

RESEARCH ARTICLE

The Application of DNA Barcodes for the Identification of Marine Crustaceans from the North Sea and Adjacent Regions

Michael J. Raupach^{1*}, Andrea Barco^{1#a}, Dirk Steinke², Jan Beermann³, Silke Laakmann¹, Inga Mohrbeck¹, Hermann Neumann⁴, Terue C. Kihara¹, Karin Pointner¹, Adriana Radulovici², Alexandra Segelken-Voigt⁵, Christina Wesse^{1#b}, Thomas Knebelberger¹

1 German Center of Marine Biodiversity (DZMB), Senckenberg am Meer, Wilhelmshaven, Niedersachsen, Germany, **2** Biodiversity Institute of Ontario, University of Guelph, Guelph, Ontario, Canada, **3** Alfred Wegener Institute Helmholtz Centre for Polar and Marine Research, Biologische Anstalt Helgoland, Helgoland, Schleswig-Holstein, Germany, **4** Department for Marine Research, Senckenberg am Meer, Wilhelmshaven, Niedersachsen, Germany, **5** Animal Biodiversity and Evolutionary Biology, Institute for Biology and Environmental Sciences, V. School of Mathematics and Science, Carl von Ossietzky University Oldenburg, Oldenburg, Niedersachsen, Germany

#a Current address: GEOMAR, Helmholtz-Centre for Ocean Research Kiel, Kiel, Schleswig Holstein, Germany

#b Current address: Botany, School of Biology/Chemistry, Osnabrück University, Osnabrück, Niedersachsen, Germany

* mraupach@senckenberg.de



OPEN ACCESS

Citation: Raupach MJ, Barco A, Steinke D, Beermann J, Laakmann S, Mohrbeck I, et al. (2015) The Application of DNA Barcodes for the Identification of Marine Crustaceans from the North Sea and Adjacent Regions. PLoS ONE 10(9): e0139421. doi:10.1371/journal.pone.0139421

Editor: Roberta Cimmaruta, Tuscia University, ITALY

Received: June 18, 2015

Accepted: September 14, 2015

Published: September 29, 2015

Copyright: © 2015 Raupach et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: Relevant voucher information, taxonomic classifications, photos, DNA barcodes, used primer pairs and trace files are publicly accessible through the public data set "Crustacea of the North Sea" (Dataset ID: DS-CRNS; dx.doi.org/10.5883/DS-CRNS) on the Barcode of Life Data Systems (BOLD; www.boldsystems.org). In addition, all barcode sequences were deposited on GenBank (accession numbers KT208391 to KT209586; Bankit: 1835202).

Funding: MJR received funding from Federal Ministry of Education and Research of Germany (Projekträger Jülich); <https://www.ptj.de/index.php>;

Abstract

During the last years DNA barcoding has become a popular method of choice for molecular specimen identification. Here we present a comprehensive DNA barcode library of various crustacean taxa found in the North Sea, one of the most extensively studied marine regions of the world. Our data set includes 1,332 barcodes covering 205 species, including taxa of the Amphipoda, Copepoda, Decapoda, Isopoda, Thecostraca, and others. This dataset represents the most extensive DNA barcode library of the Crustacea in terms of species number to date. By using the Barcode of Life Data Systems (BOLD), unique BINs were identified for 198 (96.6%) of the analyzed species. Six species were characterized by two BINs (2.9%), and three BINs were found for the amphipod species *Gammarus salinus* Spooner, 1947 (0.4%). Intraspecific distances with values higher than 2.2% were revealed for 13 species (6.3%). Exceptionally high distances of up to 14.87% between two distinct but monophyletic clusters were found for the parasitic copepod *Caligus elongatus* Nordmann, 1832, supporting the results of previous studies that indicated the existence of an overlooked sea louse species. In contrast to these high distances, haplotype-sharing was observed for two decapod spider crab species, *Macropodia parva* Van Noort & Adema, 1985 and *Macropodia rostrata* (Linnaeus, 1761), underlining the need for a taxonomic revision of both species. Summarizing the results, our study confirms the application of DNA barcodes as highly effective identification system for the analyzed marine crustaceans of the North Sea and represents an important milestone for modern biodiversity assessment studies using barcode sequences.

grant no. 03F0664A). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Introduction

In recent years, the use of molecular methods for specimen identification and classification has become quite popular, including proteome [1–3] or spectroscopic data [4,5]. However, the analysis of DNA sequence data represents the most used and accepted application to date. In most animals, mitochondrial DNA (mtDNA) exhibits several characteristics that make it highly attractive for molecular specimen identification, such as an almost exclusively maternal inheritance, a high number of copies within the mitochondria, the absence of introns, typically high substitution rates, and the absence of recombination [6–8]. Furthermore, as a consequence of uniparental inheritance and haploidy, mtDNA has a four-fold smaller effective population size compared to nuclear DNA, resulting in faster lineage sorting [9]. In this context, the standardized use of an approx. 650 base pair (bp) fragment of the cytochrome *c* oxidase subunit 1 (CO1) as DNA barcode represents a very successful mtDNA-based approach for the identification of animal specimens [10–12]. The idea of DNA barcoding is based on the assumption that each species will have similar DNA barcodes representing its intraspecific variability. In addition, the variation between species needs to exceed the variation within species, which allows a clear genetic delineation of species by so-called barcoding gaps [10,11,13]. The two main goals of DNA barcoding are (i) to assign unknown specimens to already described and classified species, and (ii) to enhance the discovery of new species and facilitate identification, particularly in cryptic, microscopic, and other organisms with complex or inaccessible morphology [10,11]. Whereas various phenomena may affect the application of DNA barcodes or mitochondrial DNA in general for successful specimen identification, e.g. heteroplasmy [14,15], incomplete lineage sorting [16], the presence of mitochondrial pseudogenes (numts) [17,18] or introgressive hybridization [19,20], DNA barcoding has become an important tool in numerous biological disciplines, e.g. modern biodiversity assessment studies [12,21–23], conservation biology [12,24], or the authentication of sea food [25,26]. As consequence, many recently published species descriptions and taxonomic studies included barcode sequence data [27–31].

Within the Arthropoda, most DNA barcoding publications focus on insects [32–38], whereas the number of comprehensive studies analyzing the utility of DNA barcodes for the discrimination of crustacean species is still limited [39–43]. Nevertheless, crustaceans represent one of the most ecologically and economically important invertebrate groups [44]. Currently, more than 67,000 extant species have been described so far [45], and probably five or ten times of that number are waiting to be discovered in the marine realm [46]. Crustaceans successfully colonized every marine, brackish, and freshwater environment on Earth, and exhibit an astonishing diversity of form, habit, and size. No other group of plants or animals shows a morphological diversity as seen among the extant Crustacea [47], ranging from the tiny tantulocarid species *Stygotantulus stocki* Boxshall & Huys, 1989 with a body length less than 0.1 mm up to the giant spider crab *Macrocheira kaempferi* Temminck, 1836 with a documented leg span of up to 3.7 m [48]. In the case of parasitic crustaceans, many species can only be identified as crustaceans by reference to their larval stages, for example species of the rhizocephalan genus *Sacculina* Thompson, 1836 (Thecostraca) or various copepod species (e.g. the genus *Lernaenicus* Le Sueur, 1824).

In this study we present a comprehensive DNA barcode library of various crustacean taxa found in the North Sea, one of the most extensively studied ecosystems of the world. The North Sea is characterized by a high amount of anthropogenic pressure such as intensive fishing and ship traffic as well as offshore installations. Environmental parameters (e.g. depth, sediment characteristics, temperature and salinity) of this semi-enclosed shelf sea follow a distinct pattern: high seasonal fluctuations can be observed in southern areas, less fluctuations are found in the northern regions [49,50]. This heterogeneity is also displayed in macrobenthic

community structures, with a lower number of species in the shallow southern parts (i.e. the German Bight) and more species in the central and northern North Sea [51–54]. Furthermore, species with a typical Mediterranean-Lusitanian distribution are also known to occur in parts of the North Sea where oceanic influences prevail [55].

Our new barcode library includes a broad coverage of crustacean species of different taxa inhabiting the North Sea, ranging from large king crabs (e.g. *Lithodes maja* (Linnaeus, 1758)) to minute species that are elements of the benthic meiofauna (e.g. *Asellopsis intermedia* (Scott T., 1895)) as well as highly modified parasites of crustaceans (e.g. *Peltogaster paguri* Rathke, 1842) or fish (e.g. *Chondracanthus merluccii* (Holten, 1802)).

Material and Methods

Sampling of specimens

All analyzed crustaceans were collected between 2003 and 2014 using various sampling methods (i.e. hand collecting, Van Veen grab sampler, various dredges, bottom trawls). The Nationalparkverwaltung Niedersächsisches Wattenmeer (Wilhelmshaven) and the Johann Heinrich von Thünen-Institut, Abteilung Seefischerei (Hamburg), issued the permission to conduct this study. Our field studies have not involved endangered or protected species. All crustaceans were morphologically identified to species level by eight of the authors (JB, TCK, SL, IM, HN, KP, AS-V, MR) or by other taxonomic experts and matched with the online database World Register of Marine Species [56]. The applied taxonomic classification is based on the most recent system [45] (Table 1).

For our analysis we also included 136 DNA barcodes of 13 calanoid copepod species of a previous study [3]. Most specimens were collected in the North Sea ($n = 1,285$, 96.5%), but for comparison some specimens from the English Channel (30, 2.3%), the Baltic Sea (6, 0.4%), and

Table 1. Number of barcoded species of different crustacean orders from the North Sea. Note that 136 DNA barcodes of 13 species of the Calanoida (Maxillopoda: Copepoda: Gymnoplea) were already published as part of a previous study [3].

Class	Subclass	Superorder	Order	Number of analyzed species	Number of analyzed specimens	
Branchiopoda	Phyllopoda		Diplostraca	5	24	
Maxillopoda	Thecostraca	Rhizocephala	Aktentrogonida	1	7	
			Kentrogonida	2	8	
			Thoracica	Lepadiformes	1	4
			Scalpelliformes	1	6	
			Sessilia	7	67	
			Copepoda	Gymnoplea	Calanoida	14
			Podoplea	Cyclopoida	4	22
				Harpacticoida	10	31
				Siphonostomatoidea	5	26
	Malacostraca	Hoplocarida		Monstrilloida	1	8
			Stomatopoda	1	1	
Eumalacostraca		Peracarida	Mysida	7	35	
				Amphipoda	59	305
				Isopoda	16	84
				Cumacea	4	14
			Eucarida	Euphausiacea	1	4
				Decapoda	66	529
Total				205	1332	

doi:10.1371/journal.pone.0139421.t001

some other locations of Germany (11, 0.8%) were also included. All specimens were stored in ethanol (96%). The number of analyzed specimens per species ranged from one individual (26 species, 12.7%) to a maximum of 32 for the long-clawed porcelain crab *Pisidia longicornis* (Linnaeus, 1767) (Malacostraca: Decapoda).

DNA sequencing and data depository

Laboratory operations were carried out either at the Canadian Center for DNA Barcoding (CCDB), University of Guelph, following standardized high-throughput protocols for DNA barcode amplification and sequencing [57,58], or at the molecular lab of the German Center of Marine Biodiversity Research, Senckenberg am Meer, in Wilhelmshaven, Germany. For small specimens with a body length <3 mm, complete specimens were used for DNA extraction, whereas tissue samples (e.g. legs or pleon muscles) were used for individuals >3 mm. In Wilhelmshaven, DNA was extracted using the QIAmp Tissue Kit (Qiagen GmbH, Hilden, Germany) or NucleoSpin Tissue Kit (Macherey-Nagel, Düren, Germany), following the manufactures protocol. Polymerase chain reaction (PCR) was performed for amplifying the CO1 barcode fragment using two primer pairs (LCO1480/HCO2198 [59]; or jgLCO1490/jgHCO2198 [60]). For the primer pair jgLCO1490/jgHCO2198 we added M13 forward and reverse tails to provide defined nucleotide sequences for sequencing [61]. All PCR products were amplified using illustra puReTaq Ready-To-Go PCR Beads (GE Healthcare, Buckinghamshire, UK) in a total volume of 20 µl, containing 17.5 µl sterile molecular grade H₂O, 2 µl DNA template with an DNA amount between 2 to 150 ng/µl, and 0.25 µl of each primer (20 pmol/µl). The PCR thermal conditions included an initial denaturation at 94°C (5 min), followed by 38 cycles at 94°C (denaturation, 45 s), 48°C (annealing, 45 s), 72°C (extension, 80 s), and a final extension step at 72°C (7 min). All PCR reactions were conducted using an Eppendorf Mastercycler Pro system (Eppendorf, Hamburg, Germany). Negative and positive controls were included with each round of reactions. Two µl of the amplified products were verified for size conformity by electrophoresis in a 1% agarose gel with GelRed using commercial DNA size standards, whereas the remaining PCR product was purified with the QIAquick PCR Purification Kit (Qiagen GmbH, Hilden, Germany). Purified amplicons were cycle sequenced and sequenced in both directions at a contract sequencing facility (GATC, Konstanz, Germany) using LCO1480 and HCO2198 as sequencing primers or the M13 sequence tails for jgLCO1490/jgHCO2198 as matrix (see above). Double stranded sequences were assembled and checked for the presence of mitochondrial pseudogenes (numts) with the Geneious version 7.0.4 program package [62] by translating all nucleotide sequences in amino acid sequences. BLAST searches were performed to confirm the identity of all new sequences [63,64].

All analyzed barcodes had a length of at least 500 base pairs (bp). Relevant voucher information, taxonomic classifications, photos, DNA barcodes, used primer pairs and trace files are publicly accessible through the public data set “Crustacea of the North Sea” (Dataset ID: DS-CRNS; dx.doi.org/10.5883/DS-CRNS) on the Barcode of Life Data Systems (BOLD; www.boldsystems.org) [65]. In addition, all barcode sequences were deposited on GenBank (accession numbers KT208391 to KT209586; Bankit: 1835202).

DNA barcode analysis

Intra- and interspecific nucleotide variability of the analyzed crustaceans was based on the Kimura 2-parameter model (K2P; [66]), using the analytical tools on BOLD (align sequences: BOLD aligner; ambiguous base/gap handling: pairwise deletion). The BOLD workbench was also used to calculate base frequencies. In addition, all barcodes were subject to the Barcode Index Number (BIN) system implemented in BOLD [67]. This approach clusters DNA

barcodes to calculate operational taxonomic units (OTUs) that closely correspond to species. BIN clusters are indexed in a regimented way which means that genetically identical taxa of different studies reside under a shared identifier [67]. However, BINs are not necessarily stable over time and may change, for example as a consequence of the addition of new barcode sequences to BOLD. A recommended threshold of 2.2%, as it has been demonstrated in eight test datasets, was used for a rough differentiation of low and high intraspecific as well as interspecific K2P distances [67].

We performed neighbor joining cluster analyses (NJ; [68]) to construct a graphical representation of patterns of nucleotide divergences based on K2P distances using MEGA6.4 [69] for all Copepoda, Amphipoda, Decapoda, Thecostraca, Isopoda, and all other taxa. Non-parametric bootstrap support values were obtained by resampling and analyzing 1,000 replicates [70]. Another NJ analysis (K2P distances) was performed for all analyzed specimen with non-parametric bootstrap replicates ($n = 1,000$). For all analyses, barcodes were aligned using MUSCLE [71], implemented in MEGA6.4. The sequence alignment for all taxa and the MEGA-generated K2P NJ tree file in text format were uploaded to TreeParser [72], producing an output FASTA file that followed the order of terminals in the tree. A Klee diagram was generated by indicator vector analysis [73] with parameters $n = 1$ sequence/vector and bp window size = 10–600.

For species that showed identical haplotypes, statistical maximum parsimony networks were constructed with TCS 1.21 using default settings [74]. Such networks allow the identification of haplotype sharing between species as a consequence of recent speciation or on-going hybridization.

Finally we performed a simulation of sequence-based identification of specimens using the R library SPIDER [75]. Each sequence was used as a query against the entire dataset. Identification was provided following three different criteria: Best Match (BM); Best Close Match (BCM); and All Species Barcode (ASB). The BM criterion assigns identifications to the closest match regardless of the distance. The BCM criterion [76] is similar to BM, but the query is identified by the closest match with a distance below a defined threshold. Finally, the ASB criterion simulates the BOLD ID engine by applying a threshold and querying at all the sequences within it. A query is identified when all the matching sequences below the threshold are conspecific. Results are reported as correct when corresponding to prior morphological identifications, otherwise a result counts as incorrect. For BCM and ASB, a query may provide ambiguous results if sequences divergences of different species are below the threshold (ASB) or sequences from different species are the closest match below threshold (BCM). A query resulting in “no ID” has no match below the defined threshold. For BCM and ASB we used three different thresholds: the value of 1% (K2P), which is the standard used by the BOLD ID engine [65]. As second threshold we used the value that minimizes the cumulative identification errors (function ‘threshVal’ in SPIDER), i.e. the sum of false positive (no conspecific matches within threshold of query) and false negative (sequences from multiple species within threshold). Finally, we used a density plot of genetic distances and evaluated where a minimum in the density corresponds to the transition between intra- and interspecific distances (function ‘localMinima’ in SPIDER) as third threshold. All simulations were run twice, with and without singletons (species represented by a single sequence).

Results

In total, 1,332 DNA barcodes of 205 species were analyzed. No numts were found. A full list of the analyzed species can be found in the supporting information (S1 Table). Fragment lengths of the analyzed DNA barcodes ranged from 514 to 667 bp. For 129 species (61.4%), five or

Table 2. Table of 13 species of the Crustacea with a maximum intraspecific distance (K2P) of >2.2%. At least two specimens of the listed species showed a distance value higher than the threshold as part of a pairwise comparison.

Order	Species	Number of analyzed specimens (n)	Mean pairwise K2P distance (%)	Maximum pairwise K2P distance (%)	BINs
Amphipoda	<i>Photis longicaudata</i>	6	0.98	2.36	ACG9506
Calanoida	<i>Temora longicornis</i>	19	0.48	2.66	AAO2762
Decapoda	<i>Pagurus pubescens</i>	12	1.6	2.89	AAB6221
Calanoida	<i>Calanus helgolandicus</i>	14	0.64	2.99	AAB3934
Amphipoda	<i>Monocorophium insidiosum</i>	9	1.24	3.41	AAE9749, AAE1628
Sessilia	<i>Austrominius modestus</i>	11	1.39	3.81	ABX4245, ACR4768
Decapoda	<i>Pandalus montagui</i>	21	1.49	4	AAB2199
Amphipoda	<i>Gammarus salinus</i>	6	2.01	4.14	AAB7068, ACG9079, ACG8870
Calanoida	<i>Pseudocalanus elongatus</i>	9	1.93	4.43	AAF6145
Decapoda	<i>Eriocheir sinensis</i>	6	2.06	4.78	ABB0750, AAA8754
Isopoda	<i>Astacilla intermedia</i>	3	3.19	4.79	ACP7495, ACP7496
Calanoida	<i>Anomalocera patersoni</i>	22	1.87	6.11	ACM8185, ACM8186
Siphonostomatoida	<i>Caligus elongatus</i>	10	6.9	14.87	AAE8403, AAE8404

doi:10.1371/journal.pone.0139421.t002

more DNA barcodes have been generated (S1 Fig). Similar to other arthropod studies, our data indicated a high AT-content for this mitochondrial gene fragment: the mean sequence compositions were A = 26%, C = 19%, G = 19% and T = 36%. Intraspecific K2P divergences ranged from zero to 14.87% whereas interspecific distances were between 0% and 44.38% (S1 Table). The lowest intraspecific distances of clearly distinct barcode clusters were revealed for the closely related spider crab species pair *Hyas araneus* (Linnaeus, 1758) and *Hyas coarctatus* Leach, 1816 with a value of 2.36%. Maximum intraspecific pairwise distances >2.2% were found for 13 species, including one thecostrac, one isopod, three amphipod, three decapod and five copepod species (Table 2). In contrast to this we found low pairwise distances with values <2.2% only for one decapod species pair: *Macropodia parva* Van Noort & Adema, 1985 and *Macropodia rostrata* (Linnaeus, 1761). Unique BINs were revealed for 198 species (96.7%), two BINs for six species (2.9%), and three BINs for the amphipod species *Gammarus salinus* Spooner, 1947 (0.4%).

Our NJ analyses based on K2P genetic distances revealed non-overlapping species clusters with bootstrap support values of 99 or 100% for all Copepoda (Fig 1), all Amphipoda (Fig 2), most Decapoda (Fig 3) as well as all Thecostraca, Isopoda, Cumacea, Diplostraca, Euphausiacea, Mysida and Stomatopoda (all in Fig 4). A NJ topology of all analyzed crustacean specimens based on K2P is presented in the supporting information (S2 Fig).

A Klee diagram of the TreeParser-ordered alignment showed blocks of high correlation on the diagonal, reflecting the affinity among species (Fig 5). The used NJ topology is presented in the supporting information (S3 Fig). Maximum correlation was observed among neighboring species, and decorrelation among more distant species. Given the broad sampling across all crustaceans the latter occurs much more frequently. Unusual strong correlation was observed among two species (*Macropodia parva* and *Macropodia rostrata*) as a result of haplotype sharing.

The statistical maximum parsimony analysis also revealed multiple sharing of haplotypes for *Macropodia parva* (n = 9) and *Macropodia rostrata* (n = 7) (Fig 6). In total, seven

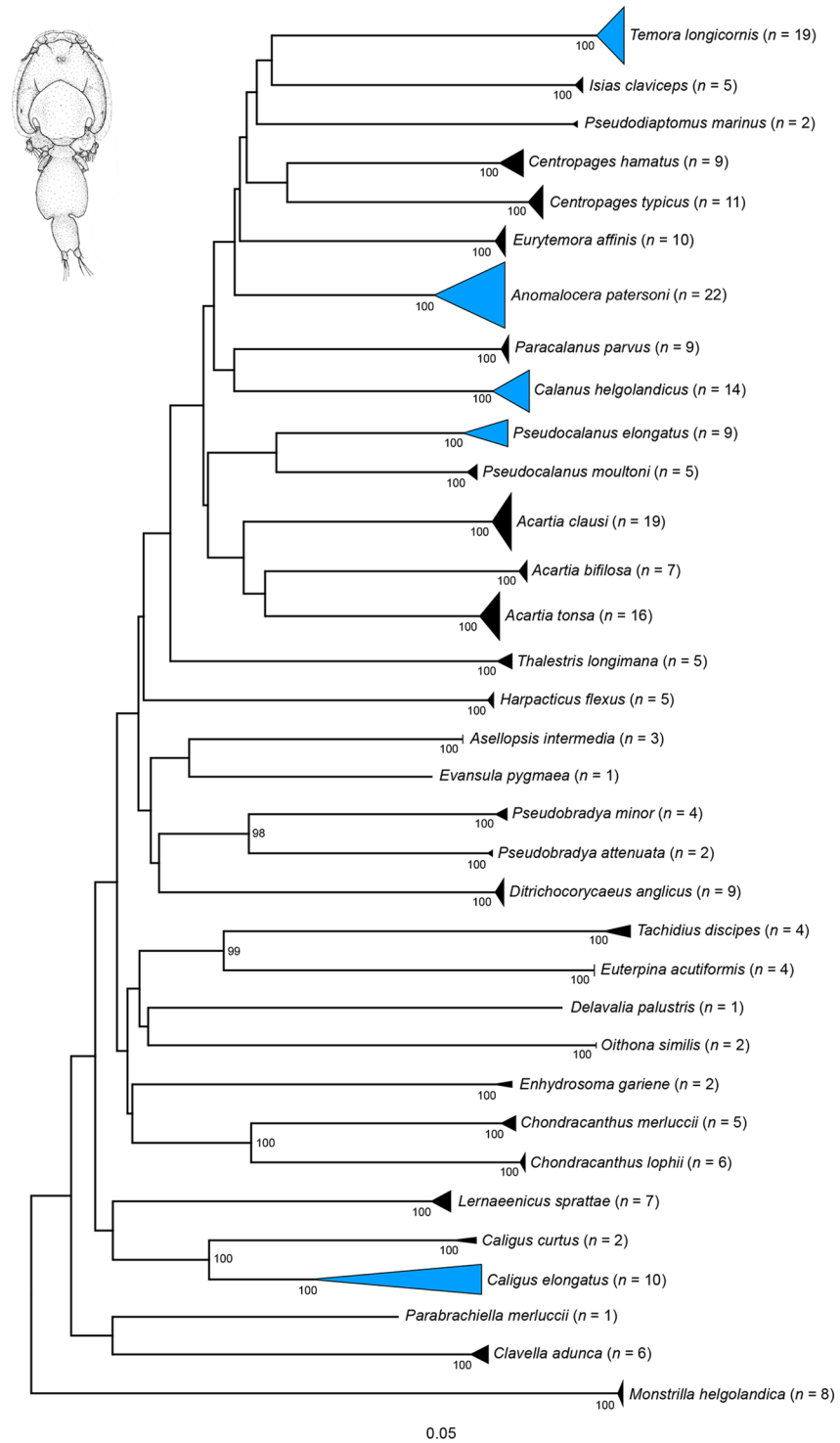


Fig 1. Neighbor joining topology of the analyzed Copepoda based on Kimura 2-parameter distances. The number of analyzed specimens collapsed into a single node is provided following the species name. Triangles indicate the relative number of individual's sampled (height) and sequence divergence (width). Blue triangles indicate species with intraspecific maximum pairwise distances >2.2%. Numbers next to nodes represent non-parametric bootstrap values >90% (1,000 replicates). Drawing of *Caligus curtus* O.F. Müller, 1789 is taken and modified from a previous publication [77].

doi:10.1371/journal.pone.0139421.g001

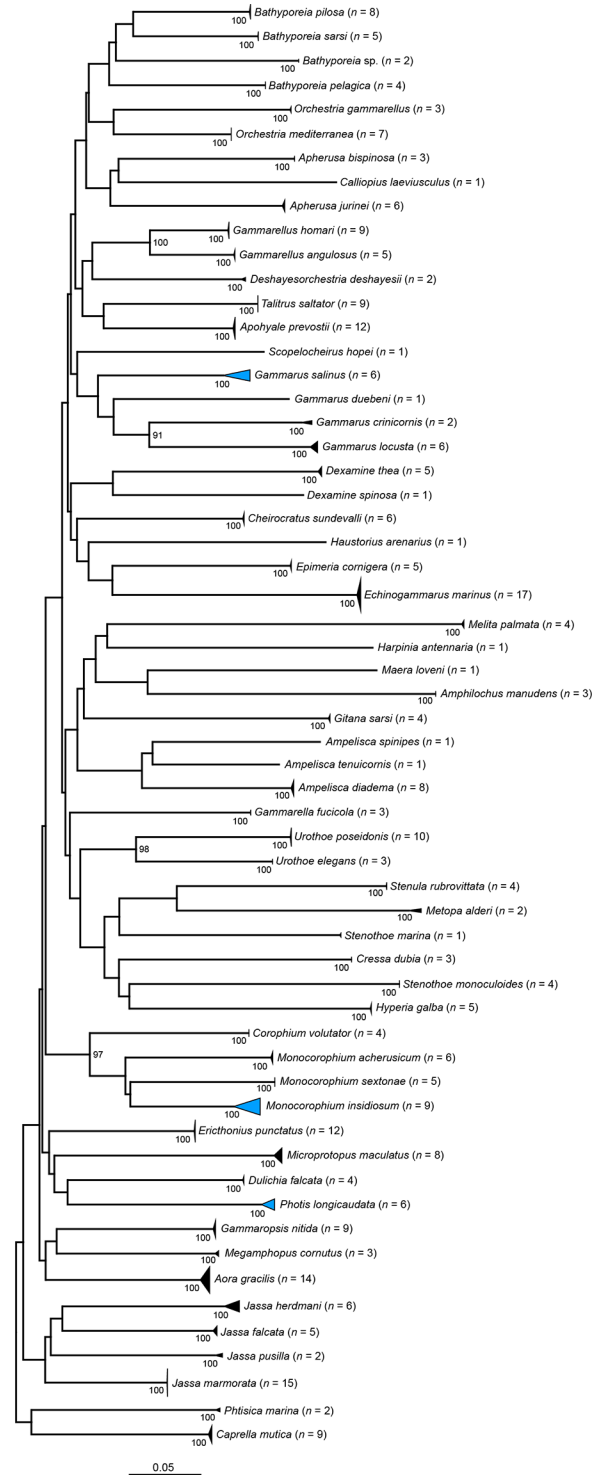


Fig 2. Neighbor joining topology of the analyzed Amphipoda based on Kimura 2-parameter distances. The number of analyzed specimens collapsed into a single node is provided following the species name. Triangles indicate the relative number of individual's sampled (height) and sequence divergence (width). Blue triangles indicate species with intraspecific maximum pairwise distances >2.2%. Numbers next to nodes represent non-parametric bootstrap values >90% (1,000 replicates). Drawing of *Melita palmata* Montagu, 1804 is taken and modified from a previous publication [78].

doi:10.1371/journal.pone.0139421.g002

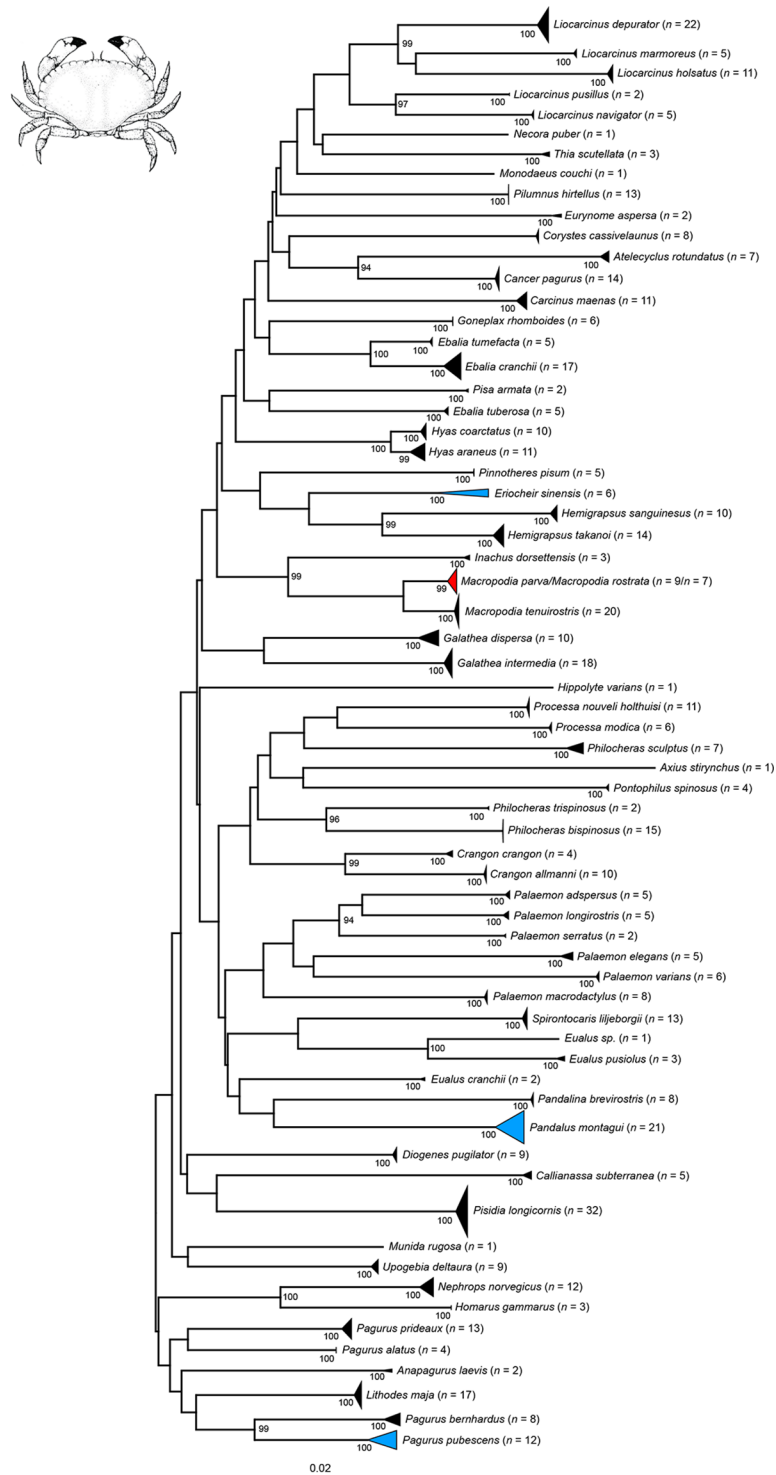


Fig 3. Neighbor joining topology of the analyzed Decapoda based on Kimura 2-parameter distances. The number of analyzed specimens collapsed into a single node is provided following the species name. Triangles indicate the relative number of individual's sampled (height) and sequence divergence (width). Blue triangles indicate species with intraspecific maximum pairwise distances >2.2%, the red triangle species with interspecific distance values <2.2%. Numbers next to nodes represent non-parametric bootstrap values >90% (1,000 replicates). Drawing of *Cancer pagurus* Linnaeus, 1758 is taken and modified from a previous publication [79].

doi:10.1371/journal.pone.0139421.g003

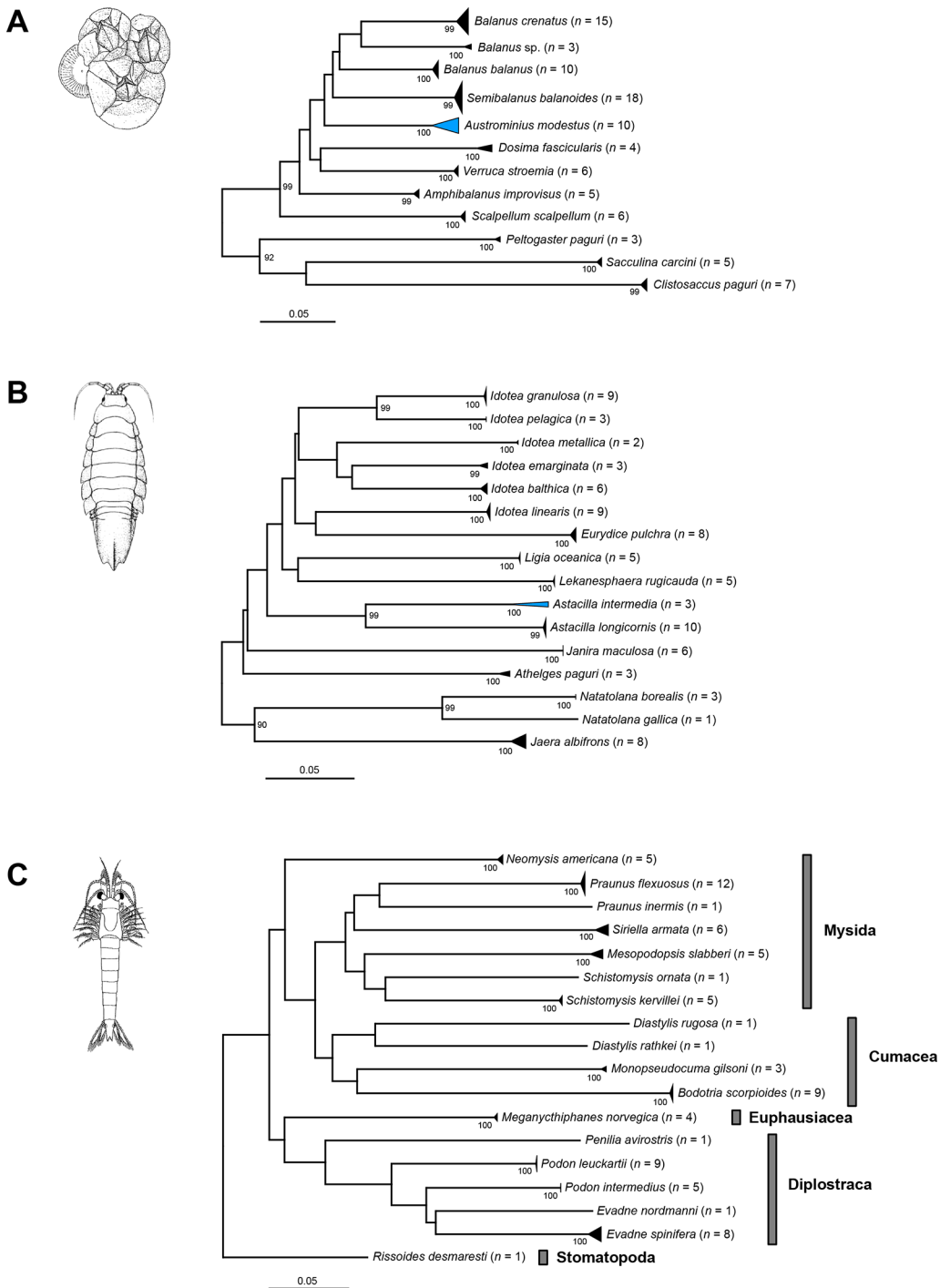


Fig 4. Neighbor joining topologies of various crustacean taxa based on Kimura 2-parameter distances. The number of analyzed specimens collapsed into a single node is provided following the species name. Triangles indicate the relative number of individual's sampled (height) and sequence divergence (width). Blue triangles indicate species with intraspecific maximum pairwise distances >2.2%. Numbers next to nodes represent non-parametric bootstrap values >90% (1,000 replicates). Drawings of species are taken and modified from previous publications (*Balanus crenatus* Bruguère, 1789 (A) and *Praunus inermis* (Rathke, 1843) (C): [79]; *Idotea balthica* (Pallas, 1772) (B): [80]).

doi:10.1371/journal.pone.0139421.g004

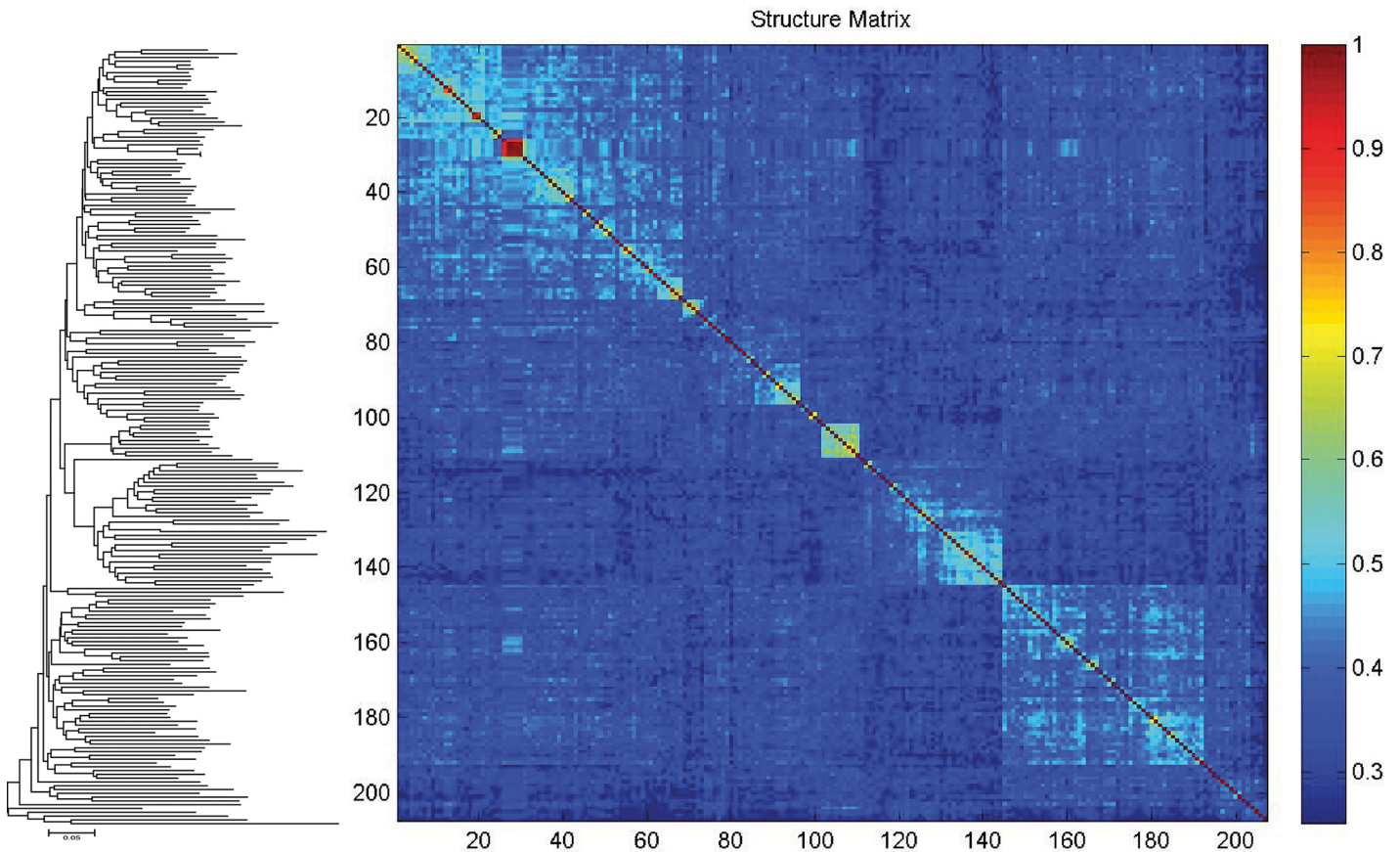


Fig 5. Klee diagram of the analyzed crustacean species. The image was generated from a TreeParser-ordered alignment with a correlation scale presented at the right of the diagram. Sequence clusters appear as blocks of higher correlation along the diagonal, on the left a corresponding NJ topology in identical order (see [S3 Fig](#) for details).

doi:10.1371/journal.pone.0139421.g005

haplotypes were identified for both species. Two haplotypes were shared by specimens of both species (h1 and h2), whereas the remaining five haplotypes were only scored in one specimen (singletons), with one haplotype found for *Macropodia parva* and four for *Macropodia rostrata*. K2P distances ranged from 0.48 to 1.12%. The nearest neighbor species was *Macropodia tenuirostris* (Leach, 1814) ($n = 20$, 7 haplotypes) with one dominant haplotype h1 ($n = 14$), separated by more than 20 additional mutational steps from the *Macropodia parva/rostrata* cluster. The minimum distance value between the *Macropodia parva/rostrata* cluster and *Macropodia tenuirostris* was 4.32%.

Using SPIDER, we obtained 1,292 correct and 40 incorrect identifications for the BM approach ([Table 3](#)). In total, 26 identifications were associated to singletons without conspecific sequences to match. After removing these singletons, incorrect identifications were reduced to 14 (*Macropodia parva* and *Macropodia rostrata*). Details of sequence comparisons are available as supporting information ([S2 Table](#)). Using a threshold of 1% as applied in the BOLD ID engine, for the BCM and ABB approaches a value of 2.1% was found for the threshold optimization method whereas a value of 3.7% was proposed by the local minima approach, respectively. For the BCM approach, correct identifications ranged from 1,249 (1% threshold) to 1,288 (3.7% threshold). Sequences with no ID ranged from 68 (1% threshold) to 29 (3.7% threshold). Incorrect and ambiguous identifications were one and 14 with all threshold values, respectively. The exclusion of singletons exclusively influenced the identifications without ID,

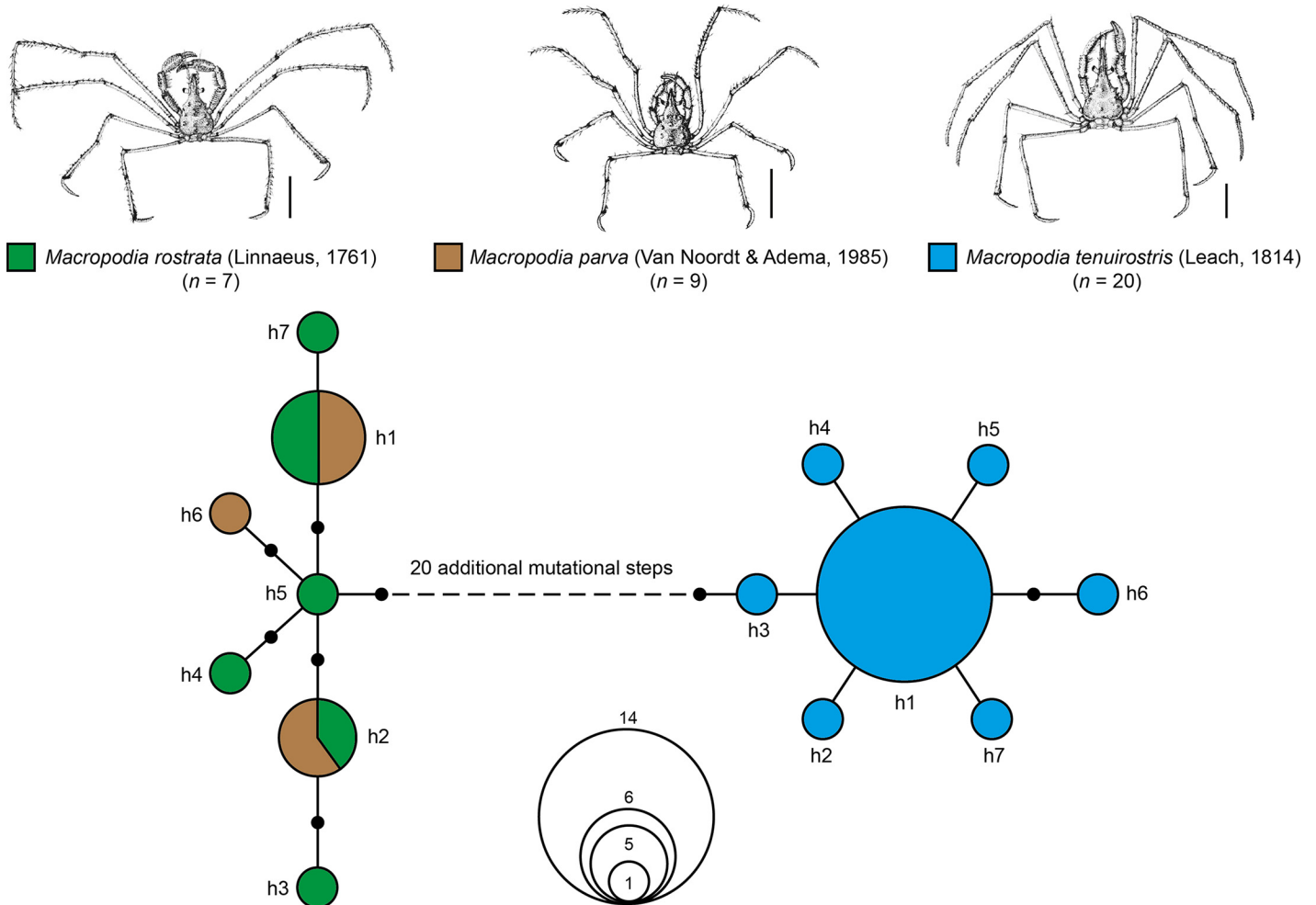


Fig 6. Maximum statistical parsimony network of the three analyzed *Macropodia* species. Settings included a user specified maximum of connection steps at 25 and gaps treated as fifth state. Each line in the network represents a single mutational change; small black dots indicate missing haplotypes. The numbers of analyzed specimens (*n*) are listed, while the diameter of the circles is proportional to the number of haplotypes sampled (see given Open circles with numbers). Scale bars = 1 cm. Illustrations were taken and modified from a previous publication [81].

doi:10.1371/journal.pone.0139421.g006

reducing them to 42 (1% threshold), six (2.1% threshold), and three (3.7% threshold). For the ASB approach, correct identifications ranged from 1,248 (1% and 2.1% thresholds) to 1,266 (3.7% threshold), ambiguous identifications ranged from 16 (1% and 2.1% thresholds) to 37 (3.7% threshold), and identification without ID had values from 68 (1% threshold) to 29 (3.7% threshold). After removing the singletons, only the identifications without ID were reduced to 42 (1% threshold), six (2.1% threshold), and three (3.7% threshold).

Discussion

Our sequence library represents an important step towards the application of DNA barcodes for the identification of crustacean taxa in the North Sea. For 86 taxa (42%), our sequence data represent the first published DNA barcodes. In total, unique BINs were found for 198 (96.6%) species, indicating a high coverage of unique BINs and analyzed species. Shared haplotypes were only found for two decapod species, whereas high intraspecific distances were only documented in one copepod species. This high efficiency of specimen identification was corroborated by our

Table 3. Results of the identification simulations using Best Match (BM), Best Close Match (BCM) and All Species Barcode (ASB) criteria based on SPIDER. Correct and incorrect identifications indicate positive and negative outcome for the respective test. An “ambiguous” outcome corresponds to the presence of both correct and incorrect identifications within the threshold (ASB) or more than one equally close match with different identification including the correct one (BCM). A “no ID” outcome corresponds to no matches found within the threshold (both BCM and ASB). Values in brackets represent the results of simulation with the exclusion of singletons.

	BM	BCM 1%	BCM 2.1%	BCM 3.7%	ASB 1%	ASB 2.1%	ASB 3.7%
Correct	1292 (1295)	1249 (1249)	1285 (1285)	1288 (1288)	1248 (1248)	1248 (1248)	1266 (1266)
Incorrect	40 (14)	1 (1)	1 (1)	1 (1)	- (-)	- (-)	- (-)
Ambiguous	-	14 (14)	14 (14)	14 (14)	16 (16)	16 (16)	37 (37)
No ID	-	68 (42)	32 (6)	29 (3)	68 (42)	32 (6)	29 (3)

doi:10.1371/journal.pone.0139421.t003

simulations using SPIDER, which were based on three criteria each with a different level of tolerance. The BM approach provided the highest number of correct identifications, i.e. the largest congruence between sequence-based identifications and prior taxonomic assignments based on morphology. However, this approach has an intrinsic bias, for it only considers the closest match regardless of distance. In fact, even singletons received a taxonomic assignment, and the amount of discordance with prior morphological identification was the highest (Table 3). Therefore, this method cannot be recommended as long as the reference database is not complete, as it is currently the case for most marine invertebrates. By using a distance threshold (BCM and ASB) it was possible to highlight cases of low interspecific distances (“ambiguous” entries in Table 3) and to exclude matches with high distances (“no ID” entries in Table 3). Of all used approaches, the ASB is the strictest one, providing no identification if query sequence matches were found below the proposed threshold. This criterion highlights specimens requiring further investigation, either for potential cases of cryptic diversity or misidentifications. We like to point out that the approaches used in our simulation are not to be confused with “species delimitation” [82]. Certainly, some concepts applied here overlap with those used for species delimitation, but our simulations exclusively tested the performance of a molecular dataset for identifying represented species. We believe that for species delimitation, and thus discovery of putative cryptic species, a different sampling design should be applied covering specimens from various locations of the entire known distribution of a species.

Whereas the species numbers of many terrestrial taxa in Europe are well known, e.g. the diurnal butterflies of Europe [83], ground beetles of Germany [84], or grasshoppers of East Austria [85], no comprehensive and reliable information about the total number of crustaceans or at least specific groups of the North Sea are available. For many terrestrial taxa, e.g. birds, reptiles and various insects, a large number of active amateurs continuously help updating distribution maps and check lists. Unfortunately, this is not the case for marine crustaceans of the North Sea, and until now only very rough estimates exist for many taxa. Most available lists rely on taxon-specific publications [86,87] or national red lists that focus on specific areas of the North Sea [88]. As consequence we are unable to provide valid estimates of species coverage for the crustacean taxa analyzed in our study. In the following we will discuss our results of the analyzed species of the Thecostraca, Copepoda, Decapoda, Amphipoda and Isopoda in detail.

Maxillopoda: Thecostraca

The Thecostraca are a large assemblage of diverse crustaceans in which parasitism and the adaptation to unusual habitats resulted in a wide range of unusual morphologies and life styles [89]. Within the Thecostraca, barnacles are the best known species that typically can be found in the rocky intertidal with high abundance and which pose severe problems as biofouling organisms that settle and accumulate on wetted submerged, man-made surfaces [90]. As a consequence of

the growing global cargo ship traffic [91,92], numerous species have become known as invasive species, e.g. *Austrominius modestus* (Darwin, 1854) which invaded the North Sea in the 1940s from Australia and/or New Zealand [93,94]. While nine barnacle species are recorded for the German sectors of the North and Baltic Sea [95], the total number of thecostracan species inhabiting the North Sea is unknown. In our study we generated 92 barcodes of 12 species, including one currently unknown barnacle species (*Balanus* sp.) and three parasitic taxa (*Clistosaccus paguri* Lilljeborg, 1861, *Peltogaster paguri* Rathke, 1842, and *Sacculina carcini* Thompson, 1836). All analyzed species form distinct sequence clusters, supported by high bootstrap values (Fig 4A). With one exception (*Austrominius modestus*), all analyzed species possess unique BINs. For this taxon we found a somewhat higher genetic variability among the five studied specimens (maximum pairwise K2P distance: 3.81%) as well as two BINs (Table 2). However, the analyzed specimens form a monophyletic lineage with high bootstrap support (100%). Additional specimens from different localities would be useful to analyze these results more in detail. Nevertheless, our results clearly show that the application of DNA barcoding is highly successful for the identification of thecostracan specimens of the North Sea.

Maxillopoda: Copepoda

With more than 15,000 described species to date [45], copepods are the dominant component of the holozooplankton, both numerically and in terms of biomass [96], and represent an essential element of aquatic food chains [97,98]. The number of calanoid copepods of the North Sea ranges between 15 and 25 species [99], and about 50 parasitic species of fish are known [87,100]. In contrast to this, the total number of harpacticoid copepods is still unclear. A synoptic meiobenthic survey of 171 stations in the North Sea, ranging from the Straits of Dover in the South to the 100 m isobath in the North, revealed 278 copepod species, with more than 40% of them being new to science [86]. Ten coastal harpacticoid copepod species were included in our study, representing a very first step to analyze this taxonomically difficult but vast group of tiny crustaceans using DNA barcodes. Similar to all other analyzed copepod species the harpacticoids grouped unambiguously with high bootstrap support (Fig 1). This efficacy of DNA barcoding for the identification of copepod species has been already demonstrated in various studies for other regions [40,43,101]. Nevertheless, we found five species of copepods with maximum pairwise distances >2.2%, including four species of the Calanoida (*Temora longicornis* (Müller O.F., 1785) with 2.76%, *Calanus helgolandicus* (Claus, 1863) with 2.99%, *Pseudocalanus elongatus* (Boeck, 1865) with 4.43%, and *Anomalocera pater-soni* Templeton, 1837 with 6.11%) and one species of the Siphonostomatoida (*Caligus elongatus* Nordmann, 1832 with 14.87%) (Table 2). We found no distinct lineages within the four analyzed calanoid species, but our data revealed two distinct monophyletic clusters within the parasitic copepod *Caligus elongatus* with distances ranging from 6.9 to 14.87% (Table 2). These results are concordant with previous studies which also found two different genotypes within this sea louse species [102–104]. As part of these studies, the molecular analyses two mitochondrial genes (16S rDNA, CO1) and a number of selected morphological characters gave evidence of the presence of two sibling species [104]. Unfortunately, a final taxonomic revision of this abundant taxon is still missing.

Malacostraca: Peracarida: Amphipoda

Aside from the Isopoda and Tanaidacea, the ecologically diverse order of the Amphipoda is one of the most species-rich groups within the Peracarida with more than 170 extant families and approx. 10,000 described species so far [45]. For the German sectors of the North and Baltic Sea, 186 amphipod species are recorded [88]. As part of our study we analyzed 305

specimens representing 59 species. All species formed monophyletic clusters with high bootstrap support (Fig 2). Nevertheless, maximum pairwise distances with values higher than 2.2% were revealed for three species: *Photis longicaudata* (Bate & Westwood, 1862) with 2.36% and one BIN, *Monocorophium insidiosum* (Crawford, 1937) with 3.41% and two BINs, and *Gammarus salinus* Spooner, 1947 with 4.14% and three BINs (Table 2). Due to the fact that the taxonomic status of all three species is uncontested, we assume that the observed variability may result from phylogeographic processes that have also been documented for other amphipod species inhabiting the North and Baltic Sea [105–108].

Malacostraca: Peracarida: Isopoda

The Isopoda comprise about 10,500 described marine, freshwater and terrestrial species [45]. About 28 species are documented for the German sectors of the North and Baltic Sea [88]. Similar to almost all other analyzed crustacean taxa our barcode analysis revealed coherent monophyletic clusters with high bootstrap support for all analyzed 16 species (Fig 4B). We found maximum pairwise distances higher than 2.2% only within one species, namely *Astacilla intermedia* (Goodsir, 1841) with 4.79% and two BINs (Table 2). For this species, two of the three analyzed female specimens showed identical barcode sequences. Given the fact that all three specimens are morphologically highly similar and valid identifications are solely based on male characteristics, we are currently unable to ascertain if the observed genetic distances simply represent a high level of intraspecific variation or reflect cryptic diversity. To answer this question, more specimens need to be collected and analyzed, using both morphological characters and nuclear sequence data [30,109].

Malacostraca: Eucarida: Decapoda

With approx. 15,000 described species [45], the Decapoda represent one of the best-known taxa of the Crustacea. Decapod crustaceans are familiar to most people and represent a dominant group of benthic invertebrates of the continental shelf and slope, including many species of economic importance [42]. Thus, it is no surprise that many barcoding studies in the past focused on decapods [29,31,39–41,110,111]. Despite of their economic and ecological importance, the total number of decapod species that inhabit the North Sea is still unclear. For the German sectors of the North and Baltic Sea, 76 species of decapod crustaceans are documented [88]. As part of this study, 529 specimens of 66 decapod species were analyzed. With one exception (see below), all analyzed species can be characterized as monophyletic clusters with high bootstrap support (99–100%) (Fig 3). Within the analyzed taxa, three species showed intraspecific maximum pairwise distances greater than 2.2%: *Pagurus pubescens* Krøyer, 1838 (2.89%, one BIN), *Pandalus montagui* Leach, 1814 (4%, one BIN), and *Eriocheir sinensis* Milne Edwards, 1853 (4.78%, two BINs) (Table 2). Whereas the observed molecular variability of *Pagurus pubescens* and *Pandalus montagui* may result from phylogeographic effects, a previous barcoding study already highlighted problems in species identification within the crab genus *Eriocheir* De Haan, 1835 as a consequence of unresolved taxonomy [40]. In our case, the observed high distances for the notorious invasive Chinese mitten crab *Eriocheir sinensis* were caused by one specimen that was sampled downstream of the river Rhine close to Bonn (approx. 350 km from the river mouth). All other four specimens were collected at the coast, showing distance values ranging from zero to 0.5%. Interestingly, all specimens could be identified by morphological characters without any difficulties. It is obvious that a comprehensive taxonomic revision of this important genus using both morphological and molecular data is urgently needed.

Our data also revealed haplotype sharing for *Macropodia parva* van Noort and Adema, 1985 and *Macropodia rostrata* (Linnaeus, 1761) (Fig 6). Morphological differences between both

species of the family Inachidae are generally very subtle, plastic and difficult to spot, e.g. the presence and size of spines on the fifth pereopod, the curvature of the dactylus of the fifth pereopod and the length of the rostrum [81]. For the analyzed barcode fragment, both species shared identical haplotypes, and there was no evidence for any differentiation between them. Nevertheless, when species pairs have very recent origins or hybridize, the utility of mtDNA sequences for species identification is very limited [112,113]. After the initial “split”, the new sister species will share alleles and mutations especially in slowly evolving genes [31,112,113], and faster evolving nuclear markers as SNPs or RAD tags may be more useful for species delineation [114–117]. Based on our data, however, it is also possible that the species pair represents one species and *Macropodia parva* needs to be synonymized with *Macropodia rostrata*. Additional analyses of morphological as well as molecular data have to be performed to answer this question.

Other analyzed crustacean taxa

In addition to the already discussed data, various other crustacean taxa were analyzed, including five species of the Diplostraca, seven species of the Mysida, four species of the Cumacea, one species of the Euphausiacea and one species of the Stomatopoda. For all these taxa, the total number of species occurring in the North Sea is unknown. All analyzed species, however, formed cohesive clusters with high support, each correlating with a single BIN (Fig 4C). For all taxa, maximum pairwise distances were lower than 2.2%.

Conclusions

Our data represent the first important step towards the establishment of a comprehensive DNA barcode library of the Crustacea of the North Sea. Despite the fact that various taxa are still missing (e.g. Tanaidacea or Ostracoda) or are currently underrepresented (e.g. harpacticoid copepods), our results clearly underline the usefulness of DNA barcodes to discriminate the vast majority of the analyzed species. It should be also kept in mind that the benefits of DNA barcoding are not restricted to taxonomic or systematic research only. The rise of modern high-throughput sequencing technologies will change biomonitoring applications and surveys in the coming years significantly [23,118,119]. As consequence, reference datasets such as ours will become essential for the correct identification of specimens sequenced as part of metabarcoding studies. This is especially true for the North Sea, a marine region that has been massively affected by cargo ship traffic, the exploitation of oil and gas resources, the rise of offshore wind parks and in particular extensive long-term fisheries.

Supporting Information

S1 Fig. Frequency histogram of the number of barcodes per species. Twenty six were represented by one barcode (12.7%), whereas 129 species (61.4%) had five or more DNA barcodes. (TIF)

S2 Fig. Neighbor joining topology of all analyzed crustacean specimens based on Kimura 2-parameter distances. Specimens are classified using ID numbers from BOLD and species name. Numbers next to nodes represent non-parametric bootstrap values (1,000 replicates, in %). (PDF)

S3 Fig. Neighbor joining topology of all analyzed crustacean species based on Kimura 2-parameter distances used for the Klee diagram. (PDF)

S1 Table. Molecular distances based on the Kimura 2-parameter model of the analyzed specimens of the Crustacea. Divergence values were calculated for all studied sequences, using the Nearest Neighbor Summary implemented in the Barcode Gap Analysis tool provided by the Barcode of Life Data System (BOLD). Align sequencing option: BOLD aligner (amino acid based HMM), ambiguous base/gap handling: pairwise deletion. ISD = intraspecific distance. BINs are based on the barcode analysis from 02-06-2015. AphiaID codes were retrieved from the World Register of Marine Species (WoRMS; www.marinespecies.org) on 02-06-2015. Asterisks indicate new species to BOLD.
(DOCX)

S2 Table. Results of the SPIDER simulations of sequence-based specimen identification. Query: Each sequence (excluded singletons) used as a query against the complete crustacean COI library (including singletons). Best Match: Results of query identification using the Best Match criterion. BCM: results of identifications based on Best Close Match criterion and three different thresholds (1%, 2.1% and 3.7%). Results were: "correct" or "incorrect" depending on the comparison with our prior morphological identification; "ambiguous" when the simulations returned more sequences from different species with the same distance from the query and below the given threshold; and "no ID" when no matches were found below the given threshold. ASB: results of identifications based on All Species Barcodes and three different thresholds (1%, 2.1% and 3.7%).
(XLSX)

Acknowledgments

The following colleagues are thanked for either collecting, providing, sorting and/or identifying specimens (in alphabetical order): Holger Bäcker, Carola Becker, Dirk Brandis, Angelika Brandt, Andreas Dürr, Christiane Eichner, Madeline Gerlach, Dominik Gloe, Lars Gutow, Carina Harms, Inga Hellige, Heike Kück, Alexandra Markert, Pedro Martínez Arbizu, Karin Meisner, Laura Ney, Marthe Otto, Stefan Richter, Imke Rottgardt, Franziska Schade, Isabel Schmalenbach, Katharina Schmidt, Myriam Schüller, Sabine Schüchel, Ulrike Schüchel, Meike Seefeldt, Moritz Sonnwald, Max Theisen, Anja Thomann, and Michael Türkay. We are grateful to Valeska Borges, Corinna Girschik, and Rebekka Schüller for their help in the molecular lab at the DZMB. Finally, we are thankful to all crew members of RV Walter Herwig III, FK Senckenberg and all other vessels for their professional help, support and advice as well as the BOLD team in managing the barcode data.

Author Contributions

Conceived and designed the experiments: MJR SL IM TCK. Performed the experiments: MJR SL IM KP CW. Analyzed the data: MJR AB DS TCK. Contributed reagents/materials/analysis tools: MJR AB DS JB SL IM HN TCK KP AR AS-V CW TK. Wrote the paper: MJR AB DS JB SL IM HN TCK KP AR AS-V TK.

References

1. Ortea I, Barros L, Gallardo JM. Closely related shrimp species identification by MALDI-ToF mass spectrometry. *J Aquat Food Prod T* 2009; 18: 146–155. doi: [10.1080/10498850802584280](https://doi.org/10.1080/10498850802584280)
2. Riccardi N, Lucini L, Benagli C, Welker M, Wicht B, Tonolla M. Potential of matrix-assisted laser desorption/ionization time-of-flight mass spectrometry (MALDI-TOF MS) for the identification of freshwater zooplankton: a pilot study with three Eudiaptomus (Copepoda: Diaptomidae) species. *J Plankton Res* 2012; 34: 484–492. doi: [10.1093/plankt/fbs022](https://doi.org/10.1093/plankt/fbs022)

3. Laakmann S, Gerds G, Erler R, Knebelberger T, Martínez Arbizu P, Raupach MJ. Comparison of molecular species identification for North Sea calanoid copepods (Crustacea) using proteome fingerprints and DNA sequences. *Mol Ecol Resour* 2013; 13: 862–876. PMID: [23848968](#)
4. Rodríguez-Fernández JI, de Carvalho CJB, Pasquini C, de Lima KMG, Moura MO, Arizaga GGC. Barcoding without DNA? Species identification using near infrared spectroscopy. *Zootaxa* 2011; 2933: 46–54.
5. Fischnaller S, Dowell FE, Lusser A, Schlick-Steiner BC, Steiner FM. Non-destructive species identification of *Drosophila obscura* and *Drosophila subobscura* (Diptera) using near-infrared spectroscopy. *Fly* 2012; 6: 284–289. doi: [10.4161/fly.21535](#) PMID: [22885252](#)
6. Ballard JWO, Whitlock MC. The incomplete natural history of mitochondria. *Mol Ecol* 2004; 13: 729–744. doi: [10.1046/j.1365-294X.2003.02063.x](#) PMID: [15012752](#)
7. Ballard JWO, Rand DM. The population biology of mitochondrial DNA and its phylogenetic implications. *Annu Rev Ecol Evol S* 2005; 36: 621–642. doi: [10.1146/annurev.ecolsys.36.091704.175513](#)
8. Bernt M, Braband A, Schierwater B, Stadler PF. Genetic aspects of mitochondrial genome evolution. *Mol Phylogenet Evol* 2013; 69: 328–338. doi: [10.1016/j.ympev.2012.10.020](#) PMID: [23142697](#)
9. Birky CW Jr, Maruyama T, Fuerst PA. An approach to population and evolutionary genetic theory for genes in mitochondria and chloroplasts, and some results. *Genetics* 1983; 103: 513–527. PMID: [6840539](#)
10. Hebert PDN, Ratnasingham S, deWaard JR. Barcoding animal life: cytochrome c oxidase subunit 1 divergences among closely related species. *Proc Biol Sci* 2003; 270: S96–S99. doi: [10.1098/rsbl.2003.0025](#) PMID: [12952648](#)
11. Hebert PDN, Cywinska A, Ball SL, deWaard JR. Biological identifications through DNA barcodes. *Proc Biol Sci* 2003; 270: 313–321. doi: [10.1098/rspb.2002.2218](#) PMID: [12614582](#)
12. Bucklin A, Steinke D, Blanco-Bercial L. DNA barcoding of marine Metazoa. *Ann Rev Mar Sci* 2011; 3: 471–508. doi: [10.1146/annurev-marine-120308-080950](#) PMID: [21329214](#)
13. Hebert PDN, Penton EH, Burns JM, Janzen DH, Hallwachs W. Ten species in one: DNA barcoding reveals cryptic species in the neotropical skipper butterfly *Astrapes fulgerator*. *Proc Natl Acad Sci U S A* 2004; 101: 14812–14817. doi: [10.1073/pnas.0406166101](#)
14. Doublet V, Souty-Grosset C, Bouchon D, Cordaux R, Marcade I. A thirty million year-old inherited heteroplasmy. *PLoS One* 2008; 3: e2938. doi: [10.1371/journal.pone.0002938](#) PMID: [18698356](#)
15. Doublet V, Helleu Q, Raimond R, Souty-Grosset C, Marcadé I. Inverted repeats and genome architecture conversions of terrestrial isopods mitochondrial DNA. *J Mol Evol* 2013; 77: 107–118. doi: [10.1007/s00239-013-9587-7](#) PMID: [24068302](#)
16. Baeza JA, Fuentes MS. Phylogeography of the shrimp *Palaemon floridanus* (Crustacea: Caridea: Palaemonidae): a partial test of meta-population genetic structure in the wider Caribbean. *Mar Ecol* 2013; 34: 381–393. doi: [10.1111/maec.12038](#)
17. Buhay JE. “COI-like” sequences are becoming problematic in molecular systematic and DNA barcoding studies. *J Crustacean Biol* 2009; 29: 96–110. doi: [10.1651/08-3020.1](#)
18. Song H, Buhay JE, Whiting MF, Crandall KA. Many species in one: DNA barcoding overestimates the number of species when nuclear mitochondrial pseudogenes are coamplified. *Proc Natl Acad Sci U S A* 2008; 105: 13486–13491. doi: [10.1073/pnas.0803076105](#) PMID: [18757756](#)
19. Taylor DJ, Hebert PDN. Cryptic intercontinental hybridization in *Daphnia* (Crustacea): the ghost of introductions past. *Proc Biol Sci* 1993; 254: 163–168. doi: [10.1098/rspb.1993.0141](#)
20. Barber BR, Xu J, Pérez-Losada M, Jara CG, Crandall KA. Conflicting evolutionary patterns due to mitochondrial introgression and multilocus phylogeography of the Patagonian freshwater crab *Aegla neuquensis*. *PLoS One* 2012; 7: e37105. doi: [10.1371/journal.pone.0037105](#) PMID: [22685541](#)
21. Barber PH, Boyce SL. Estimating diversity of Indo-Pacific coral reef stomatopods through DNA barcoding of stomatopod larvae. *Proc Biol Sci* 2006; 273: 2053–2061. doi: [10.1098/rspb.2006.3540](#) PMID: [16846913](#)
22. Radulovici AE, Archambault P, Dufresne F. DNA barcodes for marine biodiversity: moving fast forward? *Diversity* 2010; 2: 450–472. doi: [10.3390/d2040450](#)
23. Leray M, Knowlton N. DNA barcoding and metabarcoding of standardized samples reveal patterns of marine benthic diversity. *Proc Natl Acad Sci U S A* 2015; 112: 2076–2081. doi: [10.1073/pnas.1424997112](#) PMID: [25646458](#)
24. Witt JDS, Threlloff DL, Hebert PDN. DNA barcoding reveals extraordinary cryptic diversity in an amphipod genus: implications for desert spring conservation. *Mol Ecol* 2007; 15: 3073–3082. doi: [10.1111/j.1365-294X.2006.02999.x](#) PMID: [16911222](#)

25. Haye PA, Segovia NI, Vera R, Gallardo MdIA, Gallardo-Escárate C. Authentication of commercialized crab-meat in Chile using DNA barcoding. *Food Control* 2012; 25: 239–244. doi: [10.1016/j.foodcont.2011.10.034](https://doi.org/10.1016/j.foodcont.2011.10.034)
26. Nicolé S, Negrisoló E, Eccher G, Mantovani R, Patarnello T, Erickson DL, et al. DNA barcoding as reliable method for the authentication of commercial seafood products. *Food Technol Biotech* 2012; 50: 387–398.
27. Chen Y-Y, Lin H-C, Chan BKK. Description of a new species of coral-inhabiting barnacle, *Darwiniella angularis* sp. n. (Cirripedia, Pyrgomatidae) from Taiwan. *ZooKeys* 2012; 214: 43–74. doi: [10.3897/zookeys.214.3291](https://doi.org/10.3897/zookeys.214.3291) PMCID: PMC3426880 PMID: [22936866](https://pubmed.ncbi.nlm.nih.gov/22936866/)
28. Riehl T, Kaiser S. Conquered from the deep sea? A new deep-sea isopod species from the Antarctic shelf shows pattern of recent colonization. *PLoS One* 2012; 7: e49354. doi: [10.1371/journal.pone.0049354](https://doi.org/10.1371/journal.pone.0049354) PMCID: PMC3492298 PMID: [23145160](https://pubmed.ncbi.nlm.nih.gov/23145160/)
29. De Grave S, Chu KH, Chan T-Y. On the systematic position of *Galatheacaris abyssalis* (Decapoda: Galatheacaridoidea). *J Crustacean Biol* 2013; 30: 521–527. doi: [10.1651/10-3278.1](https://doi.org/10.1651/10-3278.1)
30. Khalaji-Pirbalouty V, Raupach MJ. A new species of *Cymodoce* Leach, 1814 (Crustacea: Isopoda: Sphaeromatidae) based on morphological and molecular data, with a key to the Northern Indian Ocean species. *Zootaxa* 2014; 3826: 230–254. doi: [10.11646/zootaxa.3826.1.7](https://doi.org/10.11646/zootaxa.3826.1.7) PMID: [24990044](https://pubmed.ncbi.nlm.nih.gov/24990044/)
31. Markert A, Raupach MJ, Segelken-Voigt A, Wehrmann A. Molecular identification and morphological characteristics of native and invasive Asian brush-clawed crabs (Crustacea: Brachyura) from Japanese and German coasts: *Hemigrapsus penicillatus* (De Haan, 1835) versus *Hemigrapsus takanoi* Asakura & Watanabe 2005. *Org Divers Evol* 2014; 14: 369–382. doi: [10.1007/s13127-014-0176-4](https://doi.org/10.1007/s13127-014-0176-4)
32. Zhou X, Robinson JL, Geraci CJ, Parker CR, Flint OS Jr, Etnier DA, et al. Accelerated construction of a regional DNA-barcode reference library: caddisflies (Trichoptera) in the Great Smoky Mountains National Park. *J N Am Benthol Soc* 2011; 30: 131–162. doi: [10.1899/10-010.1](https://doi.org/10.1899/10-010.1)
33. Hausmann A, Haszprunar G, Hebert PDN. DNA barcoding the Geometrid fauna of Bavaria (Lepidoptera): successes, surprises, and questions. *PLoS One* 2011; 6: e17134. doi: [10.1371/journal.pone.0017134](https://doi.org/10.1371/journal.pone.0017134) PMID: [21423340](https://pubmed.ncbi.nlm.nih.gov/21423340/)
34. Woodcock TS, Boyle EE, Roughley RE, Kevan PG, Labbee RN, Smith AB, et al. The diversity and biogeography of the Coleoptera of Churchill: insights from DNA barcoding. *BMC Ecology* 2013; 13: 40. doi: [10.1186/1472-6785-13-40](https://doi.org/10.1186/1472-6785-13-40) PMID: [24164967](https://pubmed.ncbi.nlm.nih.gov/24164967/)
35. Morinière J, Hendrich L, Hausmann A, Hebert PDN, Haszprunar G, Gruppe A. Barcoding Fauna Bavarica: 78% of the Neuropterida fauna barcoded! *PLoS One* 2014; 9: e109719. doi: [10.1371/journal.pone.0109719](https://doi.org/10.1371/journal.pone.0109719) PMID: [25286434](https://pubmed.ncbi.nlm.nih.gov/25286434/)
36. Raupach MJ, Hendrich L, Küchler SM, Deister F, Morinière J, Gossner MM. Building-up of a DNA barcode library for true bugs (Insecta: Hemiptera: Heteroptera) of Germany reveals taxonomic uncertainties and surprises. *PLoS One* 2014; 9: e106940. doi: [10.1371/journal.pone.0106940](https://doi.org/10.1371/journal.pone.0106940) PMCID: PMC4159288 PMID: [25203616](https://pubmed.ncbi.nlm.nih.gov/25203616/)
37. Hendrich L, Morinière J, Haszprunar G, Hebert PDN, Hausmann A, Köhler F, et al. A comprehensive DNA barcode database for Central European beetles with a focus on Germany: adding more than 3500 identified species to BOLD. *Mol Ecol Resour* 2015; 15: 795–818. doi: [10.1111/1755-0998.12354](https://doi.org/10.1111/1755-0998.12354) PMID: [25469559](https://pubmed.ncbi.nlm.nih.gov/25469559/)
38. Schmidt S, Schmid-Egger C, Morinière J, Haszprunar G, Hebert PDN. DNA barcoding largely supports 250 years of classical taxonomy: identifications for Central European bees (Hymenoptera, Apoidea partim). *Mol Ecol Resour* 2015; 15: 985–1000. doi: [10.1111/1755-0998.12363](https://doi.org/10.1111/1755-0998.12363) PMID: [25588628](https://pubmed.ncbi.nlm.nih.gov/25588628/)
39. Lefébure T, Douady CJ, Gouy M, Gibert J. Relationships between morphological taxonomy and molecular divergence within Crustacea: proposal of a molecular threshold to help species delimitation. *Mol Phylogenet Evol* 2006; 40: 435–447. doi: [10.1016/j.ympev.2006.03.014](https://doi.org/10.1016/j.ympev.2006.03.014) PMID: [16647275](https://pubmed.ncbi.nlm.nih.gov/16647275/)
40. Costa FO, deWaard JR, Boutillier J, Ratnasingham S, Dooh RT, Hajibabaei M, et al. Biological identifications through DNA barcodes: the case of the Crustacea. *Can J Fish Aquat Sci* 2007; 64: 272–295. doi: [10.1139/f07-008](https://doi.org/10.1139/f07-008)
41. Radulovici AE, Sainte-Marie B, Dufresne F. DNA barcoding of marine crustaceans from the estuary and Gulf of St Lawrence: a regional-scale approach. *Mol Ecol Resour* 2009; 9: 181–187. doi: [10.1111/j.1755-0998.2009.02643.x](https://doi.org/10.1111/j.1755-0998.2009.02643.x) PMID: [21564977](https://pubmed.ncbi.nlm.nih.gov/21564977/)
42. Matzen da Silva J, Creer S, dos Santos A, Costa AC, Cunha MR, Costa FO, et al. Systematic and evolutionary insight derived from mtDNA COI barcode diversity in the Decapoda (Crustacea: Malacostraca). *PLoS One* 2011; 6: e19449. doi: [10.1371/journal.pone.0019449](https://doi.org/10.1371/journal.pone.0019449) PMID: [21589909](https://pubmed.ncbi.nlm.nih.gov/21589909/)
43. Blanco-Bercial L, Cornils A, Copley N, Bucklin A. DNA barcoding of marine copepods: assessment of analytical approaches in species identification. *PLoS Currents Tree of Life* 2014; 6. doi: [10.1371/currents.tol.cdf8b74881f87e3b01d56b43791626d2](https://doi.org/10.1371/currents.tol.cdf8b74881f87e3b01d56b43791626d2) PMCID: PMC4073882

44. Brusca RC, Brusca GJ. *Invertebrates*. 2nd ed. Sunderland: Sinauer Associates; 2003.
45. Ah Yong ST, Lowry JK, Alonso M, Bamber RN, Boxshall GA, Castro P, et al. Subphylum Crustacea Brönnich, 1772. *Zootaxa* 2011; 3148: 165–191.
46. Appeltans W, Ah Yong ST, Anderson G, Angel MV, Artois T, Bailly N, et al. The magnitude of global marine species diversity. *Curr Biol* 2012; 22: 2189–2202. doi: [10.1016/j.cub.2012.09.036](https://doi.org/10.1016/j.cub.2012.09.036) PMID: [23159596](https://pubmed.ncbi.nlm.nih.gov/23159596/)
47. Martin JW, Davis GE. *An Updated Classification of the Recent Crustacea*. Los Angeles: Natural History Museum of Los Angeles County; 2001.
48. McClain CR, Balk MA, Benfield MC, Branch TA, Chen C, Cosgrove J, et al. Sizing ocean giants: patterns of intraspecific size variation in marine megafauna. *PeerJ* 2015; 2: e715. doi: [10.7717/peerj.715](https://doi.org/10.7717/peerj.715) PMID: [25649000](https://pubmed.ncbi.nlm.nih.gov/25649000/)
49. Dyer MF, Fry WG, Fry PD, Cranmer GJ. Benthic regions within the North Sea. *J Mar Biol Assoc UK* 1983; 63: 683–693. doi: [10.1017/S0025315400070995](https://doi.org/10.1017/S0025315400070995)
50. Reiss H, Cunze S, König K, Neumann H, Kröncke I. Species distribution modelling of marine benthos: a North Sea case study. *Mar Ecol Prog Ser* 2011; 442: 71–86. doi: [10.3354/meps09391](https://doi.org/10.3354/meps09391)
51. Heip CHR, Basford D, Craeymeersch JA, Dewarumez J-M, Dörjes J, de Wilde P, et al. Trends in biomass, density and diversity of the North Sea macrofauna. *ICES J Mar Sci* 1992; 49: 13–22. doi: [10.1093/icesjms/49.1.13](https://doi.org/10.1093/icesjms/49.1.13)
52. Callaway R, Alsvåg J, de Boois I, Cotter J, Ford A, Hinz H, et al. Diversity and community structure of epibenthic invertebrates and fish in the North Sea. *ICES J Mar Sci* 2002; 59: 1199–1214. doi: [10.1006/jmsc.2002.1288](https://doi.org/10.1006/jmsc.2002.1288)
53. Neumann H, Ehrich S, Kröncke I. Spatial variability of epifaunal communities in the North Sea in relation to sampling effort. *Helgoland Mar Res* 2008; 62: 215–225. doi: [10.1007/s10152-008-0109-8](https://doi.org/10.1007/s10152-008-0109-8)
54. Kröncke I, Reiss H, Dippner JW. Effects of cold winters and regime shifts on macrofauna communities in shallow coastal regions. *Estuar Coast Shelf S* 2013; 119: 79–90. doi: [10.1016/j.ecss.2012.12.024](https://doi.org/10.1016/j.ecss.2012.12.024)
55. Beermann J, Franke HD. A supplement to the amphipod (Crustacea) species inventory of Helgoland (German Bight, North Sea): indication of rapid recent change. *Mar Biodivers Rec* 2011; 4: 1–15. doi: [10.1017/S1755267211000388](https://doi.org/10.1017/S1755267211000388)
56. WoRMS Editorial Board. *World Register of Marine Species*. 2015. Available: <http://www.marinespecies.org>
57. Ivanova NV, de Waard JR, Hebert PDN. An inexpensive, automation-friendly protocol for recovering high-quality DNA. *Mol Ecol Notes* 2006; 6: 998–1002. doi: [10.1111/j.1471-8286.2006.01428.x](https://doi.org/10.1111/j.1471-8286.2006.01428.x)
58. de Waard JR, Ivanova NV, Hajibabaei M, Hebert PDN. Assembling DNA barcodes: analytical protocols. In: Martin C, editor. *Methods in Molecular Biology: Environmental Genetics*. Totowa: Humana Press; 2008. pp. 275–293.
59. Folmer O, Black M, Hoeh W, Lutz R, Vrijenhoek R. DNA primers for amplification of mitochondrial cytochrome c oxidase subunit I from diverse metazoan invertebrates. *Mol Mar Biol Biotechnol* 1994; 3: 294–299. PMID: [7881515](https://pubmed.ncbi.nlm.nih.gov/7881515/)
60. Geller J, Meyer C, Parker M, Hawk H. Redesign of PCR primers for mitochondrial cytochrome c oxidase subunit I for marine invertebrates and application in all-taxa biotic surveys. *Mol Ecol Resour* 2013; 13: 851–861. doi: [10.1111/1755-0998.12138](https://doi.org/10.1111/1755-0998.12138) PMID: [23848937](https://pubmed.ncbi.nlm.nih.gov/23848937/)
61. Ivanova NV, Zemlak TS, Hanner RH, Hebert PDN. Universal primer cocktails for fish DNA barcoding. *Mol Ecol Notes* 2007; 7: 544–548. doi: [10.1111/j.1471-8286.2007.01748.x](https://doi.org/10.1111/j.1471-8286.2007.01748.x)
62. Kearse M, Moir R, Wilson A, Sone-Havas S, Cheung M, Sturrock S, et al. Geneious Basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 2012; 15: 1647–1649. doi: [10.1093/bioinformatics/bts199](https://doi.org/10.1093/bioinformatics/bts199) PMID: [22543367](https://pubmed.ncbi.nlm.nih.gov/22543367/)
63. Zhang Z, Schwartz S, Wagner L, Miller W. A greedy algorithm for aligning DNA sequences. *J Comput Biol* 2000; 7: 203–214. doi: [10.1089/10665270050081478](https://doi.org/10.1089/10665270050081478) PMID: [10890397](https://pubmed.ncbi.nlm.nih.gov/10890397/)
64. Morgulis M, Coulouris G, Rayseis Y, Madden TL, Agarwala R, Schäffer AA. Database indexing for production MegaBLAST searches. *Bioinformatics* 2008; 24: 17157–1764. doi: [10.1093/bioinformatics/btn322](https://doi.org/10.1093/bioinformatics/btn322) PMID: [18567917](https://pubmed.ncbi.nlm.nih.gov/18567917/)
65. Ratnasingham S, Hebert PDN. BOLD: The Barcode of Life Data Systems. *Mol Ecol Notes* 2007; 7: 355–364. doi: [10.1111/j.1471-8286.2007.01678.x](https://doi.org/10.1111/j.1471-8286.2007.01678.x) PMID: [18784790](https://pubmed.ncbi.nlm.nih.gov/18784790/)
66. Kimura M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* 1980; 16: 111–120. PMID: [7463489](https://pubmed.ncbi.nlm.nih.gov/7463489/)
67. Ratnasingham S, Hebert PDN. A DNA-based registry for all animal species: the Barcode Index Number (BIN) system. *PLoS One* 2013; 8: e66213. doi: [10.1371/journal.pone.0066213](https://doi.org/10.1371/journal.pone.0066213) PMID: [23861743](https://pubmed.ncbi.nlm.nih.gov/23861743/)

68. Saitou N, Nei M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 1987; 4: 406–425. PMID: [3447015](#)
69. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* 2013; 30: 2725–2729. doi: [10.1093/molbev/mst197](#) PMID: [24132122](#)
70. Felsenstein J. Confidence limits on phylogenies: an approach using the bootstrap. *Evolution* 1985; 39: 783–791. doi: [10.2307/2408678](#)
71. Edgar RC. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics* 2004; 5: 113. doi: [10.1186/1471-2105-5-113](#) PMID: [15318951](#)
72. Stoeckle MY, Coffran C. TreeParser-aided Klee diagrams display taxonomic clusters in DNA barcode and nuclear gene datasets. *Sci Rep* 2013; 3: 2635. doi: [10.1038/srep02635](#) PMID: [24022383](#)
73. Sirovich L, Stoeckle MY, Zhang Y. A scalable method for analysis and display of DNA sequences. *PLoS One* 2009; 4: e7051. doi: [10.1371/journal.pone.0007051](#) PMID: [19798412](#)
74. Clement M, Posada D, Crandall KA. TCS: a computer program to estimate gene genealogies. *Mol Ecol* 2000; 9: 1657–1660. doi: [10.1046/j.1365-294x.2000.01020.x](#) PMID: [11050560](#)
75. Brown SDJ, Collins RA, Boyer S, Lefort M-C, Malumbres-Olarte J, Vink CJ, et al. SPIDER: An R package for the analysis of species identity and evolution, with particular reference to DNA barcoding. *Mol Ecol Resour* 2012; 12: 562–565. doi: [10.1111/j.1755-0998.2011.03108.x](#) PMID: [22243808](#)
76. Meier R, Shiyang K, Vaidya G, Ng PKL. DNA Barcoding and taxonomy in Diptera: A tale of high intra-specific variability and low identification success. *Syst Biol* 2006; 55: 715–728. doi: [10.1080/10635150600969864](#) PMID: [17060194](#)
77. Kabata Z. Parasitic Copepoda of British Fishes. London: Ray Society Publications; 1979.
78. Lincoln RJ. British Marine Amphipoda: Gammaridea. London: British Museum (Natural History); 1979.
79. Hayward PJ, Ryland JS. Handbook of the Marine Fauna of North-West Europe. Oxford: Oxford University Press; 1990.
80. Gruner H-E. Lehrbuch der Speziellen Zoologie. Kaestner Begründet von A.. Band I. Wirbellose Tiere. 4. Teil: Arthropoda (ohne Insecta). Jena: Gustav Fischer Verlag; 1993. German.
81. van Noort GJ, Adema JPHM. The genus *Macropodia* Leach, 1814 in the North Sea and adjacent waters, with the description of a new species. *Zool Med Leiden* 2011; 59: 363–379.
82. Collins RA, Cruickshank RH. The seven deadly sins of DNA barcoding. *Mol Ecol Resour* 2012; 13: 969–975. doi: [10.1111/1755-0998.12046](#) PMID: [23280099](#)
83. Kudrna O, Harpke A, Lux K, Pennerstorfer J, Schweiger O, Settele J, et al. Distribution Atlas of Butterflies in Europe. Halle: GfS; 2011.
84. Trautner J, Fritze M-A, Hannig K, Kaiser M. Distribution Atlas of Ground Beetles in Germany. Norderstedt: Books on Demand; 2014. German.
85. Zuna-Kratky T, Karner-Ranner E, Lederer E, Braun B, Berg H-M, Denner M, et al. Verbreitungsatlas der Heuschrecken und Fangschrecken Ostösterreichs. Wien: Verlag des Naturhistorischen Museum; 2009. German.
86. Huys R, Herman PMJ, Heip CHR, Soetaert K. The meiobenthos of the North Sea: density, biomass trends and distribution of copepod communities. *ICES J Mar Sci* 1992; 49: 23–44. doi: [10.1093/icesjms/49.1.23](#)
87. Koch W, Boer P, Witte JI, van der Veer HW, Thiltges DW. Inventory and comparison of abundance of parasitic copepods on fish hosts in the western Wadden Sea (North Sea) between 1968 and 2010. *J Mar Biol Assoc UK* 2014; 94: 547–555. doi: [10.1017/S0025315413001677](#)
88. Rachor E, Bönsch R, Boos K, Gosselck F, Grotjahn M, Günther CP, et al. Rote Liste und Artenliste der bodenlebenden wirbellosen Meerestiere. In: Becker N, Haupt H, Hofbauer N, Ludwig G, Nehring S, editors. Rote Liste gefährdeter Tiere, Pflanzen und Pilze Deutschlands. Band 2: Meeresorganismen. Bonn: Bundesamt für Naturschutz; 2013. pp. 81–176. German.
89. Martin J, Olesen J, Høeg JT. Atlas of Crustacean Larvae. Baltimore: Johns Hopkins University Press; 2014.
90. Holm ER. Barnacles and biofouling. *Integr Comp Biol* 2012; 52: 348–355. PMID: [22508866](#)
91. Kaluza P, Kölsch A, Gastner MT, Blasius B. The complex network of global cargo ship movement. *J R Soc Interface* 2010; 7: 1093–1103. doi: [10.1098/rsif.2009.0495](#)
92. Seebens H, Gastner MT, Blasius B. The risk of marine bioinvasion caused by global shipping. *Ecol Lett* 2013; 16: 782–790. doi: [10.1111/ele.12111](#) PMID: [23611311](#)

93. Bishop MWH. Establishment of an immigrant barnacle in British coastal waters. *Nature* 1947; 159: 501. doi: [10.1038/159501a0](https://doi.org/10.1038/159501a0)
94. Crisp DJ. The spread of *Elminius modestus* Darwin in north-west Europe. *J Mar Biol Assoc UK* 1958; 37: 483–520. doi: [10.1017/S0025315400023833](https://doi.org/10.1017/S0025315400023833)
95. Luther G. Seepocken der deutschen Küstengewässer. *Helgolander Meeresun* 1987; 41: 1–43. German.
96. Harris RP, Wiebe PH, Lenz J, Skjoldal HR, Huntley M. *ICES Zooplankton Methodology Manual*. Waltham: Academic Press; 2000.
97. Sommer U, Stibor H. Copepoda—Cladocera—Tunicata: The role of three major mesozooplankton groups in pelagic food webs. *Ecol Res* 2002; 17: 161–174. doi: [10.1046/j.1440-1703.2002.00476.x](https://doi.org/10.1046/j.1440-1703.2002.00476.x)
98. Turner JT. The importance of small planktonic copepods and their roles in pelagic marine food webs. *Zool Stud* 2004; 43: 255–266.
99. Franz HG, Colebrook JM, Gamble JC, Krause M. The zooplankton of the North Sea. *Neth J Sea Res* 1991; 28: 1–52.
100. Boxshall GA. Infections with parasitic copepods in North Sea marine fishes. *J Mar Biol Assoc UK* 1974; 54: 355–372. doi: [10.1017/S0025315400058598](https://doi.org/10.1017/S0025315400058598)
101. Bucklin A, Ortman BD, Jennings RM, Nigro LM, Sweetman CJ, Copley NJ, et al. A “Rosetta Stone” for metazoan zooplankton: DNA barcode analysis of species diversity of the Sargasso Sea (Northwest Atlantic Ocean). *Deep Sea Res PT II* 2010; 57: 2234–2247. doi: [10.1016/j.dsr2.2010.09.025](https://doi.org/10.1016/j.dsr2.2010.09.025)
102. Øines Ø, Heuch PA. Identification of sea louse species of the genus *Caligus* using mtDNA. *J Mar Biol Assoc UK* 2005; 85: 73–79. doi: [10.1017/S0025315405010854h](https://doi.org/10.1017/S0025315405010854h)
103. Øines Ø, Simonsen JH, Knutsen JA, Heuch PA. Host preference of adult *Caligus elongatus* in the laboratory and its implications for Atlantic cod aquaculture. *J Fish Dis* 2006; 29: 167–174. doi: [10.1111/j.1365-2761.2006.00702.x](https://doi.org/10.1111/j.1365-2761.2006.00702.x) PMID: [16533302](https://pubmed.ncbi.nlm.nih.gov/16533302/)
104. Øines Ø, Schram T. Intra- or inter-specific differences in genotypes of *Caligus elongatus* Nordmann 1832? *Acta Parasitol* 2008; 53: 93–105. doi: [10.2478/s11686-008-0002-2](https://doi.org/10.2478/s11686-008-0002-2)
105. Rock J, Ironside J, Potter T, Whiteley NM, Lunt DH. Phylogeography and environmental diversification of a highly adaptable marine amphipod, *Gammarus duebeni*. *Heredity* (Edinb) 2007; 99: 102–111. doi: [10.1038/sj.hdy.6800971](https://doi.org/10.1038/sj.hdy.6800971) PMID: [17426729](https://pubmed.ncbi.nlm.nih.gov/17426729/)
106. Costa FO, Henzler CM, Lunt DH, Whiteley NM, Rock J. Probing marine *Gammarus* (Amphipoda) taxonomy with DNA barcodes. *Syst Biodivers* 2009; 7: 365–379. doi: [10.1017/S1477200009990120](https://doi.org/10.1017/S1477200009990120)
107. Krebs L, Blank M, Jürss K, Zettler ML, Bastrop R. Glacial-driven vicariance in the amphipod *Gammarus duebeni*. *Mol Phylogenet Evol* 2010; 54: 372–385. doi: [10.1016/j.ympev.2009.07.034](https://doi.org/10.1016/j.ympev.2009.07.034) PMID: [19654046](https://pubmed.ncbi.nlm.nih.gov/19654046/)
108. Krebs L, Blank M, Bastrop R. Phylogeography, historical demography and postglacial colonization routes of two amphiatlantic distributed amphipods. *Syst Biodivers* 2011; 9: 259–273. doi: [10.1080/14772000.2011.604359](https://doi.org/10.1080/14772000.2011.604359)
109. Brökeland W, Raupach MJ. A species complex within the genus *Haploniscus* (Crustacea: Malacostraca: Peracarida) from the Southern Ocean deep sea: a morphological and molecular approach. *Zool J Linn Soc* 2008; 152: 655–706. doi: [10.1111/j.1096-3642.2008.00362.x](https://doi.org/10.1111/j.1096-3642.2008.00362.x)
110. Meyer R, Weis A, Melzer RR. Decapoda of southern Chile: DNA barcoding and integrative taxonomy with focus on the genera *Acanthocyclus* and *Eurypodius*. *Syst Biodivers* 2013; 11: 389–404. doi: [10.1080/14772000.2013.833143](https://doi.org/10.1080/14772000.2013.833143)
111. Negri M, Lemaitre R, Mantelatto FL. Molecular and morphological resurrection of *Clibanarius symmetricus* (Randall, 1840), a new cryptic species hiding under name for the “thin stripe” hermit crab *C. vittatus* (Bosc, 1802). *J Crustacean Biol* 2014; 34: 848–861. doi: [10.1163/1937240X-00002277](https://doi.org/10.1163/1937240X-00002277)
112. Tautz D, Arctander P, Minelli A, Thomas RH, Vogler AP. A plea for DNA taxonomy. *Trends Ecol Evol* 2003; 18: 70–74. doi: [10.1016/S0169-5347\(02\)00041-1](https://doi.org/10.1016/S0169-5347(02)00041-1)
113. Raupach MJ, Astrin JJ, Hannig K, Peters MK, Stoeckle MY, Wägele JW. Molecular species identifications of Central European ground beetles (Coleoptera: Carabidae) using nuclear rDNA expansion segments and DNA barcodes. *Front Zool* 2010; 7: 26. doi: [10.1186/1742-9994-7-26](https://doi.org/10.1186/1742-9994-7-26) PMCID: [20836845](https://pubmed.ncbi.nlm.nih.gov/20836845/)
114. Ferguson B, Street SL, Wright H, Pearson C, Jia Y, Thompson SL, et al. Single nucleotide polymorphisms (SNPs) distinguish Indian-origin and Chinese-origin rhesus macaques (*Macaca mulatta*). *BMC Genomics* 2007; 8: 43. doi: [10.1186/1471-2164-8-43](https://doi.org/10.1186/1471-2164-8-43) PMCID: [17286860](https://pubmed.ncbi.nlm.nih.gov/17286860/)
115. Nadeau NJ, Martin SH, Kozak KM, Salazar C, Dasmahapatra K, Davey JW, et al. Genome-wide patterns of divergence and gene flow across a butterfly radiation. *Mol Ecol* 2013; 22: 814–826. doi: [10.1111/j.1365-294X.2012.05730.x](https://doi.org/10.1111/j.1365-294X.2012.05730.x) PMID: [22924870](https://pubmed.ncbi.nlm.nih.gov/22924870/)

116. Leaché AD, Fujita MK, Minin VN, Bouckaert RR. Species delimitation using genome-wide SNP data. *Syst Biol* 2014; 63: 534–542. doi: [10.1093/sysbio/syu018](https://doi.org/10.1093/sysbio/syu018) PMID: [24627183](https://pubmed.ncbi.nlm.nih.gov/24627183/)
117. Pante E, Abdelkrim J, Viricel A, Gey D, France SC, Boisselier MC, et al. Use of RAD sequencing for delimiting species. *Heredity* 2015; 114: 450–459. doi: [10.1038/hdy.2014.105](https://doi.org/10.1038/hdy.2014.105) PMID: [25407078](https://pubmed.ncbi.nlm.nih.gov/25407078/)
118. Fonseca VG, Carvalho GR, Sung W, Johnson HF, Power DM, Neill SP, et al. Second-generation environmental sequencing unmasks marine metazoan biodiversity. *Nat Commun* 2010; 1: 98. doi: [10.1038/ncomms1095](https://doi.org/10.1038/ncomms1095) PMCID: PMC2963828 PMID: [20981026](https://pubmed.ncbi.nlm.nih.gov/20981026/)
119. Hajibabaei M, Shokralla S, Zhou X, Singer GAC, Baird DJ. Environmental barcoding: a next-generation sequencing approach for biomonitoring applications using river benthos. *PLoS One* 2011; 6: e17497. doi: [10.1371/journal.pone.0017497](https://doi.org/10.1371/journal.pone.0017497) PMID: [21533287](https://pubmed.ncbi.nlm.nih.gov/21533287/)