

# Lessons learned by the harmonization between spectroscopic and thermal degradation methods for the analysis of microplastics

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# Why it is important to analyze MP?



- Mandatory for monitoring<sup>1</sup> of microplastics < 500  $\mu\text{m}$
- May contain additional chemicals like plasticizers
- Risk assessment needs particle numbers, particle shape and polymer<sup>2</sup>

1. GESAMP. Guidelines for the Monitoring and Assessment of Plastic Litter in the Ocean 2019.

2. Kögel T, et al. Sci Total Environ. 2020;709:136050. doi:10.1016/j.scitotenv.2019.136050.

# How can microplastics be analyzed?

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## FTIR spectroscopy

- Determines particle numbers
- Polymer type characterization via reference databases or other chemometric approaches
- Particles >10 µm can be measured in a rapid fashion.

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## Thermoanalysis-GC-MS

- Particle mass
- Using specific degradation products of the materials for quantification using signal to mass calibrations

# FTIR Imaging

Using the common Fourier-transform infrared (FTIR) spectroscopy

Allows the analysis of large filters (diameter usually 10 - 13 mm)

Applicable in transmission and reflection mode

Can be analyzed by automated approaches

Example: Sediment sample

Primpke et. al., *Anal. Methods*, 2017, 9, 1499–1511



# Harmonization by automated analysis

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Automatization of microplastic analysis  
based on FTIR imaging

- Data analysis independent from human bias via automated analysis
- Identification and Quantification of MP already within this process
- Time saving due to parallelization
- High comparability of results!

# Harmonization!



Water samples  
(surface etc.)

Cabernard et al., 2018, ES&T  
Lorenz et al., 2019, EP  
Tekman et al., 2020, ES&T  
Mintenig et al., 2020, WR  
Primpke et al. 2020, ABC

Treated waste  
water

Primpke et al., 2017, CHIUZ  
Primpke et al., 2019, Analytical Methods  
Mintenig et al., 2020, WR  
Primpke et al. 2020, ABC

siMPle

Primpke et al., 2020,  
Applied Spectroscopy

Automatization of microplastic analysis  
based on FTIR imaging

Sediments

Bergmann et al., 2017, ES&T  
Haave et al., 2019, MBP  
Lorenz et al., 2019, EP  
Mani et al., 2019, ES&T  
Abel et al. 2021, EP  
Primpke et al. 2020, ABC

Biota

In progress

(Arctic) Sea Ice

Peeken et al., 2018,  
Nature Communications

Snow

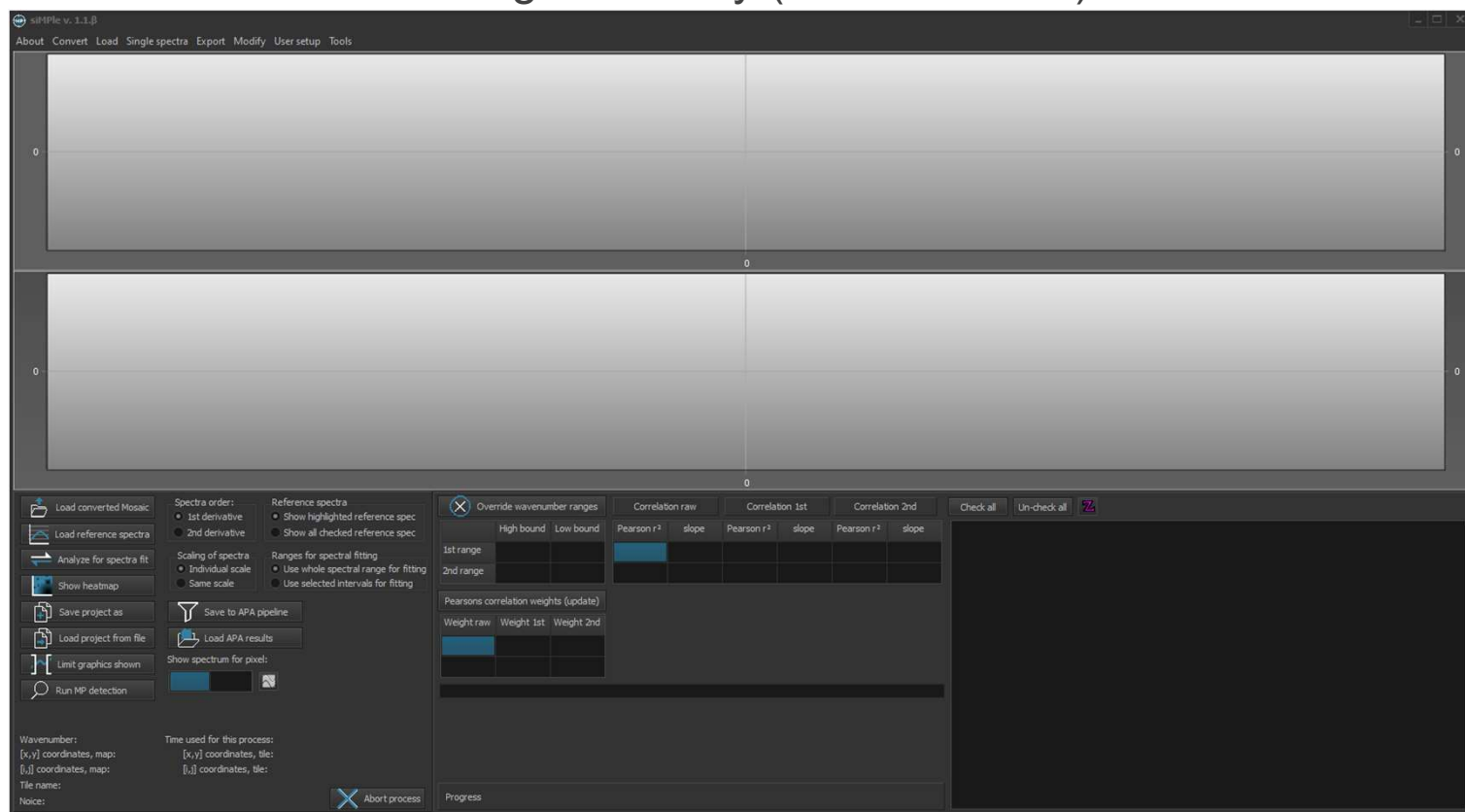
Bergmann et al., 2019,  
Science Advances



# Automatization via siMPle



- Systematic Identification of MicroPLastics in the Environment (siMPle)
- Software tool available by CC-BY-SA 4.0 on [www.simple-plastics.eu](http://www.simple-plastics.eu). In collaboration with Aalborg University (Jes Vollertsen)



See also: Pimpke, S., et al. 2020 Appl. Spectrosc. 74(9), 1127-1138. doi: 10.1177/0003702820917760

# siMPle for various IR systems

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Universal application of data analysis using the same database.

- Not limited to one manufacturer
  - Database is free of charge available
  - Software is free of charge available
  - Currently imports for Agilent, Bruker, DRS Daylight Solutions, Perkin Elmer and ThermoFischer Scientific
- Please contact us if your manufacturer is not in the list yet to find a solution

# siMPle for various IR systems

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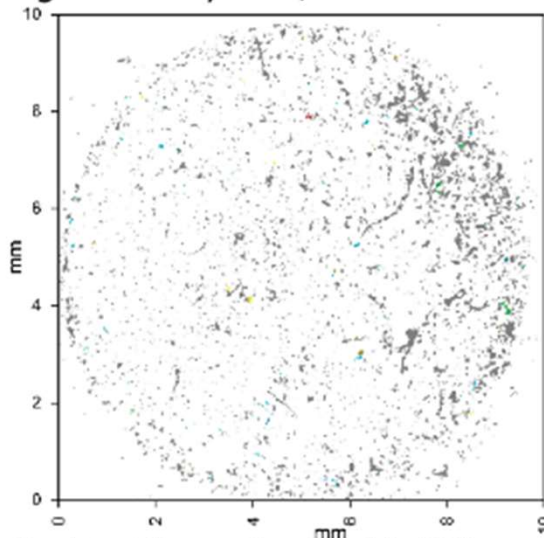
Testing the universal application of data analysis using the same database.

- Collaboration with Svenja Mintenig (Utrecht University), Richard Cross (Centre for Ecology and Hydrology/Wallingford), Alvise Vianello, Marta Simon and Jes Vollertsen (all Aalborg University)
  
- Dataset from Agilent, Bruker, Perkin Elmer and ThermoFischer Scientific systems

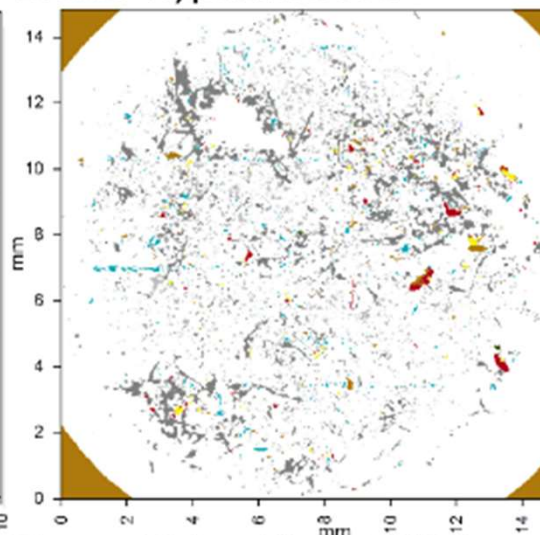
# siMPle for various IR systems



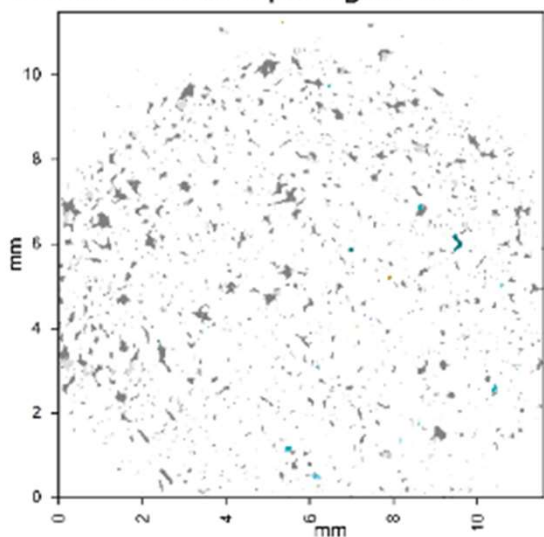
Agilent Cary 620/670



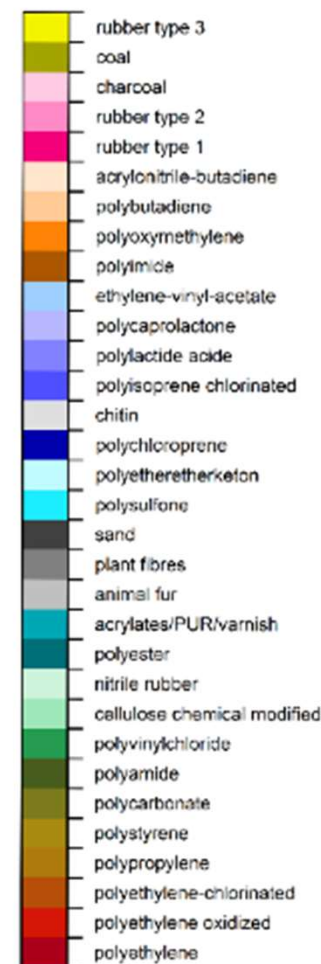
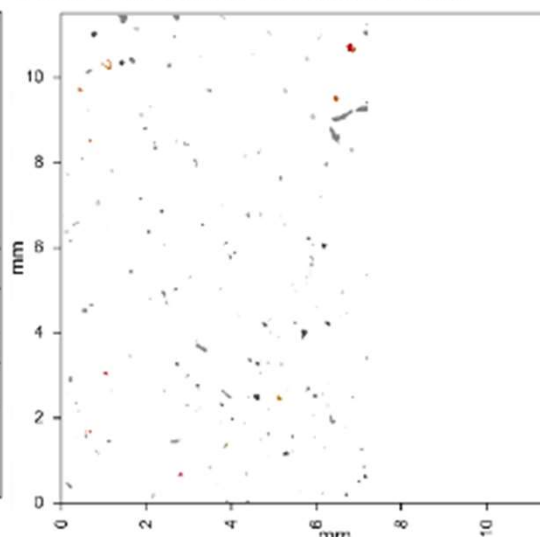
Bruker Hyperion 3000



Perkin Elmer Spotlight 400



ThermoFisher Nicolet iN10



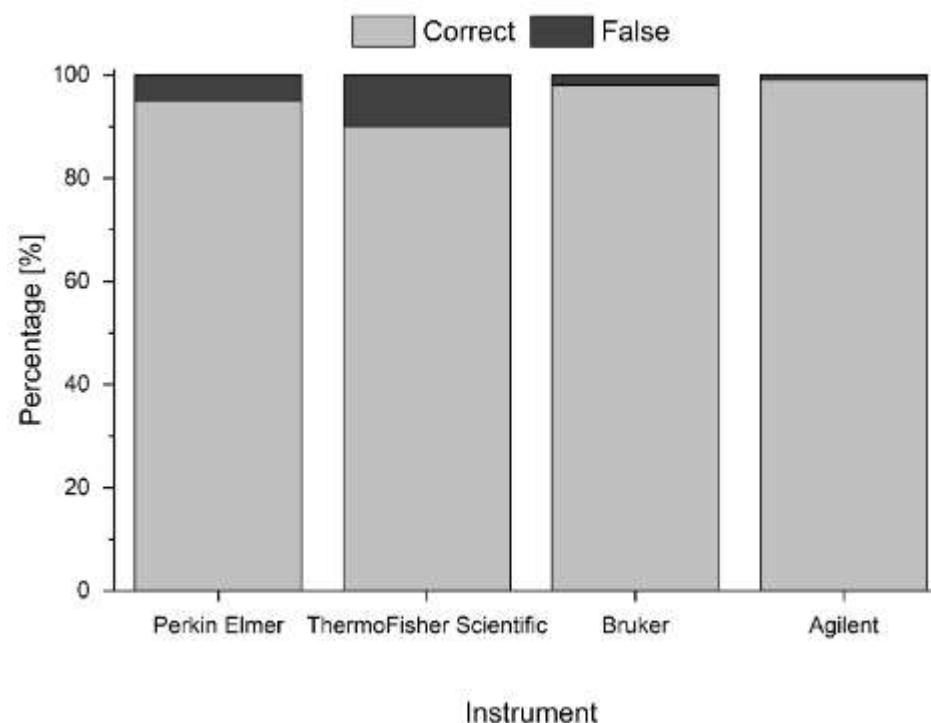
Graph: CC 4.0-BY-SA: Pimpke, S., *et al.* 2020 Appl. Spectrosc. 74(9), 1127-1138. doi: 10.1177/0003702820917760

# siMPle for various IR systems



Results of the intercomparison

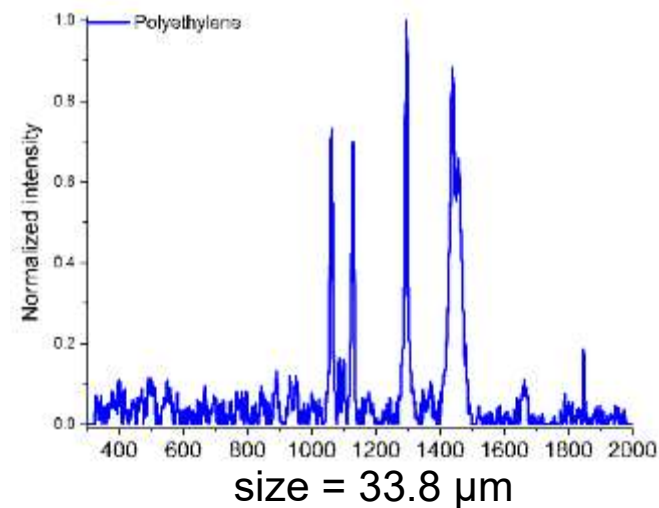
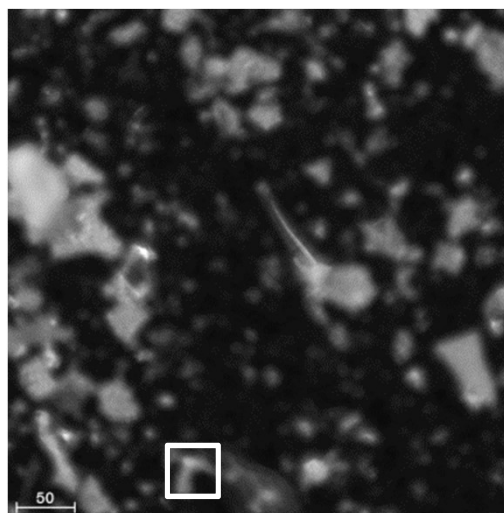
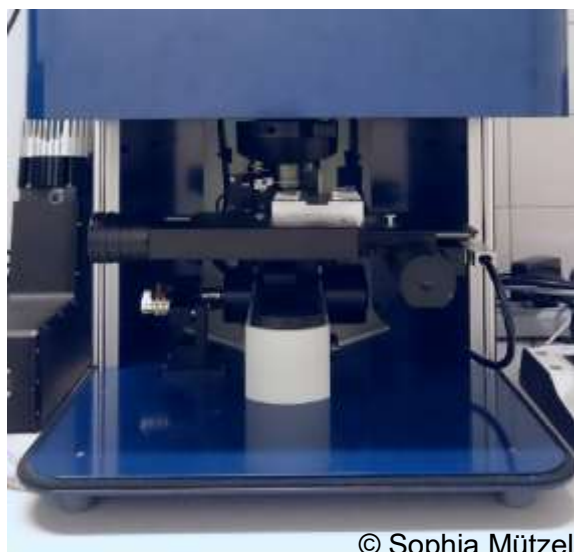
- All datasets yield identified particles
- Number of identified particles are dependent on the available pixel resolution
- All systems yielded mainly high ratios of correct assignments



# Raman for MP

Automated single particle exploring (ASPE) Raman

- Together with Livia Cabernard (AWI, now ETH Zürich), Claudia Lorenz (AWI, now Aalborg University), Lisa Roscher and Gunnar Gerdts (all AWI)
- Combines Particle analysis and counting with Raman analysis
- Each identified particle is individually targeted by Raman



1. L. Cabernard, et al. , Environmental Science & Technology 2018, 52, 13279-13288.

# Comparison of FTIR and Raman



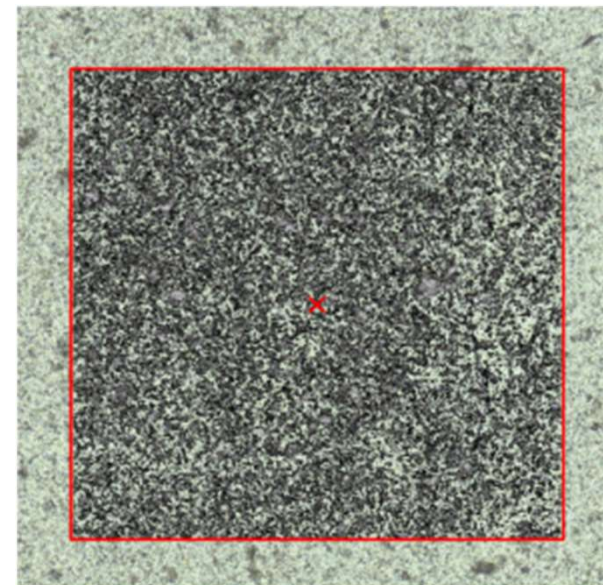
FTIR imaging and automated single particle exploring (ASPE) Raman microscopy

Seven environmental samples

Using gold coated polycarbonate filters

Measurement of FTIR (full image) and Raman (red square)

Only FTIR and Raman within the red square were compared

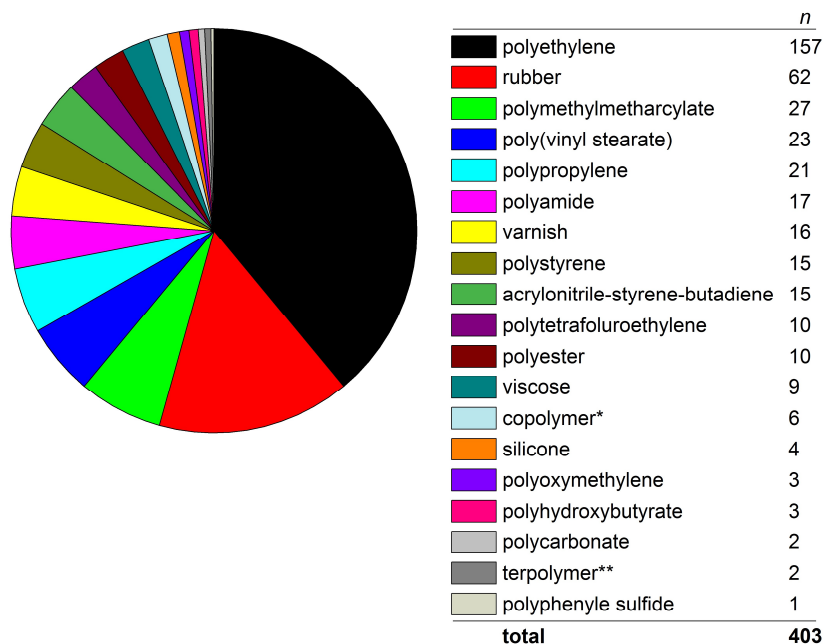


1. L. Cabernard, et al. , Environmental Science & Technology 2018, 52, 13279-13288.

# Method intercalibration



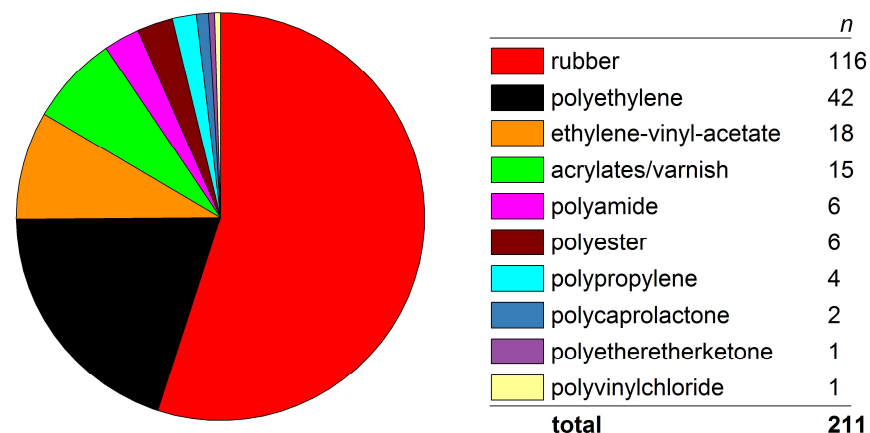
SPE- $\mu$ -Raman



\*styrene/allyl alcohol & vinyl chloride/vinylacetate

\*\*vinylchloride/vinylacetat/maleic acid

FTIR imaging

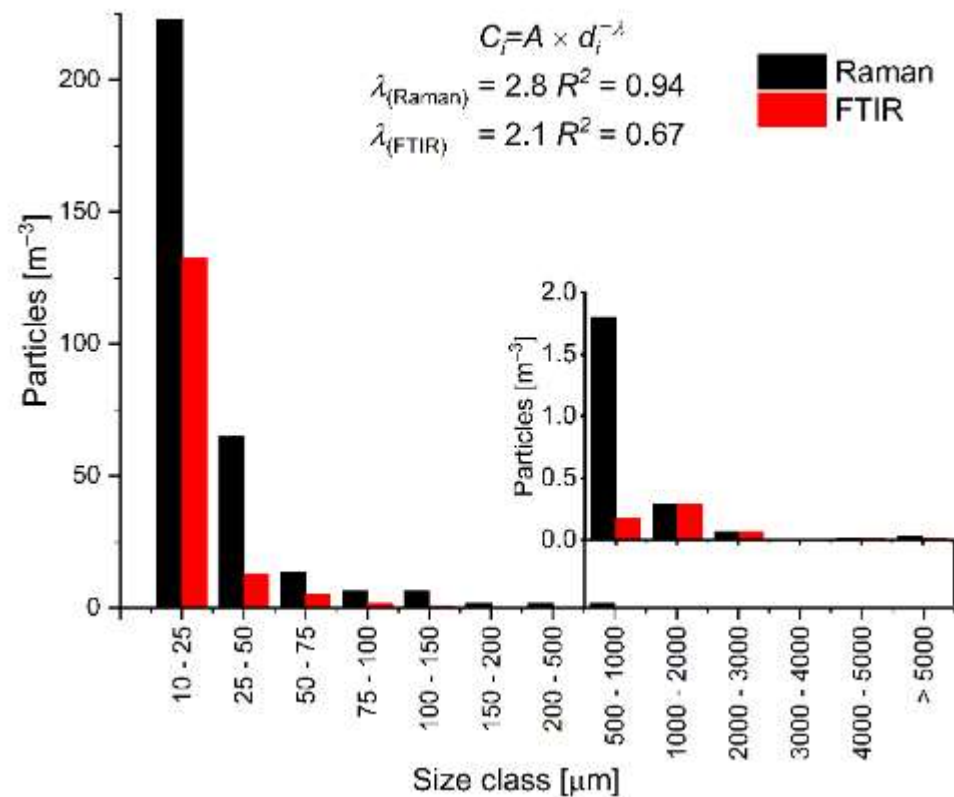


Raman: average 43 hours per sample    FTIR: 8 hours per sample

1. L. Cabernard, et al. , Environmental Science & Technology 2018, 52, 13279-13288.



# Method intercalibration

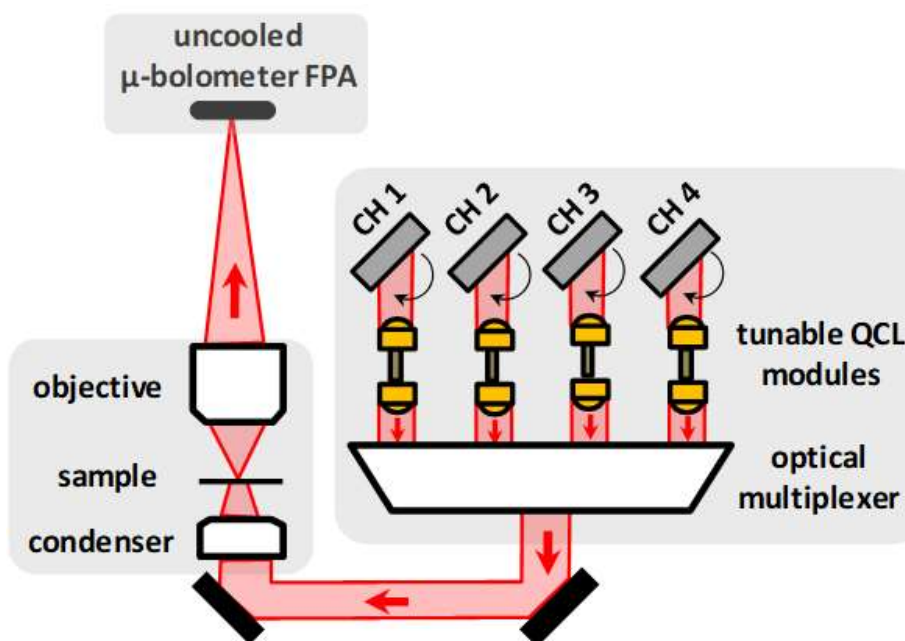


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1. Reprinted with permission from L. Cabernard, et al. , Environmental Science & Technology 2018, 52, 13279-13288. Copyright 2018 American Chemical Society.

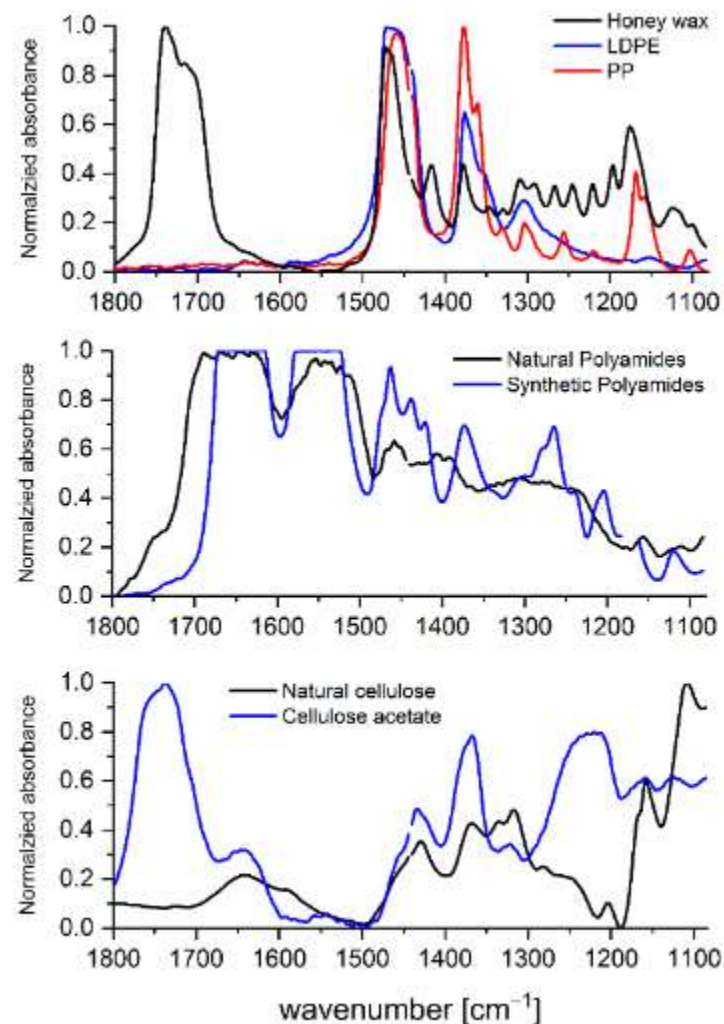
# Quantum Cascade Laser Imaging

- DRS Daylight SperoQT: Setup similar to an infra red microscope
- Infra red source is a tunable laser
- No liquid nitrogen required
- Speed: 1 minute for a  $2 \times 2$  mm field of view with a wavenumber range of  $1800 - 950 \text{ cm}^{-1}$
- Resolution:  $4.2 \text{ }\mu\text{m}$  per pixel in the field of view



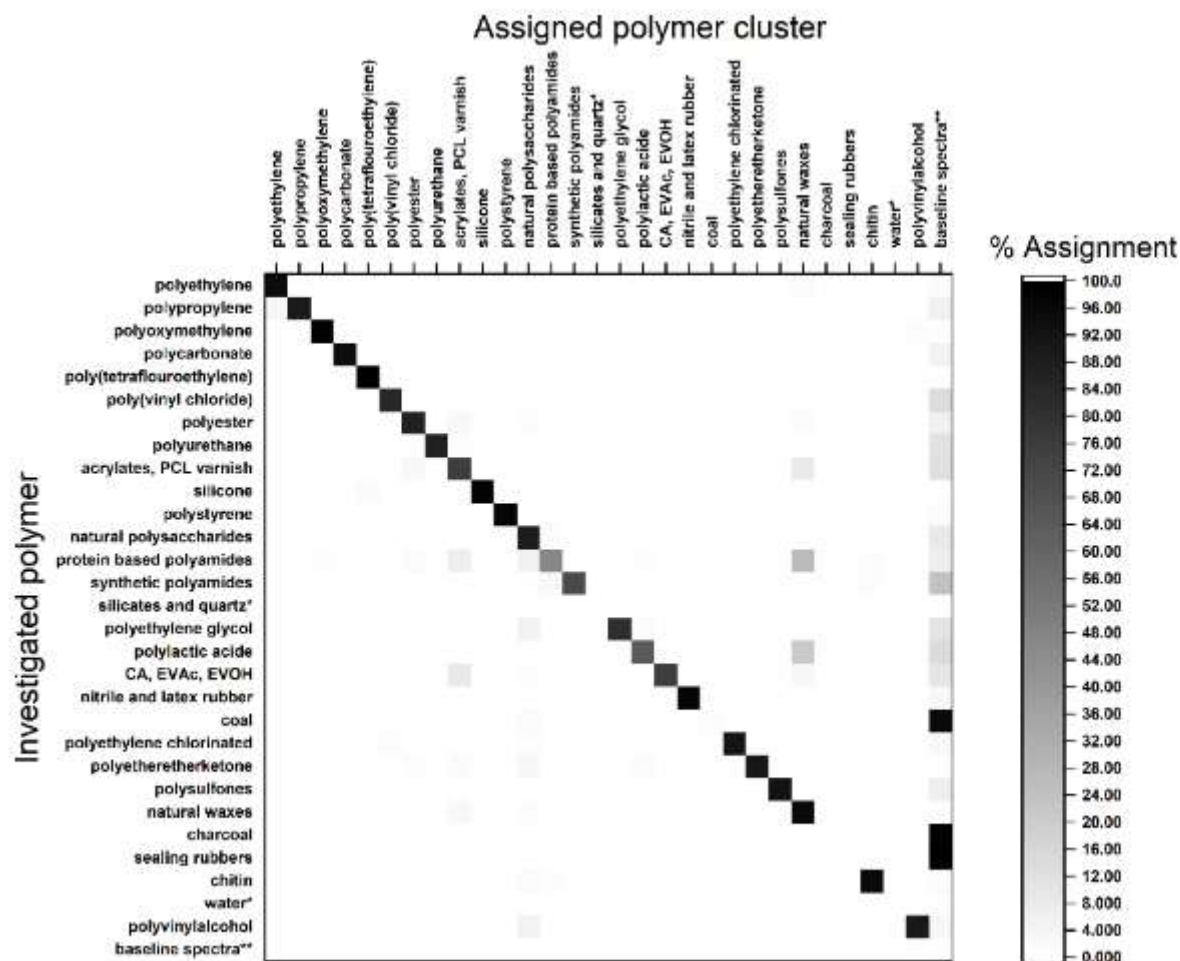
# QCL based spectra on Anodisc

- Anodisc is one of the few suitable filters for IR
- Inexpensive, but limited in wavenumber range ( $>1250\text{ cm}^{-1}$ )
- Using a QCL, measurements are possible until  $>1084\text{ cm}^{-1}$
- Separation of natural and anthropogenic materials possible



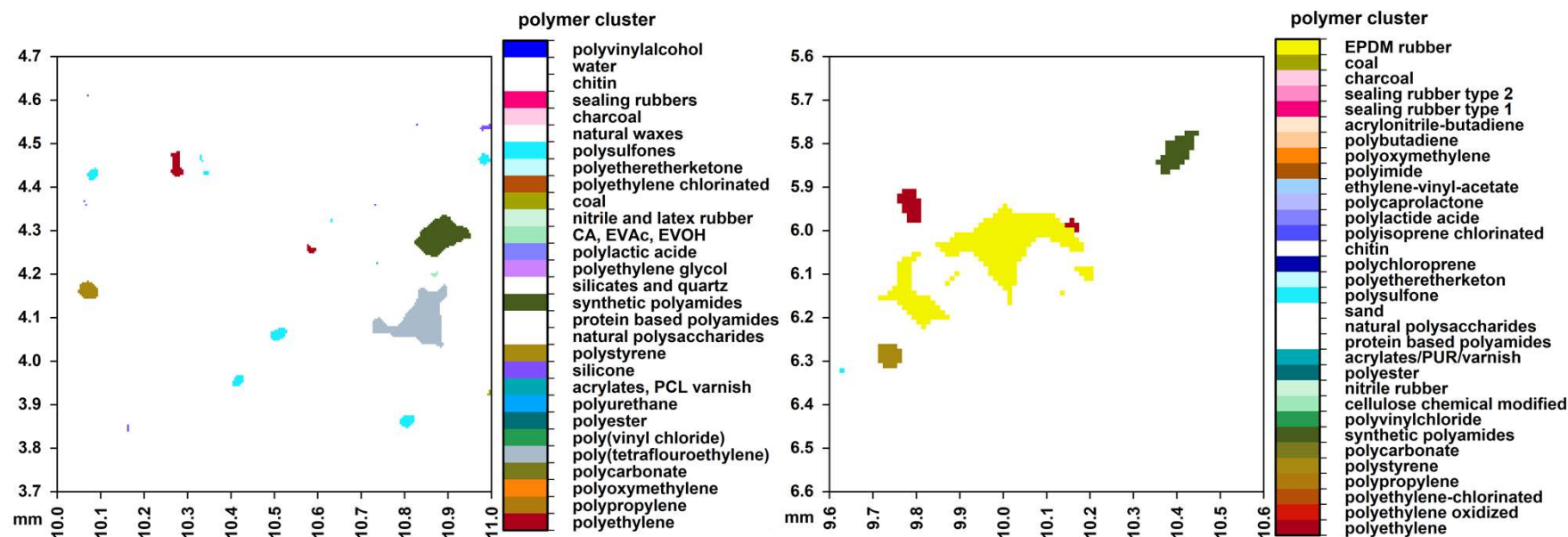
# Polymer types detectable

- Aiming for automated analysis
- Based on hierarchical cluster analysis
- Cluster generation based ATR-FTIR spectra
- Afterwards addition of QCL measured spectra



# Comparsion with FTIR imaging

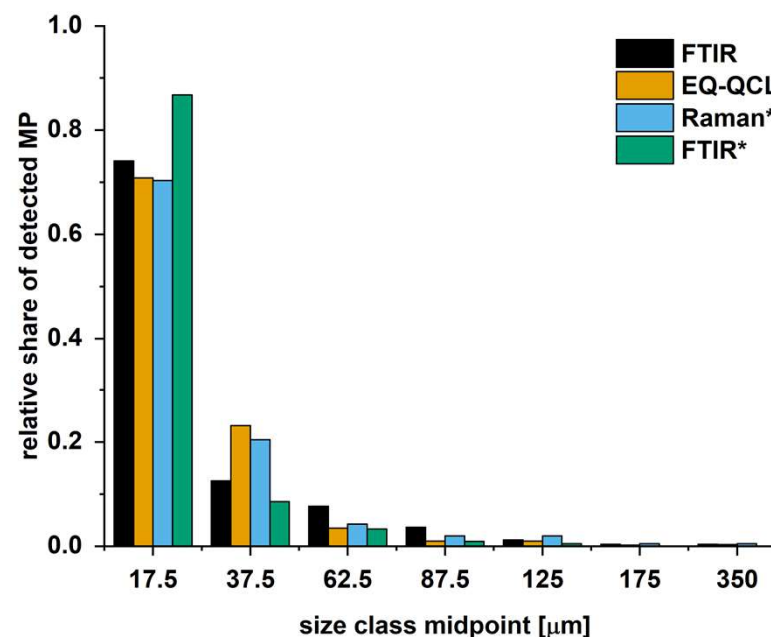
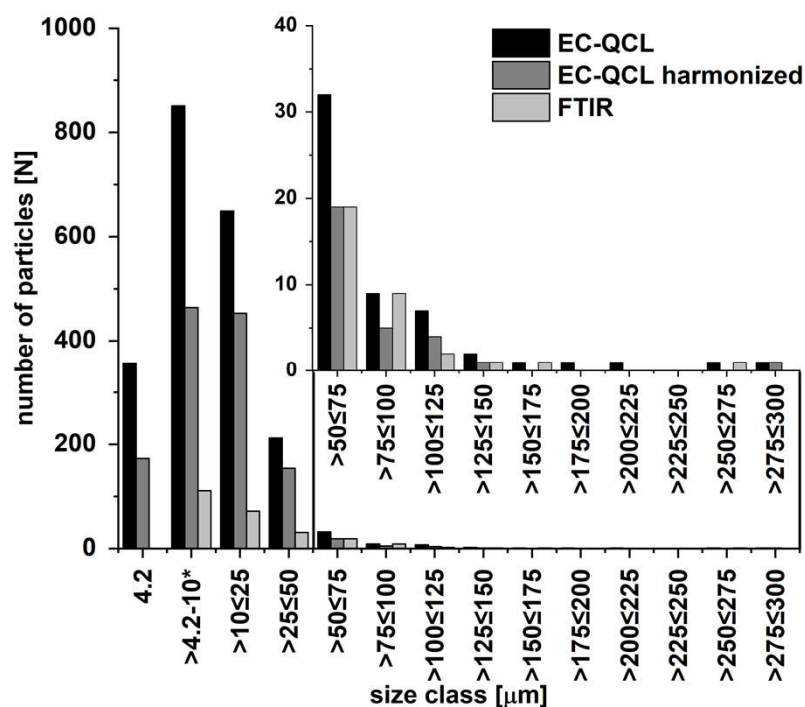
- Similar particles for main polymer types were found.
- In addition more polysulphones and also PTFE were detected.
- The large EPDM assigned particles were not detected.



Graphs: CC 4.0-BY Primpke S, et al. Environ Sci Technol. 2020;54(24):15893-903.  
 doi:10.1021/acs.est.0c05722.

# Comparison with FTIR imaging

- More particles detected compared to FTIR imaging
- Similar relative particle shares like for Raman microscopy (Cabernard et al. 2018)



Graphs: CC 4.0-BY Primpke S, et al. Environ Sci Technol. 2020;54(24):15893-903.  
 doi:10.1021/acs.est.0c05722.

# Comparsion with py-GC/MS



Together with Marten Fischer, Barbara Scholz-Böttcher (ICBM), Claudia Lorenz (AWI, now Aalborg University) and Gunnar Gerdts (AWI)

Direct measurement of particle data for numbers and polymer types followed by mass data for polymer types



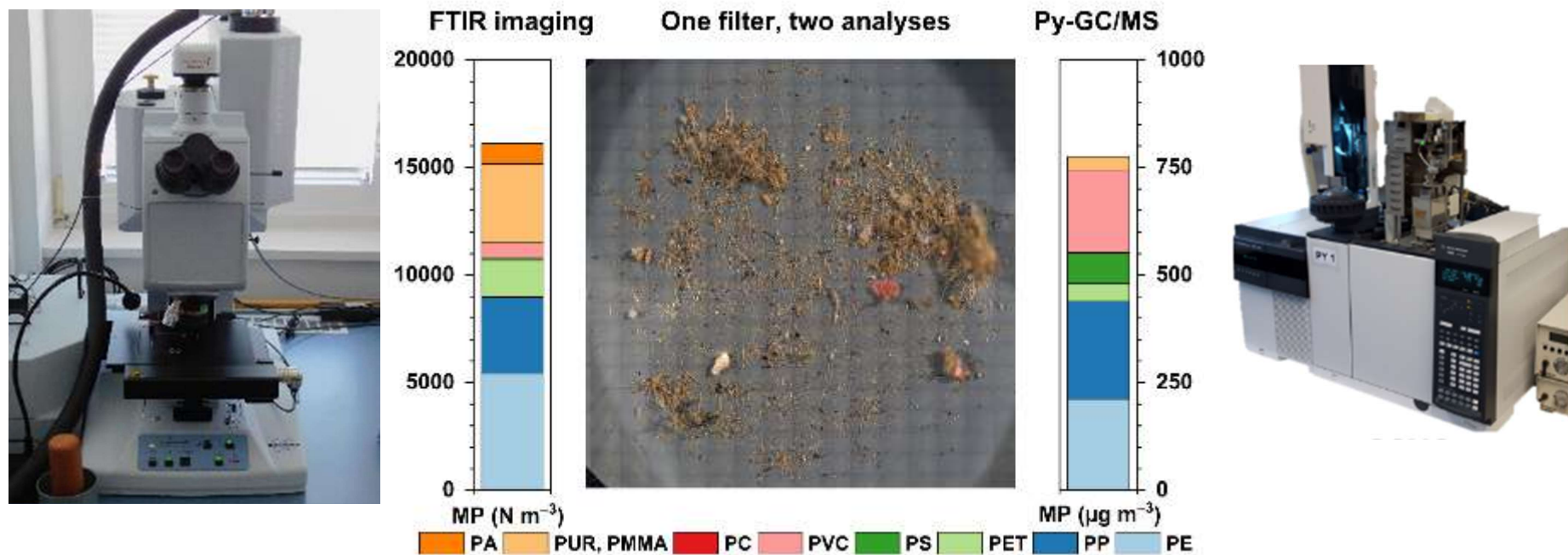
1. S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Full filter analysis

Pyrolysis GC/MS and FTIR imaging

Prior to analysis: Sample preparation via enzymatic digestion

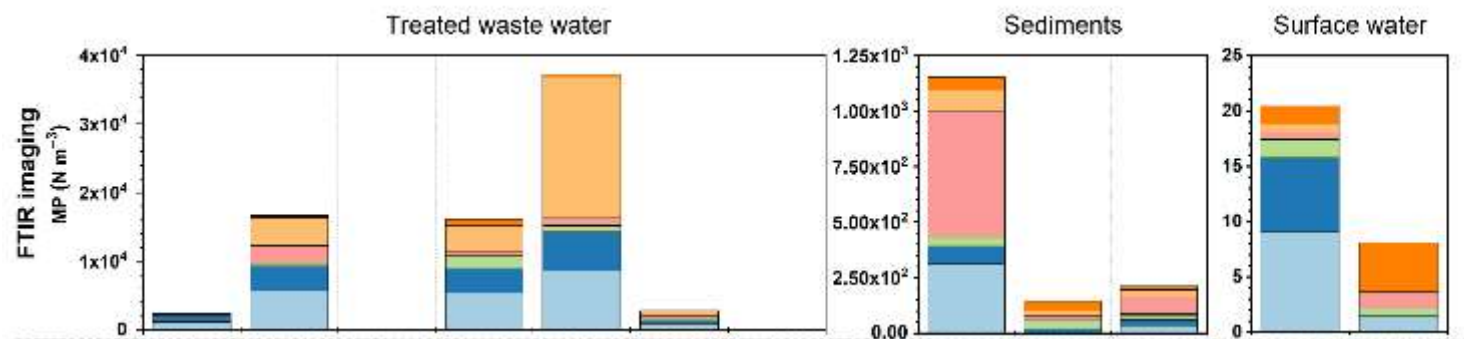
1. Measurement via FTIR imaging using 3.5x FTIR Objective on Anodisc
2. Measurement of same sample via Py-GC/MS at 590°C and on-line derivatization with TMAH



1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298



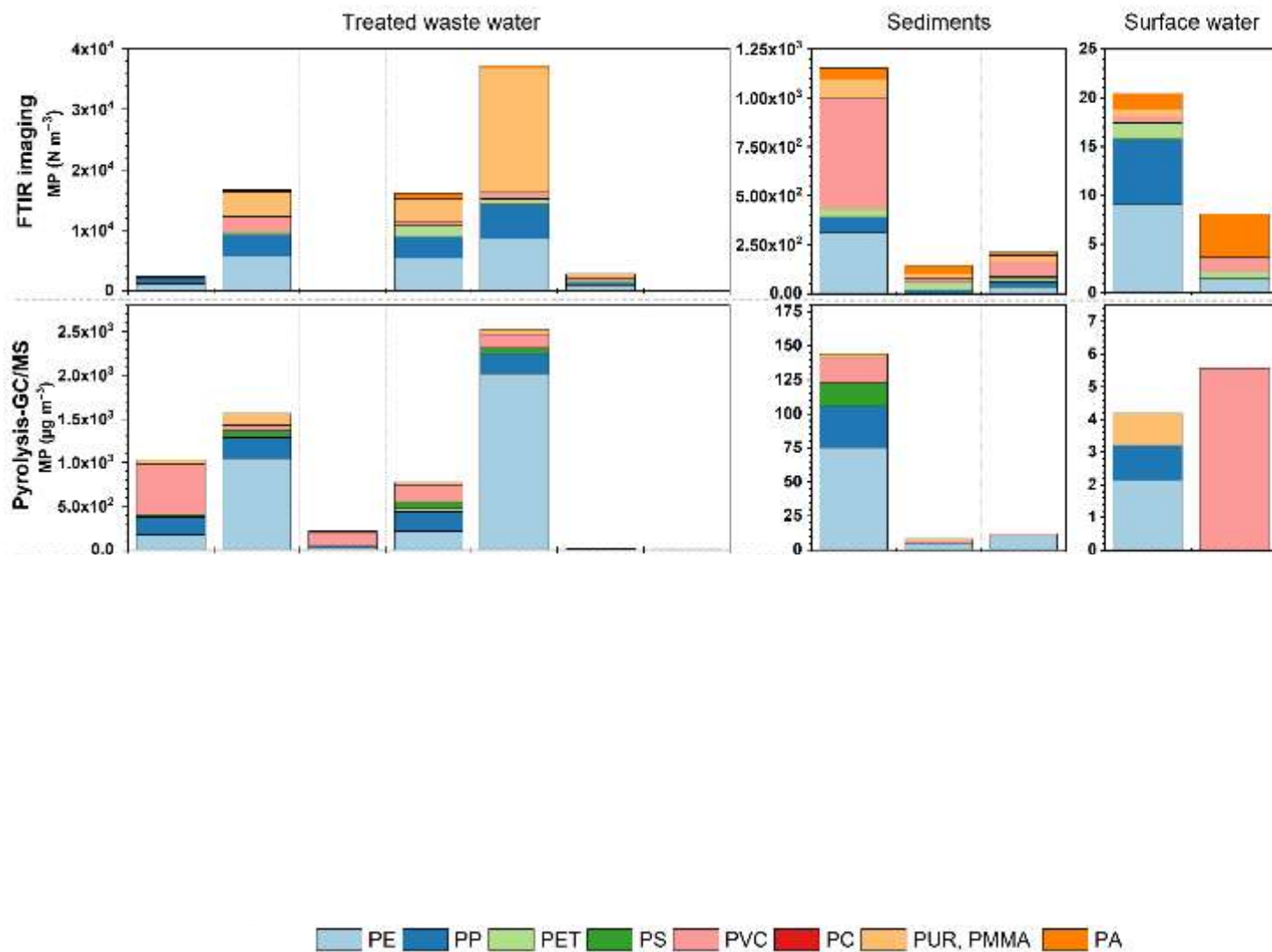
# Results



PE PP PET PS PVC PC PUR, PMMA PA

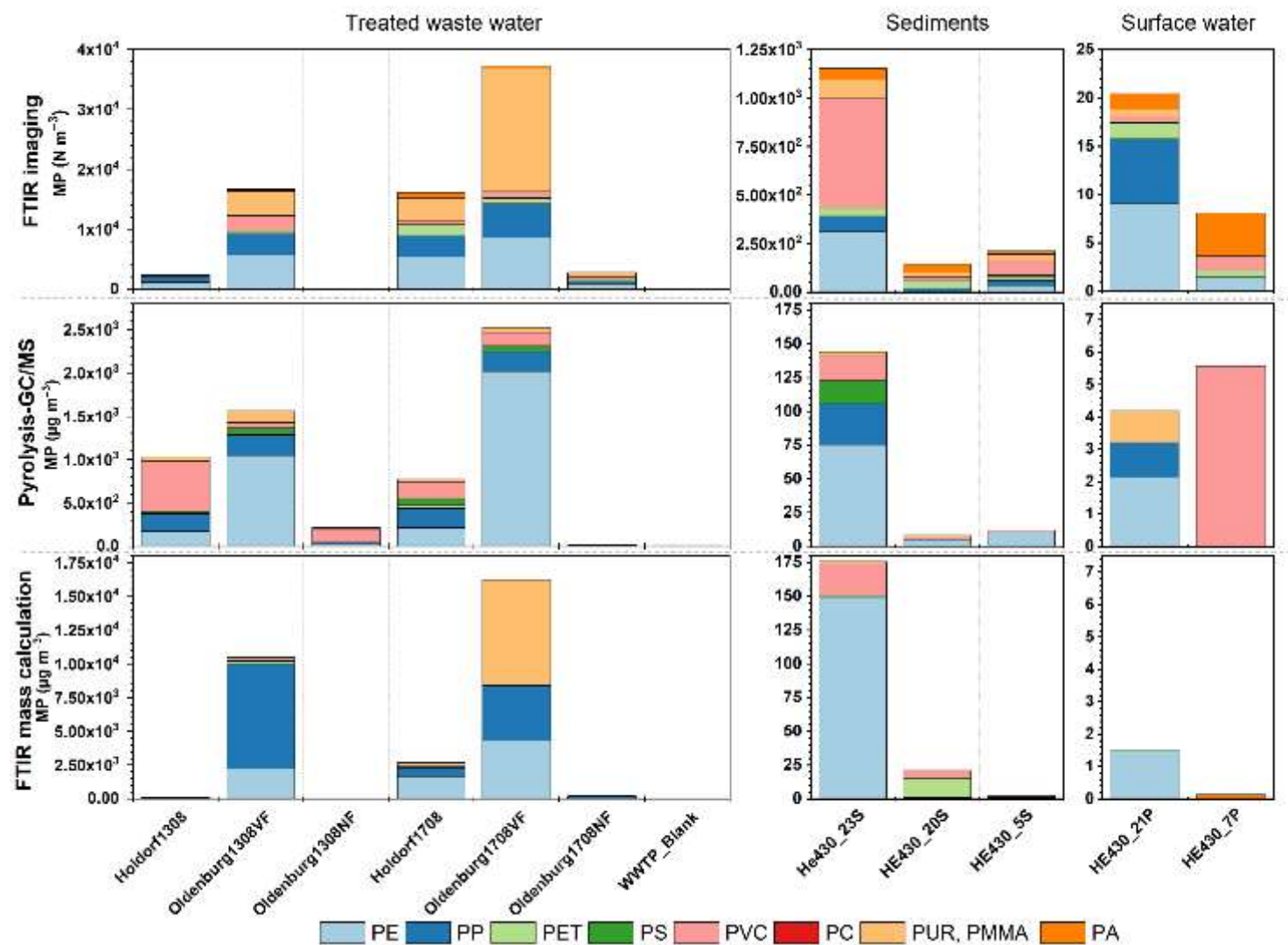
1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Results



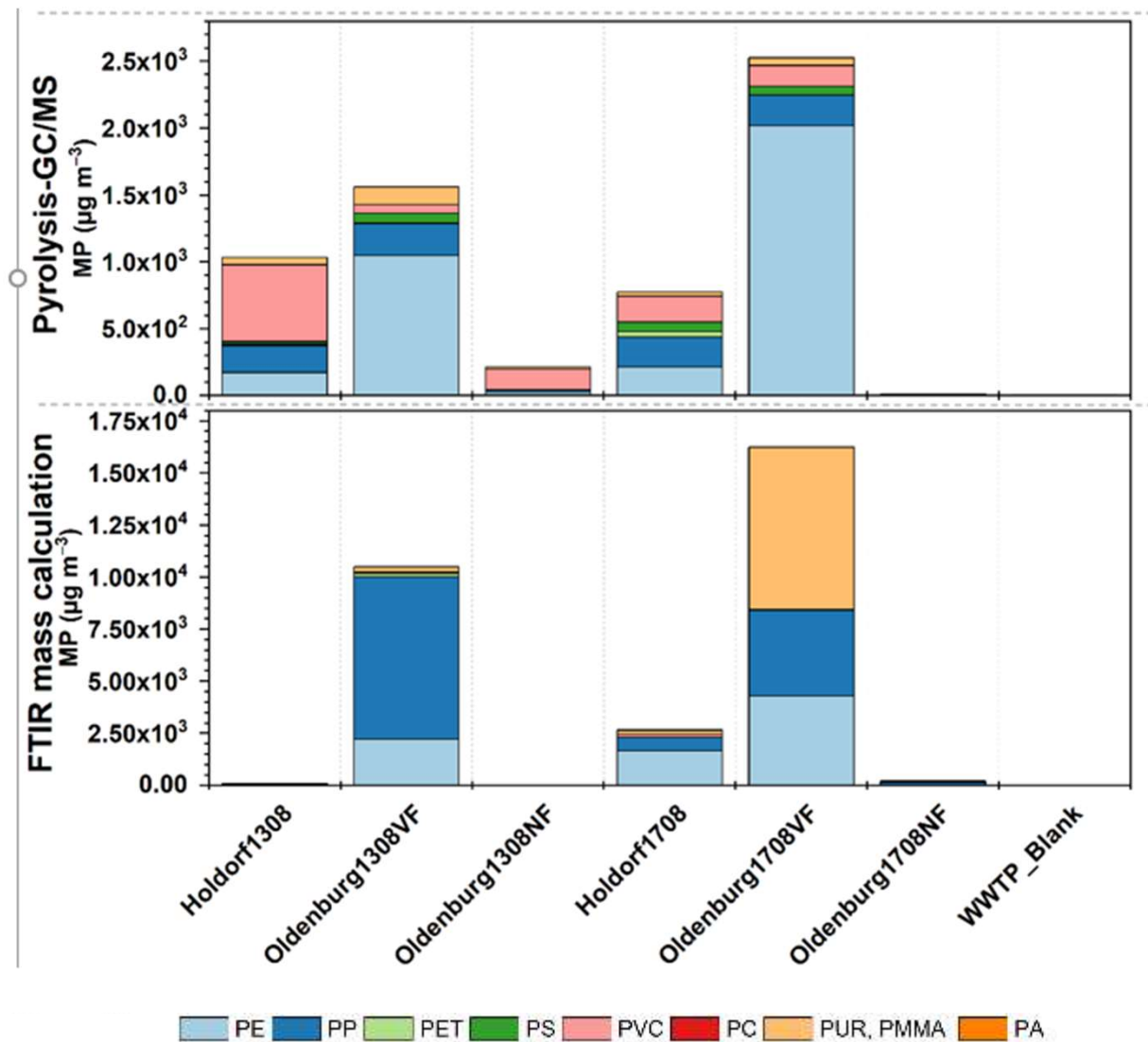
1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Results



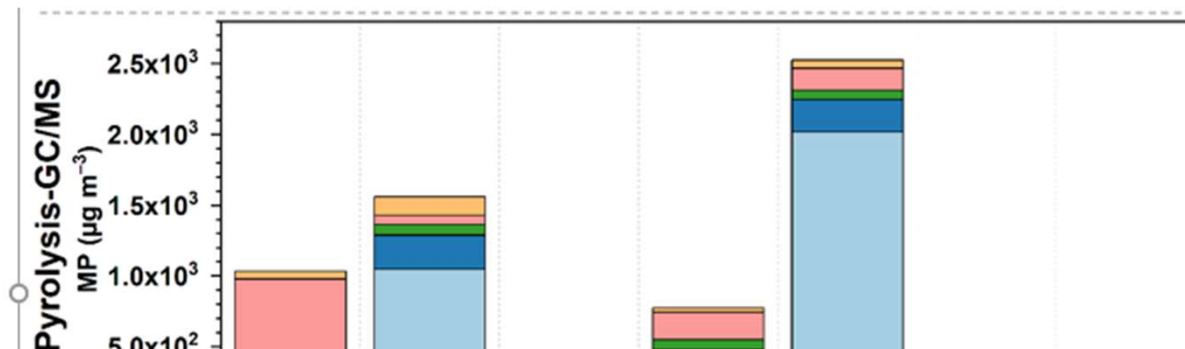
1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Results

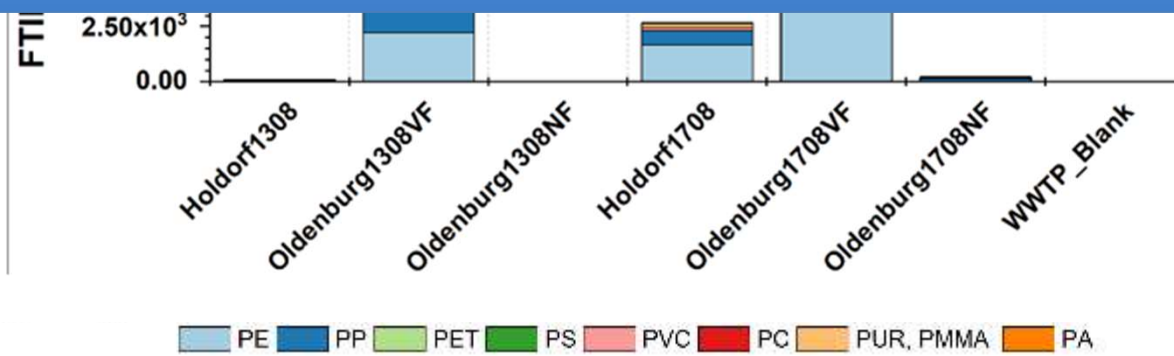


1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Results

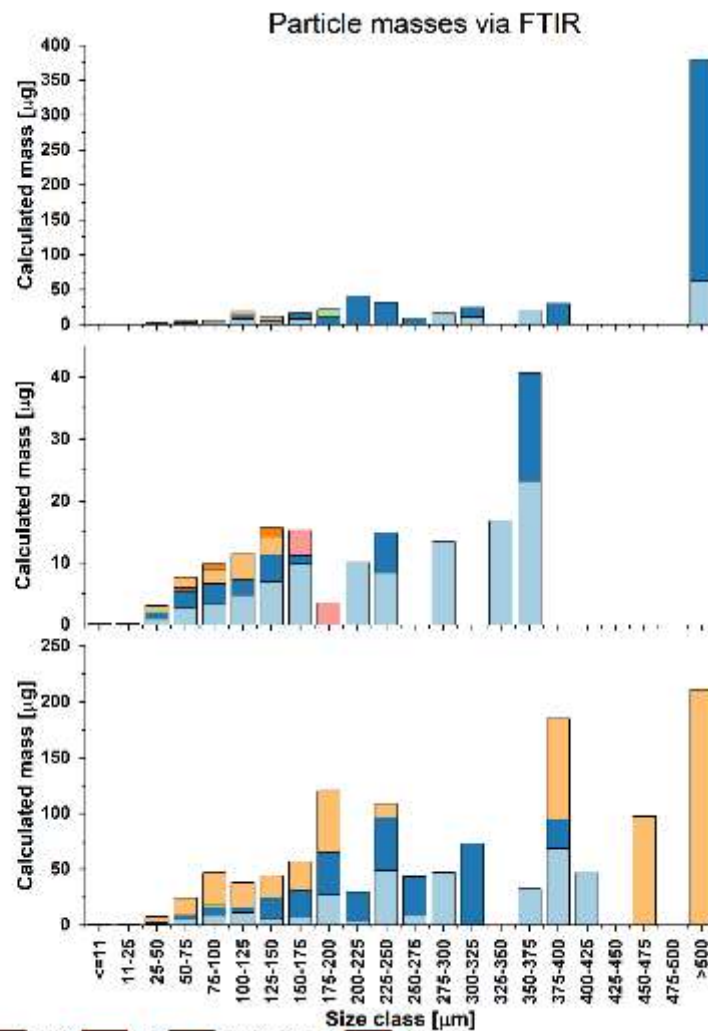
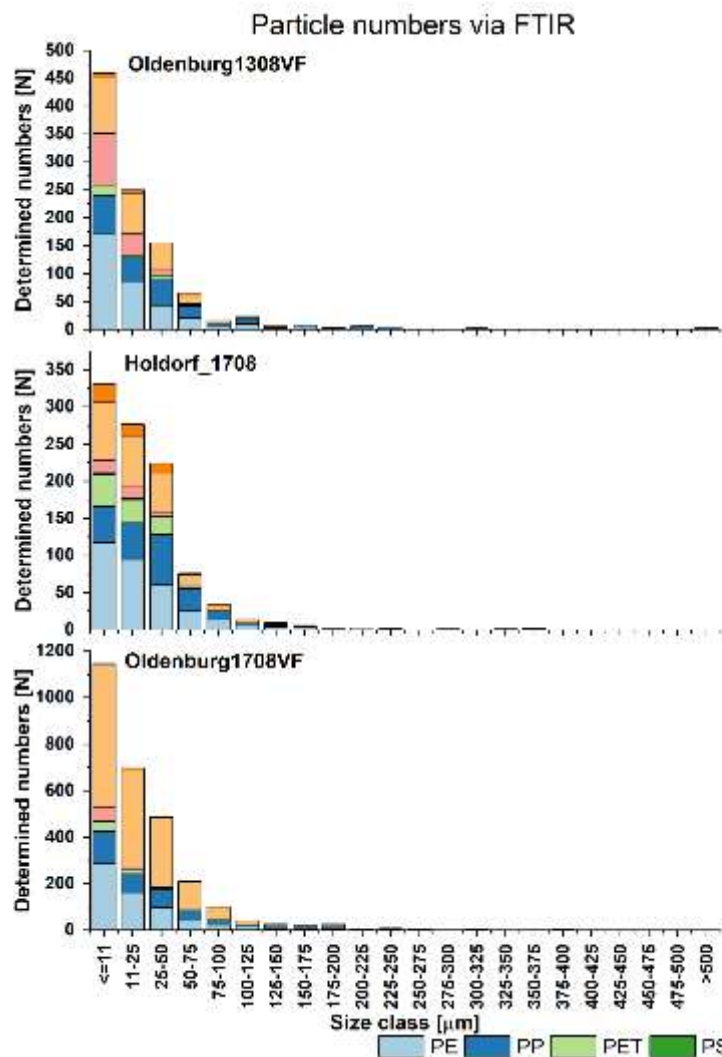


Large differences between calculated (FTIR) and measured (py-GC/MS) mass



1. S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Particle versus mass data

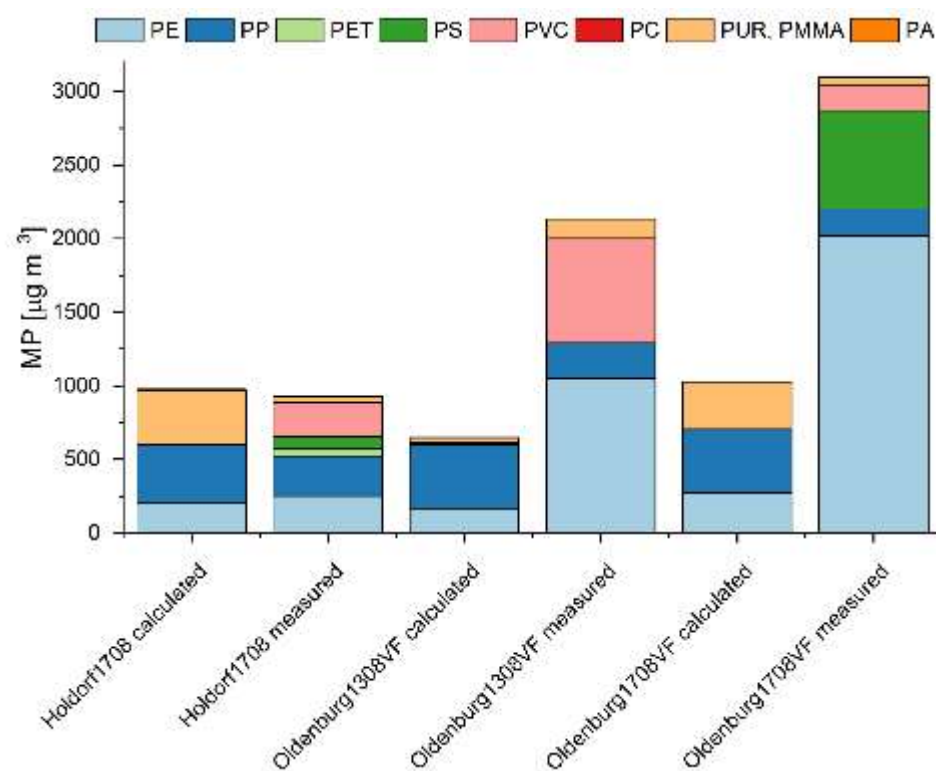


1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

# Alternative approach



- Based on average particle sizes as reference particles
- Calculation of reference particles represented per particle by the measured area
- Yields closer results if many large particles are present.

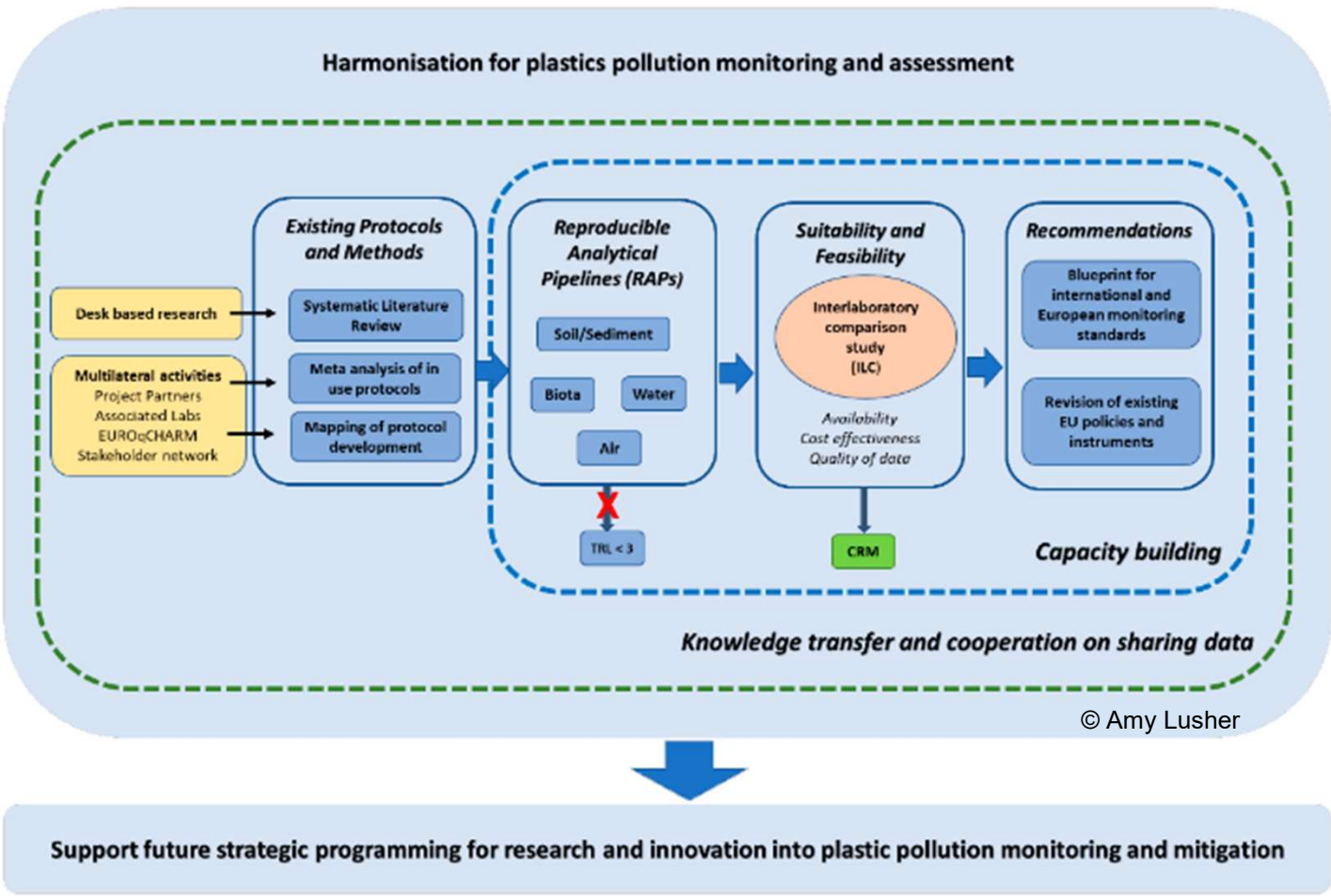


1. Graph: CC 4.0-BY S. Primpke, M. Fischer, et al., *Anal. Bioanal. Chem.*, 2020, 412, 8283-8298

## Lessons learned by the harmonization

- Fast and reliable analyses are available (< 1 hour)
- Mass conversion from spectroscopic data is currently limited for larger particles
- (FT)-IR, Raman and thermoanalytical methods are complementary and can be applied in a step-by-step approach on the same samples





<https://www.euroqcharm.eu/en>

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AWI, Germany: Gunnar Gerdts (Leader WG)  
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students), Marcus Bach (former TA)

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Marquette University, USA: Philippe Ambrozio  
Dias

Aalborg University, Denmark: Jes Vollertsen  
(Professor), Alvis Vianello (Postdoc), Claudia Lorenz  
(Postdoc), PhD students: Nikki van Alst, Márta  
Simon, Kristina B Olesen

University Oldenburg and ICBM: Barbara Scholz-  
Böttcher and Martin Fischer

Matthias Godejohann



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# Question ?

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