. *Tetrahedron* **46**, 5579–5586.
- Lindel T., Hoffmann H., Hochgürtel M. and Pawlik J. R. (2000), Structure-activity relationship of inhibition of fish feeding by sponge-derived and synthetic pyrrole-imidazole alkaloids. *J. Chem. Ecol.* **26**, 1477–1496.
- Shen X., Perry T. L., Dunbar C. D., Kelly-Borges M. and Hamann M. T. (1998), Debromosceptrin, an alkaloid from the Caribbean sponge *Agelas conifera*. *J. Nat. Prod.* **61**, 1302–1303.
- Walker R. P., Faulkner D. J., van Engen D. and Clardy J. (1981), Sceptrin, an antimicrobial agent from the sponge *Agelas sceptrum*. *J. Am. Chem. Soc.* **103**, 6772–6773.
- Wilson D. M., Puyana M., Fenical W. and Pawlik J. R. (1999), Chemical defense of the Caribbean reef sponge *Axinella corrugata* against predatory fishes. *J. Chem. Ecol.* **25**, 2811–2823.